

Doubling diet fat on sugar ratio in children with mitochondrial OXPHOS disorders: Effects of a randomized trial on resting energy expenditure, diet induced thermogenesis and body composition

Laurent Beghin, Stephanie Coopman, Manuel Schiff, Joseph Vamecq, Karine Mention-Mulliez, Regis Hankard, Jean-Marie Cuisset, Helene Ogier, Frederic Gottrand, Dries Dobbelaere

▶ To cite this version:

Laurent Beghin, Stephanie Coopman, Manuel Schiff, Joseph Vamecq, Karine Mention-Mulliez, et al.. Doubling diet fat on sugar ratio in children with mitochondrial OXPHOS disorders: Effects of a randomized trial on resting energy expenditure, diet induced thermogenesis and body composition. Clinical Nutrition, Elsevier, 2016, 35 (6), pp.1414-1422. 10.1016/j.clnu.2016.03.015 . hal-02177005

HAL Id: hal-02177005 https://hal.univ-lille.fr/hal-02177005

Submitted on 8 Jul 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Doubling diet fat on sugar ratio in children with mitochondrial OXPHOS disorders: Effects of a randomized trial on resting energy expenditure, diet induced thermogenesis and body composition

4

5
6 Laurent Béghin ^{a,b,*}, Stéphanie Coopman ^a, Manuel Schiff ^c, Joseph Vamecq ^d, Karine Mention7 Mulliez ^{e,}, Régis Hankard ^{f,g}, Jean-Marie Cuisset ^h, Hélène Ogier ^c, Frédéric Gottrand F ^{a,b}, Dries
8 Dobbelaere ^e.

- 9
- 10

^a Centre d'Investigation Clinique, CIC-1403–Inserm–CH&U, Lille University Hospital F-59037 Lille
 France.

- 13 ^b LIRIC- Lille Inflammation Research International Center/UMR U995 Inserm, Lille France
- ¹⁴ ^c Reference center for Inherited Metabolic Diseases, Robert Debré University Hospital, Paris, France

^d Inserm, Department of Biochemistry and Molecular Biology, HMNO, CBP, CHRU Lille and and RADEME EA 7364, Lille Nord of France University, F-59037 Lille, France

- ^e Reference center for Inherited Metabolic Diseases in child and adulthood, Lille University Children's Hospital Jeanne de Flandre, and RADEME EA 7364, Lille Nord of France University, F-59037 Lille, France
- 20 ^f Inserm U 1069, F Rabelais University, Tours, F-37000, France
- 21 ^g CIC-1426- Inserm-CH&U of Robert Debré University Hospital, F-75935 Paris, France
- 22 ^h Pediatric Neurology Unit Lille University Hospital, F-59037 Lille, France
- 24 Short title: Doubling diet fat on sugar ratio in mitochondriopathies
- ** Corresponding author* : Dr Laurent BEGHIN, LIRIC-UMR-995-Inserm and CIC-1403-Inserm-CH&U
 de Lille
- 27 Centre d'Investigation Clinique, Antenne pédiatrique, Hôpital Jeanne de Flandre, CHRU de Lille, F-
- 28 59037 LILLE Cédex, Tél : +33 3 20 44 60 58, Fax : +33 3 20 44 66 87,
- 29

30 *E-mail addresses:* <u>laurent.beghin@chru-lille.fr</u> (L. Béghin), <u>Stephanie.coopman@chru-lille.fr</u> (S.

- 31 Coopman), manuel.schiff@aphp.fr (M. Schiff), <u>joseph.vamecq@inserm.fr</u> (J. Vamecq),
- 32 <u>karine.mention@chru-lille.fr</u> (K. Mention-Mulliez), regis.hankard@univ-tours.fr (R. Hankard),
- 33 (J.M. Cuisset), (H. Ogier), <u>Frederic.gottrand@chru-lille.fr</u> (F. Gottrand), <u>dries.dobbelaere@chru-</u>
- 34 <u>lille.fr</u> (D. Dobbelaere)
- 35
- 36 Abbreviations:
- 37 ANOVA : <u>A</u>nalysis <u>Of V</u>ariance.
- 38 BD : <u>B</u>asal <u>D</u>iet
- 39 CD : <u>Challenging Diet</u>
- $40 \qquad {\rm CONSORT: Consolidated \ Standards \ of \ Reporting \ Trials}$
- 41 DIT : <u>Diet Induced Thermogenesis</u>
- 42 MOD : <u>Mitochondrial OXPHOS D</u>isorder
- 43 OXPHOS : <u>Ox</u>idative <u>Phos</u>phorylation
- 44 REE : <u>R</u>esting <u>Energy</u> <u>Expenditure</u>

SUMMARY

45 Background and aims: Mitochondrial OXPHOS disorders (MODs) affect one or several complexes

- 46 of respiratory chain oxidative phosphorylation. An increased fat/low-carbohydrate ratio of the diet
- 47 was recommended for treating MODs without, however, evaluating its potential benefits through
- 48 changes in the respective contributions of cell pathways (glycolysis, fatty acid oxidation) initiating
- energy production. Therefore, the objective of the present work was to compare Resting Energy
 Expenditure (REE) under basal diet (BD) and challenging diet (CD) in which fat on sugar content
- 51 ratio was doubled. Diet-induced thermogenesis (DIT) and body compositions were also compared.
- 52 Energetic vs regulatory aspects of increasing fat contribution to total nutritional energy input were
- 53 essentially addressed through measures primarily aiming at modifying total fat amounts and not
- 54 the types of fats in designed diets.
- 55 *Methods:* In this randomized cross-over study, BD contained 10% proteins/30% lipids/60% 56 carbohydrates (fat on sugar ratio = 0.5) and was the imposed diet at baseline. CD contained 10% 57 proteins/45% lipids/45% carbohydrates (fat on sugar ratio = 1). Main and second evaluation
- 58 criteria measured by indirect calorimetry (QUARK RMR[®], Cosmed, Pavona; Italy) were REE and DIT,
- 59 respectively. Thirty four MOD patients were included; 22 (mean age 13.2±4.7 years, 50% female;
- 60 BMI 16.9±4.2 kg/m²) were evaluated for REE, and 12 (mean age 13.8±4.8 years, 60% female; BMI
- 61 17.4±4.6 kg/m²) also for DIT. OXPHOS complex deficiency repartition in 22 analysed patients was 62 55% for complex I, 9% for complex III, 27% for complex IV and 9% for other proteins.
- 63 *Results:* Neither carry-over nor period effects were detected (p=0.878; ANOVA for repeated 64 measures). REE was similar between BD vs CD (1148.8±301.7 vs 1156.1±278.8 kcal/day; p=0.942) 65 as well as DIT (peak DIT 260 vs 265 kcal/day; p=0.842) and body composition (21.9±13.0 vs 66 21.6±13.3 % of fat mass; p=0.810).
- 67 *Conclusion:* Doubling diet fat on sugar ratio does not appear to improve, per se, energetic status
- 68 and body composition of patients with MODs.69
- 70 Keywords: Mitochondrial disorders or diseases, Energy expenditure, Diet induced thermogenesis,
- 71 Body composition.
- 72

73 **1. Introduction**

74

75 Mitochondrial OXPHOS disorders (MODs) encompass a group of rare genetic disorders 76 affecting respiratory chain oxidative phosphorylation (OXPHOS) and resulting from mitochondrial 77 or nuclear DNA defects [1,2]. The OXPHOS system plays a key role in transferring energy from 78 macronutriments to ATP through a sequence of coordinated reactions by which macronutriments 79 are oxidized. OXPHOS comprises five major protein-lipid enzyme complexes which are located in 80 mitochondrial inner membrane and work locally in synergy with the labile electron acceptors 81 ubiquinone (an amphiphilic isoprenoid compound) and cytochrome c (a 12 kDa peptide). Electrons 82 generated during glucose, fatty acids and amino acid oxidations are channeled across complexes I 83 and II to be transferred sequentially to coenzyme Q10, complex III, and complex IV. Complexes I, 84 III, and IV use part of the energy produced by the electron transfer flux to extrude protons across 85 the mitochondrial inner membrane, from mitochondrial matrix to mitochondrial intermembrane 86 space. The resulting proton gradient is consumed with energy utilization and production by 87 complex V which condenses adenosine diphosphate and inorganic phosphate into adenosine 88 trisphosphate (ATP) [3,4]. These overall mechanisms and players for ATP production are impaired 89 in disorders affecting OXPHOS, jeopardizing various cell functions notably those which require a 90 high energy supply. [5]. Currently, no satisfactory curative treatment for MODs exists. Only 91 symptomatic treatments of MODs are available [2,7,8]. Interestingly, dietary modulations have 92 been considered as regards to the key-role of mitochondria in energy production from 93 macronutrients [9,10].

94 Increasing fat vs carbohydrate content in the diet has been recommended for the 95 treatment of MOD. Several reasons justify this recommendation. (i) Mitochondrial oxidation of 96 NADH is thought to be diminished in MOD [5,11] patients especially those affected with 97 respiratory chain Complex I (CI) deficiencies, it might be by-passed by FADH₂, an alternative carrier 98 of reducing equivalents, the electrons of which can enter the respiratory chain distal to complex I. 99 (ii) Supply in FADH₂ to the mitochondria can be increased (relative to NADH) by increasing the 100 amount of triacylglycerols and fatty acids in the diet. On the basis of stoichiometry of 101 mitochondrial oxidations, fatty acids yield a ratio of FADH to NADH of 0.5, whereas glucose yields 102 a much lower ratio of 0.2. In addition, fatty acids in contrast to glucose are endowed with 103 uncoupling properties towards OXPHOS [12]. Though uncoupling OXPHOS lowers net ATP 104 production by mitochondria, it virtually accelerates mitochondrial respiration through increased

105 electron transfer rates, a feature which improves mitochondrial oxidations regulated by the106 NADH/NAD+ and FADH2/FAD ratios.

In the wake of this rationale, we hypothesized that increasing fat to glucose ratio within an 107 108 isoenergetic and isonitrogenous diet should improve energy substrate oxidation in MOD patients. 109 As a corollary, this dietary measure should actually increase resting energy expenditure (REE), an 110 assumption also based on *in vitro* data on human cell [13,14] and animal studies [15], and *in vivo* 111 data obtained in complex I deficiency mitochondrial myopathy patients [16–19]. The aim of the 112 present study was, therefore, to study REE under basal diet (referred throughout the manuscript 113 to as the basal diet (BD)) challenged by a diet in which fat on carbohydrate content ratio was 114 doubled (referred to as the challenging diet (CD)) as a potential treatment in children with a MOD. 115 In these children, diet induced thermogenesis (DIT) and body composition were also measured to 116 get a better account for whole body energetic balances under the two diets.

117

118 **2.** Materials and methods

119

120 2.1. Trial design and participants

121 This 3-year prospective study (2008-2011) included MOD patients from the reference 122 centers for Inherited Metabolic Diseases of Lille and Paris (Jeanne de Flandre Lille University 123 Hospital; Robert Debré Paris University Hospital). Inclusion criteria were : (i) Child or adult 124 between 5 to 21 years with MOD [20], (ii) MOD subjects given a stable balanced diet having the 125 characteristics of basal diet (BD : 60 % carbohydrates, 30 % lipids, 10 % proteins) for at least one month. Subjects were excluded from the study when they required hospitalization or presented 126 127 with any acute condition (such as infection) known to interfere with energy metabolism 128 assessment.

As shown in flow CONSORT diagram [21] on (Figure 1), from 36 MOD subjects included over a period of four years, 22 accepted compliance to the diets and examinations, and were analysed for principal evaluation criterion (*ie* REE in Group 2), and from these 22, only 12 accepted to undergo the secondary evaluation criterion (*ie* DIT in Group 3).

This study was a randomized cross-over study (Figure 2). Method used to generate sequence was a simple randomisation by one-to-one allocation ratio. Subject randomisation was performed using sealed envelopes. REE were measured at four time points (*ie* Visit 1: baseline, Visit 2: after one month of basal or challenging diet ; Visit 3: after wash-out period consisted of one month of BD ; and Visit: 4 after one month of basal or challenging diet; see Figure 2). Neither

- carry-over effect, nor period effect was detected (p = 0.878; ANOVA for repeated measures). All
 assessments were performed in two Clinical Investigation Centers (CIC-1403-Inserm-CH&U of Lille;
 CIC-9202-Inserm-AP-HP of Paris). Before each assessment visit, all subjects arrived by car to the
 Clinical Investigation Center at 8:00 h, being fasted from 20:00 h on the previous day. Weight and
 height were first measured, and then compliance with the imposed diet was evaluated.
- 143

144 2.2. Nutrition protocol management

145 Intervention of the trial consisted of a CD diet period of one month (*ie* 10 % proteins, 45 %
146 lipids, 45 % carbohydrates). CD and BD diets were isoenergetic and isonitrogenous diets.

Each patient was randomly assigned to the basal diet (BD) for the control period vs the challenging diet (CD) for the intervention period. Each one-month period was thereafter inversed according to a cross-over trial. BD consisted of 10 % proteins, 30 % lipids, 60 % carbohydrates. To reach this BD, research dieticians used a specific booklet communicated to subjects and/or their parents to lower fat intake. CD consisted of 45 % lipids, 45 % carbohydrates ; 10 % proteins.

DIT was performed after a calibrated breakfast corresponding to 20% of recommended daily energy intake according to Martin *et al*'s formula [22]. According to the cross-over design, breakfast had a BD in control period and a CD in intervention period. Basal content breakfast consisted of bread, skimmed milk and jam. Challenging breakfast consisted of bread, full-cream milk and chocolate paste. After breakfast, meals were re-weighted and diet energy supplies were analysed by the difference between served and returned meals [23].

158 The assessment of energy intake and diet composition was performed using dietary record technique previously reported [24]. The technique used a specific chart report form including how 159 160 to record the foods consumed, using size instruments such as pictures of graduated bowls, cups, dishes, number of spoons. Each file corresponding to one day was separated in four parts: 161 162 breakfast, lunch, dinner and snack. Lunch and dinner were detailed as starter, main course, and desert [25]. Both oral and written instructions were given to each patient and/or parents on how 163 164 to keep accurate records using the size instruments, and the parents assisted the children in recording, identifying and quantifying the foods consumed. Subjects detailed daily food intakes on 165 166 the specific chart report form during 7 d [26]. The same project related dietician reviewed each 167 specific chart report. Quantification of meals and drinks were checked by the same trained 168 dietician, the parents and the child together. The dietician was well trained to identify both miss 169 and over-reporting and coherency of food records. The portion sizes were estimated using a three 170 dimensional portion size instrument with French current meal food photographs corresponding to

an exact BILNUT software (version 6 : Paris, France). This method has been shown to give unbiased records of energy intake in lean subjects up to 9 y old and showed that children aged 8-15 y were able to estimate food quantity to within = 10 % of the amount really eaten, suggesting that children could quantify their food intake with reasonable accuracy [27].

175

176 2.3. Indirect calorimetry assessment of energy expenditure

177 The subject rested recumbent on a hospital bed watching a videotape for 15 min. Same 178 quiet cartoons or videos (depending on age range of the subject) were used for all the subjects 179 [28]. Oxygen consumption (Vo₂ in mL/min) and carbon dioxide production (Vco₂ in mL/min) were 180 determined by using the open-circuit indirect calorimeter validated QUARK® RMR (Cosmed, 181 Pavona di Albano, Italy). The validity and accuracy measurements were guaranteed by yearly 182 calibration of the flow settings by the manufacturer of the calorimeter and guaranty a high 183 accuracy of QUARK[®] RMR between study centres [29]. Calibration of zero, span and delay 184 alignment of the O₂ and CO₂ gas analyzers was performed daily before each test using a certified 185 calibration gas.

186 The QUARK[®] RMR was equipped with a canopy hood for spontaneously breathing subjects. 187 With this device, flow rate was directly measured with a digital turbine flowmeter. Accuracy of the 188 flowmeter was 2%. Ventilatory rate was regulated directly by the QUARK® RMR. During each test, 189 the readings were controlled and eventually compensated by means of periodic automatic room 190 air calibrations. Response times of O₂ and CO₂ sensors were less than 120 ms. The O₂ analyzer was 191 a paramagnetic sensor, which had a measuring range from 0 to 30% in the canopy mode. Accuracy 192 of the O₂ sensor was 0.02 %. The CO₂ analyzer was an infrared digital sensor with a measuring 193 range standing from 0 to 10%. Accuracy of the CO₂ sensor was 0.02%. The flow meter which 194 detects the ventilation rate of the canopy was a bidirectional turbine with an 18 mm diameter. 195 The ventilation range was from 0 to 80 L/min. Accuracy of the flowmeter was 2%. QUARK® RMR 196 software utilized the Weir equation to assess Resting Energy Expenditure (REE) [30].

197DIT was measured at two occasions (Visit 2 and Visit 4) after REE measurement. DIT was198derived from postprandial energy expenditure (PEE) as described by Dabbech M *et al* [31]. PEE199assessment started immediately after a calibrated breakfast (see above for energetic ratio). DIT200was computed by subtracting REE from PEE (*ie* DIT = PEE – REE) according to Dabbech M *et al* [31].

- 201
- 202
- 203

204 2.4. Assessment of body composition by Dual X-ray Absorptiometry

The DXA (Dual X-Ray Absorptiometry) instrument was a Lunar DPX-IQ "pencilbeam" (Lunar 205 206 Radiation Corporation, Madison, WI, USA). Whole-body scan time was 560-590 s, and the 207 radiation dose was less than 0.03 mrem (¼ 0.3 mSv). Total body image acquisition and analysis 208 were obtained by following strictly manufacturer's specified instructions. The use of two photon 209 energies reduced errors due to irregular soft-body-mass and body contours. Two photon energies 210 discriminated two substances in a given system to a high accuracy. When more than two 211 substances, the discrimination depended on the number of additional substances, their 212 attenuation characteristics, and their relative contents [32]. FFM (fat-free mass), FM (fat mass) and 213 weight were calculated using the software provided by the manufacturer, and was adapted for the 214 subject age.

215

216 2.5. Regulatory requirement

The protocol was approved by French appropriates regulatory authorities (Direction Générale de la Santé; DGS 2007-A0018-50) and ethical review committee (Comité de Protection des Personnes Nord-Ouest IV ; CPP 07/32). Study protocol was also declared to clinical trial.gov register (NCT02385565). Participation in the study was voluntary and written informed consent was obtained from both parents and children. Written informed consent was obtained by the subject when patient was aged over 18 years.

223

224 2.6. Statistical analysis

Required sample size was computed using previously published data from complex I deficiency (CID) mitochondrial myopathy patients [16,17] with REE = 1120 ± 93 Kcal/min in basal condition (Glucose infusion). REE obtained in hyperlipid MOD condition (Triacylglycerol infusion) were 5 % higher at 1175 ± 93 Kcal/min. We hypothesised a 10% difference of REE between basal and challenging diets. According to the cross-over study design of the study and 80% statistical power, the sample size was defined at 20 patients to detect a significant difference between diets.

Statistical analyses were performed using SAS V.9 (Cary, USA). According to a Shapiro-Wilk test, all variables had a normal distribution. In this context, parametric test χ^2 for ratio, and to a cross-over trial design: period effect was tested by ANOVA analysis for repeated measures, carry over effect was tested by Student *t* test. Difference between two diets was tested by Student *t* test when number of subjects was above 15 [33,34]. When number of subjects was under 15, we

- 236 used non-parametric tests: *ie* Wilcoxon for paired analyses and Mann-Whitney for non-paired
- analyses.

3. Results

242

243 3.1. Anthropometric, biochemical defects and main clinics of enrolled patients

244 Anthropometric characteristics of patients from each of the 3 groups defined in the flow 245 CONSORT diagram illustrated at Figure 1 are presented on Table 1. No statistical differences were 246 observed between included and analysed subjects for REE, and DIT measurements. Table 2 further 247 summarizes main clinical and biochemical results of respiratory chain activities in skeletal muscle 248 and/or cultured skin fibroblasts. The most frequent defect was complex I deficiency at 55% (n=12). 249 Table 3 presents a minimum set for anthropometric data description of patients with complex I 250 deficiency and those with other defects. There was no difference between groups excepted for Z-251 score BMI which was significantly lower in the group of patients with defects other than complex I 252 deficiency.

253

254 3.2. Study of patients undergoing REE measurements

255 *3.2.1. Food intakes*

256 Food intakes in fat, carbohydrates and proteins were determined during the cross-over 257 periods in the 22 patients undergoing REE measurements (Table 4).

258 3.2.2. Metabolic REE measurements and anthropometrics

As mentioned in the experimental section, REE were measured at four time points (see Figure 2). No carry-over effect and no period effect was detected for the present study (p = 0.878; ANOVA for repeated measures).

262 Table 5 presents full metabolic data at baseline and after basal vs challenging diet 263 conditions. Measured REE were close to the predictive REE. No significant differences were 264 observed between the basal and challenging periods for any parameters (ie Vo₂, Vco₂, REE, RQ) 265 even when these parameters were adjusted by body composition parameters (ie weight, fat mass, fat free mass, muscle weight). Patient body composition parameters did not also differ between 266 267 basal and challenging diet periods for whole analyzed population (table 6). There were no 268 differences between basal and challenging diet periods for measured REE and anthropometrics in 269 patients with complex I deficiency nor in the group of patients affected by others defects. (Tables 270 7 and 8)

- 271
- 272
- 273

275

276 3.3. Study of patients undergoing DIT measurements

277 *3.3.1. Food intakes*

In a subset of 12 subjects, it was possible to measure DIT. Table 9 presents respective carbohydrate, lipid and protein contents of basal and challenging breakfasts. According to these data, designed diet conditions (*ie* basal *vs* challenging) were in practice respected

281 3.3.2. DIT measurements

Figure 3 gives time-courses of DIT over a postprandial period of 250 *min*. Maximum DIT was obtained at 100 ± 27 *min* after basal breakfast and 122 ± 29 *min* after challenging breakfast (p = 0.861). No significant differences in DIT optima, areas under the curve, time durations of postprandial periods were observed between basal *vs* challenging diet conditions. Similarly, RQ during DIT measurements were not significantly different between basal and challenging diet conditions (Figure 4).

- 288
- 289

290

4. Discussion

292 Because of the well-established ability of fats to induce mitochondrial biogenesis, we 293 aimed to test efficacy of an increased fat on carbohydrate ratio of the diet to manage nutrition in 294 MODs in order to improve energy expenditure metabolism [15–18]. Our study was designed to 295 determine to what extent and how REE, and secondarily DIT and body composition, may be 296 impacted by doubling fat vs carbohydrate contribution to nutritional energy intake. The collected 297 data do not confirm our primary hypothesis of a higher REE in challenging diet conditions. 298 Therefore, they do not a priori corroborate previous benefits suggested for high-fat diet in MODs 299 in a specific subset of patients using a specific type of fat enrichment. Our study present several 300 strengths (i) the cross-over design that is perfectly adapted to the aims of our study and the rarity 301 and heterogeneity of MODs, without period and carry-over effect, (ii) an adequate sample size, (iii) 302 a very well controlled diet, (iv) a well characterized population and (v) the use of gold standard 303 methodology based on previously validated tools (ie indirect calorimetry, body composition

304 analysis). Duration of the cross-over periods of one month was considered to be sufficient for a 305 high stability of energy metabolism and changes in REE and DIT. The absence of differences of REE 306 between basal and challenging diets observed in our study might be explained either by the wide 307 genetic heterogeneity usually observed in subjects with a mitochondrial disease [35], and masking 308 the power/homogeneity effect or by no impact of the challenging diet on REE, DIT and body 309 composition. In this respect, majority of previous trials have been performed on complex I MOD 310 [15,16,36,37] and, until now, trials mixing patients with distinct complex deficiencies were lacking. 311 The present study overcomes this gap and allows comparison with previous studies, some extra 312 statistical analyses were performed on REE and body composition in the subset of complex I MOD 313 patients included in our study. These extra analyses did not detect any difference in REE and body 314 composition in this subset of patients (Tables 7 and 8). This result does not reject the 315 heterogeneity of our population as a cause mentioned above for a lack of benefit of challenging diet in MODs as even in Complex I defect, there is a huge genetic variability [38]. Concerning 316 317 durations of the cross-over period, one month was an adapted period to observe any changes in 318 REE and DIT in view of one week as a known successful period to normalize complex I activity in 319 malnourished patients [37]. We did not observe any difference in DIT (ie energy expenditure and 320 RQ) as Flatt observed in healthy subject [39,40]. The last hypothesis of the lack of impact of CD on 321 energy metabolism despite a higher amount of energy intake at breakfast (+ 37 % see Table 9) 322 might result from the fact that the CD tested was at a 45% instead of a 50% dietary fat content, 323 though no effect of CD was also observed by De Meer K et al [41]. An important feature also is that 324 our CD was primarily aimed at doubling fat vs carbohydrate contribution to nutritional energy 325 intake and not to enrich fat qualitatively with a specific type of fats. Using a specific type of fat 326 Roef et al [16] found that triacylglycerol infusion improved some metabolic parameters (ie VCo₂, 327 respiratory quotient) during physical exercise in a sample of patients with deficient complex I. The 328 significant effect observed in their study may be accounted for by several factors :

329 (i) a specific effect of triacylglycerol intravenous infusion which is more energetic
 330 efficient than oral intake because of a high rate of systemic delivery and contents
 331 in medium-chain triglycerides (MCT). MCT are more energetic efficient that long 332 chain triglycerides because of their faster handling by metabolic pathways. This
 advantage is used in ketogenic diets where high fat ratios force the body to use
 fats instead of carbohydrates [42]. The underlying energetic change in oxidative

- phosphorylation (OXPHOS) lies in fatty acids yielding a ratio of FADH to NADH of
 0.5, whereas glucose yields a much lower ratio of 0.2
- 337 (ii) measurements performed during a physical exercise occur during sustained
 338 substrate oxidation, allowing a better detection of differences in energetic
 339 metabolism,
- 340 (iii) a specific population response (only 4 subjects in this study). The absence of 341 response in our population even if we analyzed a subgroup of complex I-deficient 342 patients might be attributed to the lack of mechanisms covered by parenteral 343 route, MCT and physical exercise which are all conditions absent from our 344 experimental protocols. This does not preclude that the ketogenic diet does not 345 improve mitochondrial oxidations as a source of metabolic energy, but this 346 feature unfortunately failed to be detected by our study. In this respect, the 347 lower fat enrichment of our CD vs classic ketogenic diet (43.6 % of fat in our CD 348 vs 90% in ketogenic diet) is another aspect to take into account.
- 349 As MODs may exhibit a wide range of symptoms including developmental delay, seizures, 350 vision and hearing impairments, autonomous nervous dysfunction, gastrointestinal signs, 351 endocrine disturbances and failure to thrive, a special interest of these symptoms were checked 352 during the study period. No adverse reactions, adverse events of special interest, or serious 353 adverse events were observed excepted some fatigue in 6 patients from 24 randomized patients 354 only during basal diet period (not significant). As a whole, doubling fat vs carbohydrate contribution to nutritional energy intake does not improve, per se, energetic status and body 355 356 composition of patients with MODs but does not also impact negatively patients. However, 357 undernutrition observed in defects other than complex I could suppose a higher defect in weight 358 energy balance in these patients vs complex I-deficient patients.
- 359

5. Conclusion

362

363 As a whole, this study does not provide evidence that doubling fat *vs* carbohydrate ratio of 364 the diet increases whole-body energy expenditure in patients with MOD. We were not able to potential benefit in CD for mitochondrial diseases [5]. Our study suggests that underlying mechanisms for a potential beneficial effect for CD does not a priori involve modulations of REE, nor DIT. The wide heterogeneity of the studied population and/or the amount/quality of lipids in the diet might play a role as well in the generated data and subsequent conclusions Conflict of Interest Statement & Statement of Authorship. Frédéric Gottrand has received consulting fees from Numico Clinical Nutrition, lecture fees from SMS and grant support from Danone Research. The remaining authors state no conflict of interest. Acknowledgments We thank parents and patients who have taken part in the study. Pascale HINCKER & Sarah KAMMENEY for diet composition management. We thank Christelle GUIMBER study nurse from CIC for paramedical/logistical assistance. We thank Brigitte PORAS, José CANOMORALES, Stéphane DESCHILDER, Nathalie DUQUESNOY, Jeanne-Marie DESCAMPS-BEN, Valérie HELD, Isabelle CHARDON, Laurence CRINON for DXA acquisition and Dr Georges LION for DXA validation and interpretation. We thank Muriel BEUVRY and Anne GAUTREAU (CIC-PT-1403-CH&U-Inserm de Lille, France) for typing this manuscript. Funding Lille University Hospital was the trial sponsor of this study. This study was founded by a grant from French ministry of health (PHRC interregional N°2003/1901). References

demonstrate a lowering of the metabolic challenge induced by substrate oxidation as a suggested

399 [1] Vafai SB, Mootha VK. Mitochondrial disorders as windows into an ancient organelle. Nature 2012;
 400 491:374-383.

- 401 [2] Koopman WJ, Willems PH, Smeitink JA. Monogenic mitochondrial disorders. N Engl J Med 2012; 366:
 402 1132-1141.
- 403[3] Di Donato S. Disorders related to mitochondrial membranes: pathology of the respiratory chain and404neurodegeneration. J Inherit Metab Dis 2000;23:247-263.
- 405[4] Roede JR, Go YM, Jones DP. Redox equivalents and mitochondrial bioenergetics. Methods Mol Biol4062012;810:249-280.
- 407[5] Munnich A, Rotig A, Chretien D, Cormier V, Bourgeron T, Bonnefont JP, Saudubray JM, Rustin P.408Clinical presentation of mitochondrial disorders in childhood. J Inherit Metab Dis 1996 ;19:409521-527.
- 410 [6] Thorburn DR. Mitochondrial disorders: prevalence, myths and advances. J Inherit Metab Dis 2004; 411 27:349-362.
- 412 [7] Schiff M, Benit P, Coulibaly A, Loublier S, El-Khoury R, Rustin P. Mitochondrial response to controlled 413 nutrition in health and disease. Nutr Rev 2011 69: 65-75.
- Schiff M, Benit P, Jacobs HT, Vockley J, Rustin P. Therapies in inborn errors of oxidative metabolism.
 Trends Endocrinol Metab 2012;23:488-495.
- 416 [9] Passarella S, Atlante A, Valenti D, de BL. The role of mitochondrial transport in energy metabolism.
 417 Mitochondrion 2003;2:319-343.
- 418 [10] Wallace DC, Fan W, Procaccio V. Mitochondrial energetics and therapeutics. Annu Rev Pathol 2010;
 419 5:297-348.
- 420 [11] Gwynne J. The role of nutrition in mitochondrial an metabolic diseases. A primary care physician's421 guide 2000;15-16.
- 422 [12] Mokhova EN, Khailova LS. Involvement of mitochondrial inner membrane anion carriers in the 423 uncoupling effect of fatty acids. Biochemistry (Mosc) 2005;70:159-63
- 424 [13] Doctor RB, Bacallao R, Mandel LJ. Method for recovering ATP content and mitochondrial function 425 after chemical anoxia in renal cell cultures. Am J Physiol 1994;266:C1803-C1811.
- 426 [14] Santra S, Gilkerson RW, Davidson M, Schon EA. Ketogenic treatment reduces deleted mitochondrial
 427 DNAs in cultured human cells. Ann Neurol 2004;56:662-669.
- 428 [15] Schiff M, Benit P, El-Khoury R, Schlemmer D, Benoist JF, Rustin P. Mouse studies to shape clinical
 429 trials for mitochondrial diseases: high fat diet in Harlequin mice. PLoS One 2011;6:e28823.
 430 10.1371/journal.
- 431[16]Roef MJ, de Meer K, Reijngoud DJ, Straver HW, de Barse M, Kalhan SC, Berger R. Triacylglycerol432infusion improves exercise endurance in patients with mitochondrial myopathy due to433complex I deficiency. Am J Clin Nutr 2002;75: 237-244.
- 434[17]Roef MJ, de Meer K, Reijngoud DJ, Straver HW, de Barse M, Kalhan SC, Berger R. Triacylglycerol435infusion does not improve hyperlactemia in resting patients with mitochondrial myopathy436due to complex I deficiency. Am J Clin Nutr 2002;75: 228-236.

- 437 [18] Roef MJ, Reijngoud DJ, Jeneson JA, Berger R, de MK. Resting oxygen consumption and in vivo ADP are
 438 increased in myopathy due to complex I deficiency. Neurology 2002;58: 1088-1093.
- 439 [19] Panetta J, Smith LJ, Boneh A. Effect of high-dose vitamins, coenzyme Q and high-fat diet in paediatric
 440 patients with mitochondrial diseases. J Inherit Metab Dis 2004;27: 487-498.
- 441 [20] Bernier FP, Boneh A, Dennett X, Chow CW, Cleary MA, Thorburn DR. Diagnostic criteria for 442 respiratory chain disorders in adults and children. Neurology 2002 ;59: 1406-1411.
- 443 [21] Schulz KF, Altman DG, Moher D. CONSORT 2010 statement: Updated guidelines for reporting parallel 444 group randomised trials. *J Pharmacol Pharmacother* 2011 ;1: 100-107.
- 445 [22] Martin A. Apports nutritionnels conseillés pour la population française. TEC et DOC Lavoisier 2000;
 446 23-43.
- 447 [23] Belko AZ, Barbieri TF, Wong EC. Effect of energy and protein intake and exercise intensity on the 448 thermic effect of food. Am J Clin Nutr 1986 ; 43: 863-869.
- 449[24] Barnard JA, Tapsell LC, Davies PS, Brenninger VL, Storlien LH. Relationship of high energy expenditure450and variation in dietary intake with reporting accuracy on 7 day food records and diet451histories in a group of healthy adult volunteers. Eur J Clin Nutr 2002;56: 358-367.
- 452 [25] Informatic Center for Quality of foods. Pictures of different meals 1993 ; 1-63.
- 453 [26] Welch AA, McTaggart A, Mulligan AA, Luben R, Walker N, Khaw KT, Day NE, Bingham SA. DINER (Data
 454 Into Nutrients for Epidemiological Research) a new data-entry program for nutritional
 455 analysis in the EPIC-Norfolk cohort and the 7-day diary method. Public Health Nutr 2001;4:
 456 1253-1265.
- 457 [27] Chattaway FW, Happold FC, Happold AM. Nutrition of school-children in Leeds, winter, 1943, and 458 summer, 1944. Br Med J 1946;1: 429.
- 459 [28] Dougherty FE. Metabolic testing in mitochondrial disease. Semin Neurol 200 ;21: 303-308.
- 460[29] Blond E, Maitrepierre C, Normand S, Sothier M, Roth H. A new indirect calorimeter is accurate and461reliable for measuring basal enerby expenditure, thermic effect of food and substrate462oxidation in obese and healthy subjects. e-jounal of Clinical nutrition 2010 ; .
- 463 [30] Weir JB. New methods for calculating metabolic rate with special reference to protein metabolism. J
 464 Physiol 1949;109: 1-9.
- [31] Dabbech M, Boulier A, Apfelbaum M, Aubert R. Thermic effect of meal and fat mass in lean and obese
 men. Nutrition Research 1996; 16:1133-1141.
- 467 [32] Goran MI, Driscoll P, Johnson R, Nagy TR, Hunter G. Cross-calibration of body-composition techniques
 468 against dual-energy X-ray absorptiometry in young children. Am J Clin Nutr 1996;63:299-305.
- 469 [33] Putt M, Chinchilli VM. A mixed effects model for the analysis of repeated measures cross-over470 studies. Stat Med 1999 18:3037-3058.
- 471 [34] Chen X, Wei L. A comparison of recent methods for the analysis of small-sample cross-over studies.
 472 Stat Med 2003;22:2821-2833.
- 473 [35] Benit P, El-Khoury R, Schiff M, Sainsard-Chanet A, Rustin P. Genetic background influences
 474 mitochondrial function: modeling mitochondrial disease for therapeutic development. Trends
 475 Mol Med 2010; 16:210-217.

- 476[36]Kirby DM, Crawford M, Cleary MA, Dahl HH, Dennett X, Thorburn DR (1999) Respiratory chain477complex I deficiency: an underdiagnosed energy generation disorder. Neurology 52:1255-4781264.
- 479 [37] Briet F, Twomey C, Jeejeebhoy KN. Effect of feeding malnourished patients for 1 mo on mitochondrial
 480 complex I activity and nutritional assessment measurements. Am J Clin Nutr 2004;79:787 481 794.
- 482 [38] Koene S, Rodenburg RJ, van der Knaap MS, Willemsen MA, Sperl W, Laugel V, Ostergaard E,
 483 Tarnopolsky M, Martin MA, Nesbitt V, Fletcher J, Edvardson S, Procaccio V, Slama A, van den
 484 Heuvel LP, Smeitink JA. Natural disease course and genotype-phenotype correlations in
 485 Complex I deficiency caused by nuclear gene defects: what we learned from 130 cases. J
 486 Inherit Metab Dis 2012;35:737-747.
- 487[39]Flatt JP, Ravussin E, Acheson KJ, Jequier E. Effects of dietary fat on postprandial substrate oxidation488and on carbohydrate and fat balances. J Clin Invest 198 ;76:1019-1024.
- 489 [40] Acheson KJ, Schutz Y, Bessard T, Ravussin E, Jequier E, Flatt JP. Nutritional influences on lipogenesis
 490 and thermogenesis after a carbohydrate meal. Am J Physiol 1984;246: E62-E70.
- 491 [41] de Meer K, Roef MJ, de Klerk JB, Bakker HD, Smit GP, Poll-The BT. Increasing fat in the diet does not
 492 improve muscle performance in patients with mitochondrial myopathy due to complex I
 493 deficiency. J Inherit Metab Dis 2005;28: 95-98.
- 494 [42] Branco AF, Ferreira A, Simoes RF, Magalhaes-Novais S, Zehowski C, Cope E, Silva AM, Pereira D, Sardao
 495 VA, Cunha-Oliviera T. Ketogenic diets : from cancer to mitochondrial diseases and beyond.
 496 Eur J Clin Nutr 2016 (*in press*)

Table 1

Anthropometric characteristics of total included subjects and those undergoing REE and DIT measurements. Results are expressed as mean values \pm SD.

	Group 1 Total included subjects	Group 2 Analysed subjects for REE	Group 3 Analysed subjects for DIT	p-value (group 1 vs 2)	p-value (group 1 vs 3)
n	36	22	12	NA	NA
Female (%)	48	50	60	0.837#	0.916 ^ø
Age : Year	13.2 ± 4.7	13.2 ± 4.6	13.8 ± 4.8	1.000^{s}	0.8238
Weight: kg	36.3 ± 15.8	36.8 ± 15.9	40.2 ± 18.4	0.928§	0.765§
Height: cm	143.8 ±22.1	143.4 ± 20.2	148 ± 21.3	0.918§	0.8118
BMI : <i>Kg/m</i> ²	17.0 ± 4.1	16.9 ± 4.2	17.4 ± 4.6	0.924 [§]	0.875 [§]
Z-score BMI	-0.96 ± 1.6	-0.94 ± 1.68	-0.75 ± 1.59	0.878^{s}	0.749§
Fat mass: %	23.1 ± 12.9	22.22 ± 12.3	22.3 ± 11.5	0.888^{s}	0.902§
Σ FFM : <i>Kg</i>	28.6 ± 11.7	23.3 ± 11.5	32.9 ± 12.2	0.817§	0.396§
^{Σ} Muscle: <i>Kg</i>	26.9 ± 10.3	22.1 ± 12.4	30.8 ± 9.4	0.902§	0.753§

BMI = Body Mass Index NA = Not Applicable NS = Not Significant FFM = Fat Free Mass measured by ^ΣDEXA

 $# = \chi^2 \text{ test} : ^{\varnothing} = \chi^2 \text{ test with Yates correction}$

= Student *t* test

Table 2

Repartition of respiratory chain complex deficiencies in the group of REE analysed subjects along with their main clinical signs (n=22; group 2). Mitochondrial respiratory chain enzyme activities were assessed by biochemical measurements on skeletal muscle biopsy or on cultured skin fibroblasts; diagnoses of MELAS and MNGIE were made on the basis of molecular studies.

Complex deficiency	n	Neurological signs	Muscular signs	Neuro-sensitive signs
Ι	12*	7	1	0
III	2**	2	2	1
IV	6	6	0	0
MELAS ¹	1	1	0	0
MNGIE ²	1	1	0	0
TOTAL	22	16	3	1

¹MELAS = Myoclonic Epilepsy Lactate Acidosis and Stroke like episodes with mutation 3243A>G (mitochondrial DNA).

² MNGIE = MyoNeuroGastro Intestinal Encephalopathy with mutation C130T-G128A.

* 3 of 12 patients are simultaneously deficient for complexes I, II and III.

** 2 patients are simultaneously deficient for complexes III and IV.

Table 3

Comparison between anthropometric characteristics of respiratory chain-deficient subjects with and without complex I deficiencies in the group analysed for REE and body composition. Results are expressed as mean values \pm SD.

	Patients with complex I deficiency	Patients with other defects	p-value
n	12	10	NA
Female (%)	61	56	0.395ø
Age : Year	12.5 ± 4.5	14.3 ± 4.9	0.693§
Weight : kg	39.9 ± 15.9	33.5 ± 15.4	0.376§
Height : cm	142.6 ± 18.3	144.4 ± 23.1	0.927§
Z-score BMI	-0.06 ± 1.6	-2.0 ± 1.11	0.010 §

BMI = Body Mass Index NA = Not Applicable FFM = Fat Free Mass measured by $^{\Sigma}$ DEXA

 $^{\varnothing} = \chi^2$ test with Yates correction

§ = U-Mann Whitney test

Table 4.

Food parameters during cross over periods for REE analysed subjects (n=22 group 2). Results are expressed as mean values \pm SD.

	Cross-over periods			
	Baseline ^a	BD	CD	p-value (BD vs CD)
Energy Intake: Kcal/d	1722.2 ± 366.1	1729.6 ± 401.6	1950.0 ± 593.5	0.089 s
Carbohydrates: %	55.0 ± 4.0	53.8 ± 4.9	43.1 ± 3.1	<0.001 §
Lipids: %	29.3 ± 3.6	31.1 ± 2.9	43.6 ± 2.5	<0.001 §
Proteins: %	15.1 ± 3.4	15.1 ± 3.3	13.3 ± 2.3	0.24 §

^a Baseline is during a basal diet period ^b significant BD = Basal diet : CD = Challenging diet

= Paired student *t* test

Table 5.

Metabolic parameters at rest of analysed subjects (n= 22 group 2) at baseline, after basal diet or after challenging diet. Results are expressed as mean values \pm SD.

	Cross-over periods			
	Baseline	BD	CD	p-value (BD vs CD)
Vo ₂				
Vo2 ml/min	210.5 ± 42.5	205.6 ± 54.2	204.7 ± 43.8	0.904^{s}
Vo2 ml/min/kg of body weight	4.5 ± 2.1	4.8 ± 2.0	4.8 ± 2.1	0.899 §
Vo2 ml/min/kg of FFM	5.4 ± 2.3	6.1 ± 2.3	6.9 ± 4.2	0.775 \$
Vo ₂ <i>ml/min/kg</i> of muscle	5.8 ± 2.2	6.5 ± 2.3	6.5 ± 2.3	0.939 §
VC02				
VCo2 ml/min	243.2 ± 55.5	235 ± 60.1	240.7 ± 51.9	0.842 \$
VCo2 ml/min/kg of body weight	5.3 ± 2.3	4.5 ± 1.9	4.4 ± 2.2	0.810 \$
VCo2 ml/min/kg of FFM	6.4 ± 2.4	5.6 ± 2.0	6.2 ± 3.8	0.436 \$
VCo2 ml/min/kg of muscle	6.9 ± 2.3	6.0 ± 2.1	5.9 ± 2.4	0.332s
REE				
REE Kcal/d	1229.6 ± 303.8	1148.8 ± 301.7	1156.1 ± 278.8	0.942 \$
Σ pREE Kcal/d	1192.4 ± 232.4	1191.0 ± 246.7	1200 ± 245.2	0.934 §
REE Kcal/d/kg of body weight	38.1 ± 13.7	34.7 ± 11.8	35.7 ± 15.1	0.720 \$
REE Kcal/d/kg of FFM	46.4 ± 13.7	43.3 ± 12.3	50.2 ± 28.9	0.624^{s}
REE Kcal/d/kg of muscle	49.1 ± 13.0	49.9 ± 12.4	48.1 ± 16.2	0.834 \$
RO				
mRO	0.86 ± 0.10	0.87 ± 0.08	0.85 ± 0.10	0.834 \$
pRQ	0.87 ± 0.11	0.88 ± 0.10	0.84 ± 0.10	0.729 \$

^a Baseline is during a basal diet period

^b significant

BD = Basal diet: CD = Challenging diet

REE = Resting Energy Expenditure

pREE = Predictive Resting Energy Expenditure

FFM = Fat Free Mass

mRQ = Measured Respiratory Quotient (measured VO₂/measured VCO₂)

pRQ = Predictive Respiratory Quotient (theoretically calculated from the nutritional contents of diets in each class of energetic substrates

[carbohydrates, lipids and proteins])

\$ = Paired student *t* test

= Paired student *t* test

 Σ Predictice REE calculated by Harris and Benedict equation

Table 6.

Body composition parameters of subjects (n= 22) undergoing REE evaluations at baseline, after one month of basal diet and after one month of challenging diet. Results are expressed as mean values \pm SD.

		Cross-over periods		
	Baseline	BD	CD	p-value (BD vs CD)
Weight : Kg	38.8 ± 15.9	36.6 ± 13.7	38.5 ± 18.0	0.698 \$
Fat mass : %	22.1 ± 12.3	21.9 ± 13.0	21.6 ± 13.3	0.810 s
FFM : <i>Kg</i>	29.3 ± 11.5	29.1 ± 12.2	26.8 ± 11.0	0.716 §
Muscle: Kg	26.9 ± 10.3	26.6 ± 11.1	26.5 ± 11.0	0.810 §

FFM = Fat Free Mass § = Paired student *t* test

^a Baseline is during a basal diet period

^b significant

BD = Basal diet : CD = Challenging diet

Table 7.

Minimum set of metabolic and anthropometrics data in complex I deficient subjects (n= 12) undergoing REE evaluations at baseline, after one month of basal diet and after one month of challenging diet. Results are expressed as mean values \pm SD.

Cross-over periods				
	Baseline	BD	CD	p-value (BD <i>vs</i> CD)
REE Kcal/d RQ Weight: Kg Fat mass: % FFM: Kg Muscle: Kg	$1229.6 \pm 258.2 \\ 0.75 \pm 0.12 \\ 39.9 \pm 15.9 \\ 27.9 \pm 14.5 \\ 29.8 \pm 8.9 \\ 26.9 \pm 8.2$	1139.5 ± 250.3 0.88 ± 0.14 40.2 ± 16.2 25.1 ± 15.9 28.9 ± 9.8 26.5 ± 9.3	1158.2 ± 255.5 0.82 ± 0.08 40.6 ± 16.1 26.1 ± 16.5 28.3 ± 9.8 25.8 ± 9.1	0.695 \$ 0.366 \$ 0.948 \$ 0.790 \$ 0.657 \$ 0.929 \$

FFM = Fat Free Mass

§ = Wilcoxon test

^a Baseline is during a basal diet period ^b significant

BD = Basal diet; CD = Challenging diet

Table 8.

Minimum set of metabolic and anthropometrics data of subjects with a respiratory chain defect other than complex I (n=10) and undergoing REE evaluations at baseline, after one month of basal diet and after one month of challenging diet. Results are expressed as mean values \pm SD.

	Cross-over periods			
	Baseline	BD	CD	p-value (BD vs CD)
REE Kcal/d	1195.1 ± 361.2	1166.5 ± 378.5	1157.0 ± 316.3	0.875 \$
RQ	0.80 ± 0.15	0.81 ± 0.16	0.89 ± 0.12	0.555 \$
Weight: Kg	33.5 ± 15.4	34.5 ± 15.5	34.5 ± 15.4	0.948 §
Fat mass: %	15.8 ± 4.6	17.5 ± 6.4	16.6 ± 6.1	0.109 \$
FFM: Kg	28.7 ± 14.3	29.2 ± 15.5	25.2 ± 12.6	0.248 §
Muscle: Kg	26.7 ± 12.7	26.8 ± 13.7	27.3 ± 13.2	0.185 \$

FFM = Fat Free Mass

§ = Wilcoxon test

^a Baseline is during a basal diet period

^b significant BD = Basal diet: CD = Challenging diet

Table 9.

Food parameters during cross over periods for DIT assessment (n = 12; group 3). Results are expressed as mean values \pm SD.

	Cross-ov		
	BD	CD	p-value (BD vs CD)
Energy Intake*: Kcal	161.0 ± 12.3	219.9 ± 13.0	0.809§
Carbohydrates: %	61.1 ± 6.6	45.5 ± 12.2	0.403§
Lipids: %	28.5 ± 10.3	42.3 ± 11.1	0.878^{s}
Proteins: %	10.4 ± 6.4	12.2 ± 5.9	0.911

*Energy intake measurement at breakfast for DIT § = Wilcoxon test BD = Basal diet

CD = Challenging diet

Figure 1 : Flow CONSORT diagram of screening, inclusion, randomisation and analysis rate in the study.

Figure 2 : Randomized cross-over design of the study.

Figure 3 : Evolution of diet induced thermogenesis (only group 3 ; n=12)

Figure 4 : Evolution of RQ during diet induced thermogenesis assessment (only group 3 ; n=12)



REE = Resting Energy Expenditure DIT = Diet Induced Thermogenesis

Figure 1 : Flow CONSORT diagram of screening, inclusion, randomisation and analysis rate in the study.



BD = Basal Diet CD = Challenging Diet

Figure 2 : Randomized cross-over design of the study.



Figure 3 : Evolution of diet induced thermogenesis (only Group 3 ; n=12)



Figure 4 : Evolution of RQ during diet induced thermogenesis assessment (only group 3 ; n=12)