



**HAL**  
open science

## Biological modeling of mucus to modulate mucus barriers.

Jean-Luc Desseyn, Valérie Gouyer, Frederic Gottrand

► **To cite this version:**

Jean-Luc Desseyn, Valérie Gouyer, Frederic Gottrand. Biological modeling of mucus to modulate mucus barriers.. *AJP - Gastrointestinal and Liver Physiology*, 2016, American journal of physiology. Gastrointestinal and liver physiology, 310 (4), pp.G225-G227. 10.1152/ajpgi.00274.2015 . hal-02176893v1

**HAL Id: hal-02176893**

**<https://hal.univ-lille.fr/hal-02176893v1>**

Submitted on 8 Jul 2019 (v1), last revised 17 Jun 2022 (v2)

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

## Perspectives

### Biological modeling of mucus to modulate mucus barriers

Jean-Luc Desseyn, Valérie Gouyer & Frédéric Gottrand

LIRIC – UMR\_995; Inserm; Univ. Lille; CHU Lille, F-59000 Lille, France

**Corresponding author:** J-L. Desseyn (jean-luc.desseyn@inserm.fr)

**Keywords:** gelling mucin; mucus; mucosal barrier; mucin CYS domain; transgenic mouse

#### Abstract

A recent study using a transgenic mouse, whose intestinal mucus contains a molecule made of 12 copies of a domain found in many gelling mucins, demonstrates that it is possible to strengthen mucus properties *in situ*, leading to promising new treatment strategies in diseases where the mucosal barrier is impaired.

*Am J Physiol Gastrointest Liver Physiol* 2016 Feb 15;310(4):G225-7.

First published December 10, 2015; doi:10.1152/ajpgi.00274.2015.

Infections are a major cause of morbidity and mortality worldwide, particularly in children, the elderly and people with specific risk factors. Epithelial cells are the main portal of entry for many pathogens. However, before entering epithelial cells or passing between the cells into tissues, pathogens often have to cross a mucus hydrogel lying on the apical side of epithelial cells. This is particularly true in the vagina, respiratory and digestive tracts.

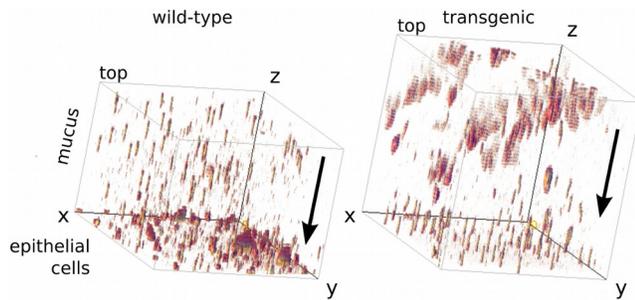
Mucus is a thick, slimy secretion, which is essential for many biological functions including lubrication, hydration and protection of the underlying epithelia (3, 22). Mucus is composed of water (~95%), salts, lipids and proteins, but its viscous and gel-like properties are largely governed by O-glycoproteins named gel-forming/gelling mucins. The intestinal mucosa contains billions of commensal bacteria, which represent a permanent challenge to the integrity of the epithelial surface (36). However, commensal bacteria compete for nutrients and sites of epithelial adherence with unwanted bacteria, protecting the underlying epithelium from penetration by pathogenic bacteria (17). Modifications to the mucus properties can greatly affect mucus layer functioning. For example, an intestinal mucus layer that is too thin, as found in inflammatory bowel diseases (IBDs), will facilitate bacteria reaching the epithelial cells which may trigger inflammation because of the dysregulated immune response to host intestinal microbiota. This has been demonstrated using several mouse models with a defective mucus layer which leads to direct contact between bacteria and the epithelium associated with a severe intestinal inflammation (15, 19). Conversely, thick mucus in the lungs makes it difficult to expel leading to lung obstruction as found in cystic fibrosis (CF) and chronic obstructive pulmonary disease (COPD). Thickened secretions are also features in the gastrointestinal system in CF (for review, see Kelly and Buxbaum (18). Meconium ileus and distal intestinal obstruction syndrome develop when thick mucous secretions occlude the hollow gastrointestinal lumen. In the pancreas and bile ducts of CF patients, thickened secretions may cause obstruction and acute and chronic inflammation. Strategies aimed at modulating mucus properties *in vivo* are limited, partly due to the complexity of mucin macromolecules.

The main characteristics of gelling mucins

result from their large, heavily O-glycosylated region, where carbohydrates retain water, and from disulfide bonds between mucin monomers. Gel-forming mucins crosslink via their amino- and carboxy-terminal regions which are enriched in cysteine residues to form either long polymers (28) or more complex structures responsible for net-like structures (16). Non-crosslinking interactions between mucin macromolecules seem also fundamental to mucus gelling (2, 3). Among the reversible cross-links within mucin polymers, intermolecular hydrophobic interactions are today the best characterized (7, 8, 10). However, few studies have investigated the mucus barrier or have attempted to modify mucin gelling *in vivo*. Several reports have suggested that a non-O-glycosylated domain interspersed within the O-glycosylated region participates in the mucin network (3, 14). This domain, named the CYS domain because it is enriched in cysteine residues, is ~110 amino acids long and is highly hydrophobic. The CYS domain has been found in two copies in human and mouse mucin MUC2, the major gelling mucin of the intestine, and in 7 and 9 copies, respectively, in the two respiratory mucins, MUC5B and MUC5AC (13). The CYS domain is not present in the two other gelling mucins, MUC6 which is expressed mainly in the stomach but also in deep glands of duodenum and ileum (34) and MUC19 which is expressed in submandibular glands and in trachea (11). In comparison to other domain sequences of gelling mucins, the CYS domain is highly conserved, with a strong selective pressure on many amino acid residues, supporting a key role of the domain (13). The CYS domain is highly hydrophobic and *ex-vivo* experiments suggested that CYS domains are able to interact with each other in a reversible manner (3, 8). Consequences of an increased number of reversible interactions between mucins on mucus properties *in vivo* have never been evaluated.

A transgenic mouse that secretes a recombinant molecule consisting of 12 consecutive identical copies of one CYS domain borrowed from human mucin MUC5B has been created (14). As expected, the gut mucus blanket is modified by the recombinant molecule. The mucus appears more robust and less permeable to inert particles (Fig. 1). In challenged mice, the transgene is associated with reduced susceptibility to chemically-induced colitis, faster clearance of the pathogen *Citrobacter rodentium* administered by gavage, and better

protection against bacterial translocation (14).



**Fig. 1:** Distribution of fluorescent microspheres (diam.  $\sim 1\mu\text{m}$ ) loaded at the surface of colonic mucus and analyzed by confocal microscopy. After 45 min incubation, there is slower sedimentation when the mucus gel is enriched by the delivery of a molecule comprised of 12 consecutive mucin CYS domains (transgenic mouse).

A compromised gut barrier function may facilitate the onset of many diseases where increased bacterial translocation and/or microbial products are a key feature such as cachexia, chronic liver diseases (31), gut infection, IBD (4), intestinal obstruction (30), chemotherapy-induced mucositis (32), and acute pancreatitis (12). Bacterial translocation may also occur after epithelial cell hypoxic injury due to trauma or as a result of bacterial overgrowth after surgery, injury (30) or antibiotic use (21). Detergent action of bile acids throughout the gastrointestinal tract has been suggested as a natural luminal aggressor. Many experimental data support that bile acids, and more especially deoxycholic acid, are cytotoxic to the epithelial cells (9, 32). However, bile acids induce also mucus secretion and expression of MUC2, two mechanisms by which the intestinal epithelium protects itself (20, 23). To date, no disease has been described where bile acids are the primary initiators of the epithelium damage (26) but we cannot rule out the exacerbation of epithelial damages by bile acids in several disease states where the mucus layer is less protective. The transgenic model shows that it is possible to reinforce the intestinal barrier by delivering a molecule made up of domains belonging to gelling mucins. This opens up new strategies to treat, limit or prevent unwanted bacterial translocation, especially from the gut.

The recent report showed that delivery of poly-CYS molecules affects mucin O-glycosylation (14). This modification has been suggested to be linked to the higher load of beneficial *Lactobacillus*

spp. found in the gut of transgenic mice (2.5 log/g of tissue). It is known that bacteria in the gut drive mucin maturation and use sugars from mucins as their energy source. Lactobacilli act as a primordial barrier to infection by competing for adhesion sites with pathogens, by producing lactic acid, bacteriocins, nonbacteriocin compounds and nonproteinaceous molecules that exercise a direct bactericidal effect (24) and stimulating the production of anti-microbial molecules by the host. Consequently, the unexpected increase in *Lactobacillus* spp. in transgenic mice may strengthen the intestinal barrier. Similarly, delivering poly-CYS molecules into the vagina could increase the abundance of lactobacilli, thereby preventing or limiting infections such as bacterial vaginosis, yeast vaginitis, urinary tract infection and sexually transmitted diseases (6). Delivering poly-CYS molecules into the cervical mucus may also represent a new method of contraception. In the cervical mucus, the poly-CYS molecule should favor reversible crosslinks between mucin macromolecules which would change the mesh size of the mucin network and decrease its permeability to sperm (8).

At least two strategies can be envisaged to enrich a mucus gel with molecules made of CYS domains. The first one consists in delivering the recombinant molecule using food-grade living lactic acid bacteria (5) or non-pathogenic yeast strains, like *Yarrowia lipolytica* (25). Delivery would be more efficient in mucus gels housing the greatest abundance of micro-organisms, i.e. colon, distal ileum and vagina. The second strategy uses the CRISPR/Cas9 system, a genomic technology which is in its infancy (29). CRISPR/Cas9 gene editing tool should enable to trans-activate in the gut the expression of the respiratory genes *MUC5B* and *MUC5AC*, which have the particularity to encode gelling mucins with 7 and 9 copies of the CYS domain, respectively.

There are many potential applications of mucus-enrichment with poly-CYS molecules. However, a high concentration of the domain in some mucus may be deleterious. For example, mucus makes it difficult for some compounds to reach the underlying epithelial cells limiting the efficacy of drugs administered orally or as aerosols. In CF, abnormal dehydrated mucus tempered hopes for correcting the mutated-*CFTR* gene, responsible for the disease. Perez-Villar and Boucher hypothesized that the increased mucus concentration in CF may result

from the formation of abnormal irreversible inter-chain bonds in airway mucins (27). Here, the increased production of respiratory mucins with 7 or 9 copies of the CYS domain may greatly favor interactions between CYS domains of gelling mucins, dangerously increasing mucus viscosity and stasis leading to obstruction of the airways. A better understanding of the interactions between CYS domains and amino acids engaged in these interactions would help identify new strategies to fluidify abnormal mucus in CF and COPD. Further studies of CYS domain properties will depend on the availability of recombinant molecules made of one or several copies of the domain, which seem particularly difficult to produce (1).

## ACKNOWLEDGMENTS

We apologize to those whose work is not cited owing to space constraints.

## GRANTS

This work was supported by the French Association against Cystic Fibrosis “Vaincre la Mucoviscidose,” the French Association against Crohn’s disease “Association François Aupetit,” the French Foundation on Digestive Tract Diseases and Nutrition “DigestScience,” the Broad and Medical Research Program (USA), the French Foundation “Fondation pour la Recherche Médicale,” and the “Région Nord-Pas de Calais.”

## DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

## AUTHOR CONTRIBUTIONS

J.-L.D. prepared figures; J.-L.D. drafted manuscript; J.-L.D., V.G., and F.G. edited and revised manuscript; J.-L.D., V.G., and F.G. approved final version of manuscript.

## References

1. **Bäckström M, Ambort D, Thomsson E, Johansson ME V, Hansson GC.** Increased understanding of the biochemistry and biosynthesis of MUC2 and other gel-forming mucins through the recombinant expression of their protein domains. *Mol Biotechnol* 54: 250–6, 2013.
2. **Bansil R, Stanley E, LaMont JT.** Mucin biophysics. *Annu. Rev. Physiol* 57: 635–657, 1995.
3. **Bansil R, Turner BS.** Mucin structure, aggregation, physiological functions and biomedical applications.

4. **Becker C, Neurath MF, Wirtz S.** The Intestinal Microbiota in Inflammatory Bowel Disease. *ILAR J* 56: 192–204, 2015.
5. **Bermúdez-Humarán LG, Aubry C, Motta JP, Deraison C, Steidler L, Vergnolle N, Chatel JM, Langella P.** Engineering lactococci and lactobacilli for human health. *Curr Opin Microbiol* 16: 278–83, 2013.
6. **Borges S, Silva J, Teixeira P.** The role of lactobacilli and probiotics in maintaining vaginal health. *Arch Gynecol Obstet* 289: 479–89, 2014.
7. **Bromberg LE, Barr DP.** Self-association of mucin. *Biomacromolecules*. 1: 325–334, 2000.
8. **Brunelli R, Papi M, Arcovito G, Bompiani A, Castagnola M, Parasassi T, Sampaolese B, Vincenzoni F, De Spirito M.** Globular structure of human ovulatory cervical mucus. *FASEB J* 21: 3872–3876, 2007.
9. **Camilleri M, Murphy R, Chadwick VS.** Dose-related effects of chenodeoxycholic acid in the rabbit colon. *Dig Dis Sci* 25: 433–8, 1980.
10. **Cao X, Bansil R, Bhaskar KR, Turner BS, LaMont JT, Niu N, Afdhal NH.** pH-dependent conformational change of gastric mucin leads to sol-gel transition. *Biophys J* 76: 1250–1258, 1999.
11. **Chen Y, Zhao YH, Kalaslavadi TB, Hamati E, Nehrke K, Le AD, Ann DK, Wu R.** Genome-wide search and identification of a novel gel-forming mucin MUC19/Muc19 in glandular tissues. *Am J Respir Cell Mol Biol* 30: 155–65, 2004.
12. **De\_Waele JJ.** Rational use of antimicrobials in patients with severe acute pancreatitis. *Semin Respir Crit Care Med* 32: 174–80, 2011.
13. **Desseyn JL.** Mucin CYS domains are ancient and highly conserved modules that evolved in concert. *Mol Phylogenet Evol* 52: 284–292, 2009.
14. **Gouyer V, Dubuquoy L, Robbe-Masselot C, Neut C, Singer E, Plet S, Geboes K, Desreumaux P, Gottrand F, Desseyn JL.** Delivery of a mucin domain enriched in cysteine residues strengthens the intestinal mucous barrier. *Sci Rep* 5: 9577, 2015.
15. **Johansson ME V, Gustafsson JK, Holmen-Larsson J, Jabbar KS, Xia L, Xu H, Ghishan FK, Carvalho FA, Gewirtz AT, Sjovall H, Hansson GC.** Bacteria penetrate the normally impenetrable inner colon mucus layer in both murine colitis models and patients with ulcerative colitis. *Gut* 63: 281–291, 2014.
16. **Johansson ME V, Sjovall H, Hansson GC, Sjövall H, Hansson GC.** The gastrointestinal mucus system in health and disease. *Nat Rev Gastroenterol Hepatol* 10: 352–361, 2013.
17. **Kaiko GE, Stappenbeck TS.** Host-microbe interactions shaping the gastrointestinal environment. *Trends Immunol* 35: 538–48, 2014.

18. **Kelly T, Buxbaum J.** Gastrointestinal Manifestations of Cystic Fibrosis. *Dig Dis Sci* 60: 1903–13, 2015.
19. **Khor B, Gardet A, Xavier RJ.** Genetics and pathogenesis of inflammatory bowel disease. *Nature* 474: 307–17, 2011.
20. **Klinkspoor JH, Mok KS, Van Klinken BJ, Tytgat GN, Lee SP, Groen AK.** Mucin secretion by the human colon cell line LS174T is regulated by bile salts. *Glycobiology* 9: 13–9, 1999.
21. **Knoop KA, McDonald KG, Kulkarni DH, Newberry RD.** Antibiotics promote inflammation through the translocation of native commensal colonic bacteria. *Gut* (June 4, 2015). doi: 10.1136/gutjnl-2014-309059.
22. **Lai SK, Wang YY, Wirtz D, Hanes J.** Micro- and macrorheology of mucus. *Adv Drug Deliv Rev* 61: 86–100, 2009.
23. **Lee HY, Crawley S, Hokari R, Kwon S, Kim YS.** Bile acid regulates MUC2 transcription in colon cancer cells via positive EGFR/PKC/Ras/ERK/CREB, PI3K/Akt/IkappaB/NF-kappaB and p38/MSK1/CREB pathways and negative JNK/c-Jun/AP-1 pathway. *Int J Oncol* 36: 941–53, 2010.
24. **Liévin-Le Moal V, Servin AL.** Anti-infective activities of lactobacillus strains in the human intestinal microbiota: from probiotics to gastrointestinal anti-infectious biotherapeutic agents. *Clin Microbiol Rev* 27: 167–199, 2014.
25. **Madzak C, Gaillardin C, Beckerich JM.** Heterologous protein expression and secretion in the non-conventional yeast *Yarrowia lipolytica*: a review. *J Biotechnol* 109: 63–81, 2004.
26. **Pavlidis P, Powell N, Vincent RP, Ehrlich D, Bjarnason I, Hayee B.** Systematic review: bile acids and intestinal inflammation-luminal aggressors or regulators of mucosal defence? *Aliment Pharmacol Ther* 42: 802–17, 2015.
27. **Perez-Vilar J, Boucher RC.** Reevaluating gel-forming mucins' roles in cystic fibrosis lung disease. *Free Radic. Biol. Med.* 37: 1564–1577, 2004.
28. **Ridley C, Kouvatso N, Raynal BD, Howard M, Collins RF, Desseyn JL, Jowitt TA, Baldock C, Davis CW, Hardingham TE, Thornton DJ.** Assembly of the respiratory mucin MUC5B: a new model for a gel-forming Mucin. *J Biol Chem* 289: 16409–16420, 2014.
29. **Sampson TR, Weiss DS.** Exploiting CRISPR/Cas systems for biotechnology. *Bioessays* 36: 34–8, 2014.
30. **Schietroma M, Pessia B, Carlei F, Cecilia EM, De Santis G, Amicucci G.** Laparoscopic versus open colorectal surgery for colon cancer: the effect of surgical trauma on the bacterial translocation. A prospective randomized study. *Am J Surg* 210: 263–9, 2015.
31. **Schnabl B, Brenner DA.** Interactions between the intestinal microbiome and liver diseases. *Gastroenterology* 146: 1513–24, 2014.
32. **Shekels LL, Lyftogt CT, Ho SB.** Bile acid-induced alterations of mucin production in differentiated human colon cancer cell lines. *Int J Biochem Cell Biol* 28: 193–201, 1996.
33. **Swank GM, Deitch EA.** Role of the gut in multiple organ failure: bacterial translocation and permeability changes. *World J Surg* 20: 411–7, 1996.
34. **Valque H, Gouyer V, Husson MO, Gottrand F, Desseyn JL.** Abnormal expression of Muc5b in Cfr-null mice and in mammary tumors of MMTV-ras mice. *Histochem. Cell Biol.* 136: 699–708, 2011.
35. **van Vliet MJ, Harmsen HJM, de Bont ESJM, Tissing WJE.** The role of intestinal microbiota in the development and severity of chemotherapy-induced mucositis. *PLoS Pathog* 6: e1000879, 2010.
36. **Zhang K, Hornef MW, Dupont A.** The intestinal epithelium as guardian of gut barrier integrity. *Cell Microbiol* 17: 1561–1569, 2015.