



HAL
open science

The Expression of the Short Isoform of Thymic Stromal Lymphopoietin in the Colon Is Regulated by the Nuclear Receptor Peroxisome Proliferator Activated Receptor-Gamma and Is Impaired during Ulcerative Colitis

Anthony Martin Mena, Audrey Langlois, Silvia Speca, Lucil Schneider, Pierre Desreumaux, Laurent Dubuquoy, Benjamin Bertin

► **To cite this version:**

Anthony Martin Mena, Audrey Langlois, Silvia Speca, Lucil Schneider, Pierre Desreumaux, et al.. The Expression of the Short Isoform of Thymic Stromal Lymphopoietin in the Colon Is Regulated by the Nuclear Receptor Peroxisome Proliferator Activated Receptor-Gamma and Is Impaired during Ulcerative Colitis. *Frontiers in Immunology*, 2017, 8, 10.3389/fimmu.2017.01052 . hal-03113349

HAL Id: hal-03113349

<https://hal.univ-lille.fr/hal-03113349>

Submitted on 18 Jan 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



The Expression of the Short Isoform of Thymic Stromal Lymphopoietin in the Colon Is Regulated by the Nuclear Receptor Peroxisome Proliferator Activated Receptor-Gamma and Is Impaired during Ulcerative Colitis

Anthony Martin Mena^{1,2}, Audrey Langlois^{1,2}, Silvia Specca^{1,2}, Lucil Schneider⁴, Pierre Desreumaux^{1,2,3}, Laurent Dubuquoy^{1,2} and Benjamin Bertin^{1,2*}

¹ Univ. Lille, U995 – LIRIC – Lille Inflammation Research International Center, Lille, France, ² INSERM, U995, Lille, France, ³ CHU Lille, Service des Maladies de l'Appareil Digestif et de la Nutrition, Hôpital Claude Huriez, Lille, France, ⁴ CHU Lille, Service de Chirurgie Digestive et Transplantations, Hôpital Claude Huriez, Lille, France

OPEN ACCESS

Edited by:

Eric Cox,
Ghent University, Belgium

Reviewed by:

Luca Pastorelli,
Università degli Studi di Milano, Italy
Bruce Vallance,
University of British Columbia,
Canada

*Correspondence:

Benjamin Bertin
benjamin.bertin-2@univ-lille2.fr

Specialty section:

This article was submitted
to Mucosal Immunity,
a section of the journal
Frontiers in Immunology

Received: 08 June 2017

Accepted: 14 August 2017

Published: 04 September 2017

Citation:

Martin Mena A, Langlois A, Specca S, Schneider L, Desreumaux P, Dubuquoy L and Bertin B (2017) The Expression of the Short Isoform of Thymic Stromal Lymphopoietin in the Colon Is Regulated by the Nuclear Receptor Peroxisome Proliferator Activated Receptor-Gamma and Is Impaired during Ulcerative Colitis. *Front. Immunol.* 8:1052. doi: 10.3389/fimmu.2017.01052

The etiology of inflammatory bowel diseases remains largely unknown. We previously demonstrated that the expression of the peroxisome proliferator activated receptor-gamma (PPAR γ) is downregulated in colonic epithelial cells of patients with ulcerative colitis (UC). PPAR γ is a nuclear receptor that modulates inflammation. We hypothesized that its deficiency may play a role in the loss of intestinal homeostasis through the control of immunomodulatory factors. We found that thymic stromal lymphopoietin (TSLP), an epithelial cytokine with pleiotropic functions, is regulated by PPAR γ . While this cytokine possesses two isoforms, only the short form (sTSLP) was regulated by PPAR γ . sTSLP mRNA expression was decreased both in PPAR γ knock-down Caco2 cells and cells treated with PPAR γ antagonist, whereas PPAR γ agonists induced the expression of sTSLP in Caco2 and T-84 cells. The response element activated by PPAR γ was identified in the promoter of the sTSLP gene by chromatin immunoprecipitation and gene reporter assays. The expression of sTSLP was significantly decreased in the colonic mucosa of UC patients compared to controls and was correlated with PPAR γ expression. Our results identified sTSLP as a new PPAR γ -target gene and support the hypothesis that, in UC, PPAR γ deficiency in colonic mucosa could play a role in the loss of intestinal tolerance through an impaired sTSLP expression.

Keywords: PPARgamma, thymic stromal lymphopoietin, intestinal epithelial cells, ulcerative colitis, colon

INTRODUCTION

In humans, the intestinal mucosal surface is an essential exchange area between the host and its environment. In the context of intestinal homeostasis, mucosal immune cells must be able to tolerate commensal and food antigens while keeping their ability to fight pathogens (1). A fundamental actor in this dialog between luminal antigens and mucosal leukocytes is the intestinal epithelial

cell (IEC) (2). Although initially considered as a simple physical barrier, it is now clear that IECs actively participate to the induction and maintenance of intestinal homeostasis. For instance, IECs respond to microorganism antigens or metabolites by increasing the synthesis of mucus, the secretion of anti-microbial peptides and/or by promoting immunoglobulin A transcytosis (2). In addition, IECs have the ability to secrete cytokines and chemokines at their basal pole, thereby regulating the functions of the underlying intestinal immune cells (2). Notably, it has been demonstrated that, through the secretion of immunoregulatory molecules such as transforming growth factor- β (TGF- β), thymic stromal lymphopoietin (TSLP), or retinoic acid, IECs have the capacity to regulate the ability of dendritic cells to induce regulatory T cells (3–5).

Recent studies suggest that IEC dysfunctions may actively participate in the pathogenesis of ulcerative colitis (UC) (6, 7). UC (along with Crohn's disease) is one of the major types of inflammatory bowel diseases (IBD) which are chronic, complex, relapsing immune-mediated diseases of the gastrointestinal tract of unknown etiology (7, 8). Our previous work has demonstrated that the expression of peroxisome proliferator activated receptor-gamma (PPAR γ) is reduced in the colonic epithelial cells (CEC) of individuals with UC (9, 10). PPAR γ is a nuclear receptor which plays a key role in lipid and glucose metabolisms as well as in gut homeostasis (11, 12). Conditional knock-out mice in which PPAR γ expression is selectively ablated in CECs demonstrate an increased susceptibility to DSS-induced colitis (13). Conversely, PPAR γ synthetic agonists inhibit inflammation and reduce disease severity both in experimental models of colitis and in UC patients (12, 14–16). These results clearly underline the importance of CEC in controlling the magnitude of the inflammatory response in the gut, at least in part, in a PPAR γ -dependent manner. Despite these observations, as well as the abundance of PPAR γ in the colon (17), how this nuclear receptor exerts its anti-inflammatory functions within the intestine is poorly known and most of its target genes, notably in CEC, remain to be characterized.

Our hypothesis is that PPAR γ deficiency in CEC during UC might play a role in the loss of intestinal homeostasis through the regulation of immunoregulatory factors. One possibility would be the potential role of PPAR γ in the control of cytokine production by CEC. In addition, CEC express some cytokine receptors and, consequently, are able to respond to the cytokine environment within the intestinal mucosa (18, 19). The aim of this study was therefore to identify PPAR γ target genes involved in CEC immune functions relevant to UC pathophysiology. Our results identified the gene encoding the short isoform of TSLP (sfTSLP) as a new gene regulated by PPAR γ in CEC and support the hypothesis that, in UC, the impaired expression of PPAR γ in colonic mucosa could play a role in the loss of intestinal tolerance through a decrease of sfTSLP expression.

MATERIALS AND METHODS

Cell Culture and Stimulation

Caco-2 cells (ATCC[®] CRL-2102[™]) were grown in Dulbecco's Modified Eagle's Medium (DMEM, Invitrogen, Life Technologies, Cergy-Pontoise, France) supplemented with 20% fetal calf serum

(FCS, Dutscher, Brumath, France), 1% penicillin-streptomycin (Invitrogen, Life technologies), and 1% non-essential amino acids (Invitrogen, Life technologies). T84 cells (ATCC[®] CCL-248[™]) were grown in a 1:1 mixture of Ham's F12 medium and DMEM supplemented with 5% FCS and 1% penicillin-streptomycin. H292 cells (ATCC[®] CRL-1848[™]) were grown in RPMI-1640 supplemented with 10% FCS and 1% penicillin-streptomycin. Generation and maintenance of PPAR γ knock-down Caco-2 cells were described in Ref. (10). All cell lines were cultured as confluent monolayers at 37°C in a controlled, 5% CO₂ atmosphere.

For cell stimulations, 1×10^6 cells per well were seeded in 6-well plates. Serum deprivation was used 16 h prior to stimulation in order to synchronize the cells. Cells were treated with GED (30 mM; Nogra Pharma Ltd, Italy), pioglitazone (10 μ M; Sigma-Aldrich), 5-ASA (30 mM; Sigma-Aldrich), GW9662 (10 μ M, Sigma-Aldrich), or poly I:C (10 μ g/L; Invivogen). When necessary, the DMSO vehicle (Sigma-Aldrich) was used as control. After 24 h of stimulation, cells were washed twice with sterile PBS and were lysed with 1 ml of TRIzol[®] reagent (Invitrogen[™], Cergy Pontoise, France). The plates were frozen at -80°C for subsequent RNA and protein extraction. Cell stimulations were performed in 3 or 6 replicates.

Culture of polarized Caco-2 cells was done as follow: 3×10^4 cells were seeded on Transwell[®] filters in 12-well plates. During the first 7 days, complete culture medium (DMEM containing FCS) was added both to the upper and lower compartment. Cell polarization is obtained between days 8 and 21 of culture by culturing cells with DMEM containing FCS in the lower compartment and DMEM without FCS in the upper compartment. Measurement of the Trans-Epithelial Resistance (TER) was done every 2 days (before medium replacement) to check cell differentiation and integrity of the cellular barriers. At day 21, polarized Caco-2 cells were stimulated with PPAR γ agonist. After 24 h of stimulation, cells were washed twice with sterile PBS and were lysed with 1 ml of TRIzol[®] reagent.

RNA Extraction

RNAs were isolated with the TRIzol[®] reagent according to the manufacturer's instructions and resuspended in RNase-free, DEPC-water. The purity and concentration of the RNAs were evaluated by UV spectroscopy on a Nanodrop system (Nyxor Biotech, Paris, France).

Quantitative PCR (qPCR)

Expression of genes was quantified by qPCR of corresponding reverse transcribed mRNA. 1 μ g of total RNA was reverse-transcribed into cDNA using the High Capacity cDNA Archive kit (Applied Biosystems). Amplification was performed using an ABI PRISM StepOnePlus detection system (Applied Biosystem) using Power SYBR[®] Green PCR master Mix (Applied Biosystem). Primer pairs were chosen with qPrimer depot software.¹ See Table S1 in Supplementary Material for the oligonucleotides used in this study. Quantification of qPCR signals was performed using ΔCt relative quantification method using GAPDH as a reference gene.

¹<http://primerdepot.nci.nih.gov>.

Values were represented in terms of relative quantity of mRNA level variation or fold increase compared to control conditions.

Protein Analysis

Peroxisome proliferator activated receptor-gamma protein was detected from total proteins obtained after TRIzol extractions. Proteins were separated by polyacrylamide gel electrophoresis and electroblotted on nitrocellulose membrane. PPAR γ was detected using a monoclonal rabbit antibody (C26H12) as primary antibody (Cell Signaling Technology[®]) and goat anti-rabbit IgG peroxidase conjugate (Jackson ImmunoResearch). Nitrocellulose membranes were probed with a mouse monoclonal anti- β actin antibody (Sigma-Aldrich[®], France) to assess equal protein loading.

Short form TSLP protein was detected by immunoprecipitation followed by western-blot analysis. 6 million of Caco-2 cells were used for each immunoprecipitation. When indicated, cells were stimulated during 24 h in 100 mm cell culture dishes. Cells were washed and scraped in PBS buffer, then lysed in IPH buffer (50 mM Tris pH 8, 150 mM NaCl, 5 mM EDTA, 0.5% NP40, and protease inhibitors) for 1 h in ice. After centrifugation 15 min at 15,000 g, the supernatants were recovered and dosed. The same amount of protein was engaged in each immunoprecipitation (1.5 mg). 2 μ g of TSLP antibody (Clone # 184638 rabbit polyclonal, Abcam) were added and the tubes were incubated overnight at 4°C under rotation. The next day, a mixture of protein A/G agarose beads (Santa Cruz) was added and incubation was continued 2 h at 4°C under rotation. Then, beads were washed 5 times with IPH buffer followed by denaturation in Laemli sample buffer at 95°C for 5 min. Immunoprecipitated material was analyzed by western-blot using the TSLP antibody (Clone # 184638 rabbit polyclonal, Abcam) as primary antibody and goat anti-rabbit IgG peroxidase conjugate (Jackson ImmunoResearch). β -actin protein was detected to assess equal protein engagement (1% input).

In Silico Analysis of sTSLP Promoter

Presence of PPAR response element sequences were investigated *in silico* in the promoter of the sTSLP. 1000 base pairs upstream of the transcription initiation site were analyzed by two different softwares: NUBIScan² (20) and PPRE Search³ (21).

Chromatin Immunoprecipitation Experiments

The physical binding of PPAR γ onto the sTSLP gene promoter was studied by ChIP experiments in Caco-2 cells (10×10^6 cells) stimulated for 24 h with 30 mM GED in 150 mm cell culture petri dishes. Caco-2 cells were synchronized by the addition of serum-free medium for 16 h and then stimulated for 24 h using the protocol described previously. Cells were then rinsed with PBS and the protein-DNA complex was fixed by adding 10% PFA for 30 min at room temperature. This binding was stopped by the addition of glycine (0.125 M). Cells were collected by scrapping in cold PBS and protease inhibitors (Sigma). The cell pellet obtained

by centrifugation was taken up in 300 μ l SDS buffer (1% SDS, 10 mM EDTA, 50 mM Tris-HCl pH 8, protease inhibitors + PMSF 0.1 mM) and sonicated (Diagenode, BioruptorUCD200TM-EX) for 30 s, followed by 30 s' resting time. For each immunoprecipitation, 125 μ l of crosslinked sonicated sample was diluted with 225 μ l of IP buffer (1% triton X-100, 150 mM NaCl, 2 mM EDTA, 20 mM Tris-HCl pH 8.1, and protease inhibitors) and precleared for 4 h by adding 40 μ l of protein A/G beads (50% slurry protein A/G Sepharose, Clinisciences) and 5 μ g of salmon sperm DNA (Invitrogen). Complexes were immunoprecipitated with specific anti-PPAR γ antibodies (C26H12 rabbit monoclonal antibody, Cell Signaling) by incubation overnight at 4°C under rotation. Immune complexes were recovered by adding 40 μ l of protein A/G Sepharose (50%) plus 2 μ g salmon sperm DNA and incubated for 4 h at 4°C. The beads were washed twice in wash buffer 1 (0.1% SDS, 1% Triton X-100, 150 mM NaCl, 0.1% Deoxycholate, 1 mM EGTA, 2 mM EDTA, 20 mM Tris-HCl pH 8.0), twice in wash buffer 2 (0.1% SDS, 1% Triton X-100, 500 mM NaCl, 0.1% Deoxycholate, 1 mM EGTA, 2 mM EDTA, 20 mM Tris-HCl pH 8.0), once in wash buffer 3 (0.25 mM LiCl, 0.5% Deoxycholate, 0.5% NP40, 0.5 mM EGTA, 1 mM EDTA, 10 mM Tris HCl pH 8.0), and 3 times in wash buffer 4 (1 mM EDTA, 10 mM Tris-HCl pH 8.0). The co-immunoprecipitated DNA was then extracted with 150 μ l of extraction buffer (0.1 M NaHCO₃, 1% SDS). Cross-linking was reverse overnight at 65°C. DNA was then purified using the PCR Clean-up kit (Macherey-Nagel) and analyzed by PCR.

Cloning

Genomic DNA fragments were cloned upstream of the luciferase reporter gene (plasmid pGL4.10 [Luc2]; Promega). The fragment corresponding to the 1,000 bp upstream of the transcription initiation site of the human *sftslp* gene (construction "Prom TSLP2") was obtained by PCR from genomic DNA of Caco-2 cells (sense oligonucleotide 5'-CGCTCGAGGGATGTCTATCCTTTGCTAAAG-3'; antisense oligonucleotide 5'-CGAAGCTTGGCGGAGGGCAC TCGTCGCGAAAAG-3'). The fragment corresponding to the 548 base pairs upstream of the transcription initiation site of the human *sftslp* gene (construct "Prom TSLP2 Δ 1") was obtained by PCR from genomic DNA of Caco-2 cells (sense oligonucleotide 5'-CGCTCGAGGGTACCTGAGTGGAAACTGTT-3'; antisense oligonucleotide 5'-CGAAGCTTGGCGGAGGGCAC TCGTCGCGAAAAG-3'). The PCR products were cloned into a TOPO pCR4 vector (TOPO TA cloning, Invitrogen) and then sequenced in order to check for potential *Taq* Polymerase errors. For each construct, a mutation-free fragment was subcloned into the vector pGL4.10 [Luc2] (Promega) using the *XhoI/HindIII* restriction sites introduced into the oligonucleotides (underlined in the sequence).

Transient Transfections and Reporter Gene Assays

Reporter plasmids and the empty vector control were transiently transfected in Caco-2 cells using Nucleofector[™] Technology. For each condition, 2×10^6 cells (50–70% confluency) were transfected with 2 μ g of plasmids using SE 4D-Nucleofector solution and program DS-123. Transfected cells were seeded in 48-well

²<http://www.nubiscan.unibas.ch/>.

³<http://www.classicus.com/PPRE/>.

plates at 100,000 cells per well and treated by PPAR γ agonists for 24 h. Luciferase activity was measured using the luciferase assay kit (Promega) in fluorometer (FLUOstar Omega, BMG Labtech) according to the manufacturers' instructions. Total protein contents were dosed using Bradford reagent (Euromedex) and used to normalize luciferase activity.

Patients and Tissue

A local ethics committee (Comité de Protection des Personnes Nord Ouest IV, CHRU Lille, France) approved the study and all subjects gave informed consent (No. DC-2008-642). Surgical colonic samples were obtained from patients with an established diagnosis of UC according to international criteria and from control patients (patients underwent surgery for colorectal cancer and patients for diverticulitis of the sigmoid). The detailed characteristics of patient cohort are given in Table S2 in Supplementary Material. Colonic epithelial cells were isolated from resected colonic specimens as previously described (10).

Statistics

The data are presented as mean with SEM or SD. All graphs were plotted and analyzed with GraphPad Prism 5 Software (San Diego, CA, USA) using a two-tailed non-parametric Mann-Whitney test. *P* values < 0.05 were considered statistically significant.

RESULTS

Reduced Expression of the Gene Encoding TSLP in PPAR γ Knock-Down Cells

In order to identify factors whose expression depends on PPAR γ in the CEC and which might be involved in the immunoregulatory properties of these cells, we have used a targeted approach and first investigated the mRNA expression of some genes encoding cytokines and cytokine receptors in PPAR γ knock-down colonic epithelial Caco-2 cells. To this end, we used a Caco-2 ShPPAR γ cell line that stably expresses a short hairpin anti-sense RNA against PPAR γ , leading to specific downregulation of PPAR γ (10) (Figure 1A). Compared to control cells (Caco-2 ShLuc cells, expressing a control shRNA directed against the luciferase gene), the expression of PPAR γ in Caco-2 ShPPAR γ cell line was reduced approximately by 70% and this decrease was equivalent to those we observed in CEC from UC patients (9, 10).

Thus, using this system, we observed that the expression of some pro-inflammatory genes such as interleukin-1 β or MCP-1 was significantly upregulated in Caco-2 ShPPAR γ cells (Figure 1B). More remarkably, we observed a dramatic decrease of expression of the mRNA encoding TSLP cytokine (Figure 1B). To confirm this result, we measured the expression level of TSLP mRNA in independent experiments and systematically found that TSLP mRNA level was significantly reduced by about 90% in ShPPAR γ Caco-2 cells compared to the ShLuc cells (Figure 1C). In order to exclude the possibility of off-target effects of anti-sense RNA, we investigated the effect of the PPAR γ antagonist GW9662. We observed that GW9662 also significantly

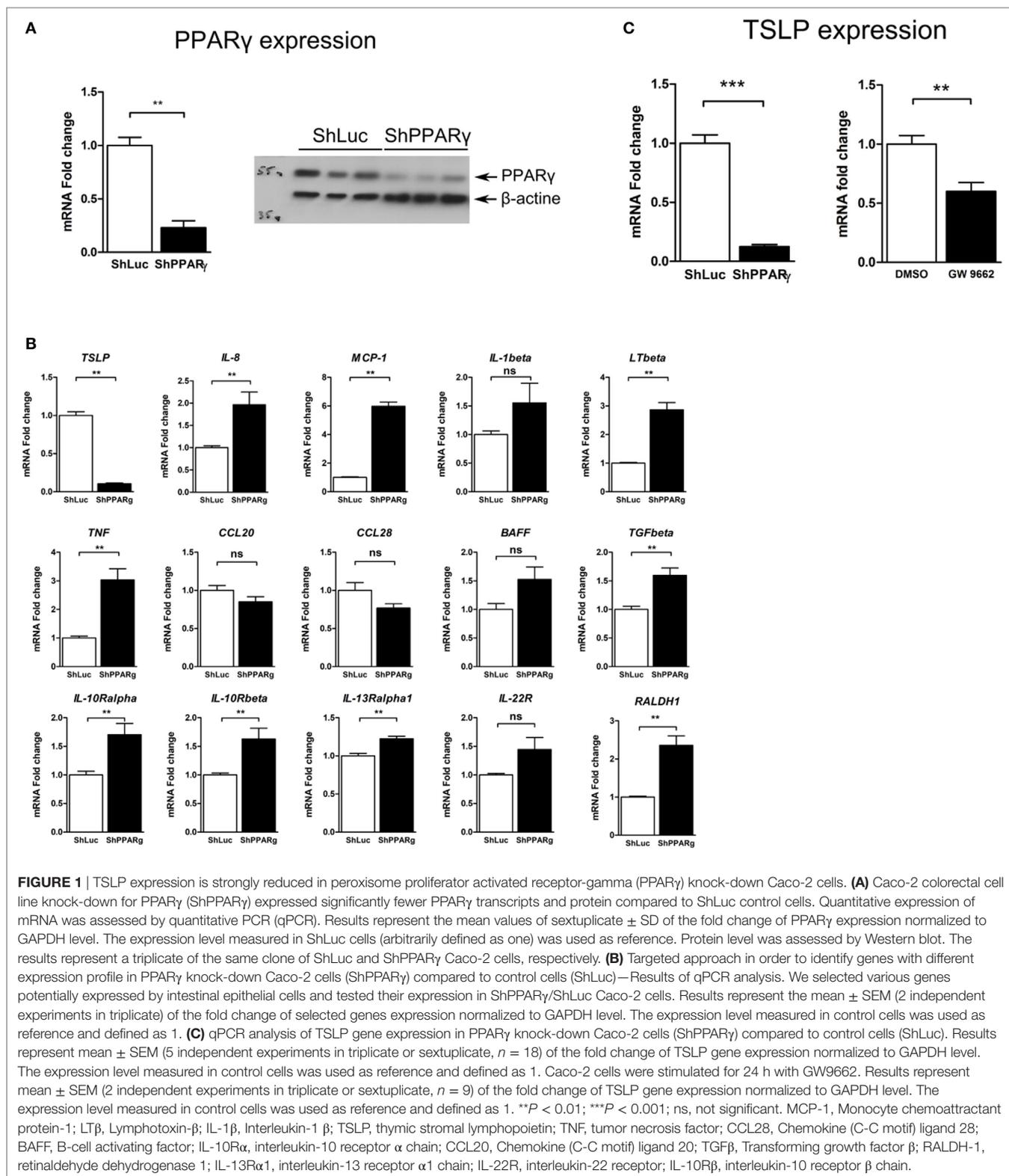
suppressed TSLP expression in Caco-2 cells (Figure 1C). These first results demonstrate that TSLP expression is abrogated in Caco-2 cells when PPAR γ is inhibited.

Activation of PPAR γ -Induced TSLP Expression

To further explore the regulation of TSLP expression by PPAR γ in CECs, we measured TSLP mRNA expression after stimulation of Caco-2 cells with 3 different PPAR γ agonists: pioglitazone (Pio; 10 μ M), a well characterized PPAR γ ligand belonging to the anti-diabetic thiazolidinedione drug class (22), 5-amino salicylic acid (5-ASA; 30 mM) (23), and a new PPAR γ modulator, we developed and named GED-0507-34-Levo (GED; 30 mM) (24). All PPAR γ ligands tested were able to significantly induce the expression of TSLP mRNA in Caco-2 cells (Figure 2A). These results were confirmed in another IEC line (T84), which expresses both TSLP and PPAR γ (Figure 2B). In order to confirm the involvement of PPAR γ in this process, we next assessed the induction of TSLP mRNA expression by PPAR γ ligands in PPAR γ knock-down Caco-2 cells. We first observed that 5-ASA and GED significantly induce TSLP mRNA expression in Caco-2 ShLuc control cells in an identical manner than in normal Caco-2 cells (Figure 2C), while in ShPPAR γ Caco-2 cells, both 5-ASA and GED-dependent expression of TSLP mRNA were strongly reduced (Figure 2C). This result shows that PPAR γ potentiates *tslp* expression in Caco-2 cells. Taken together, our results support the hypothesis that PPAR γ is a key factor controlling TSLP expression in CEC.

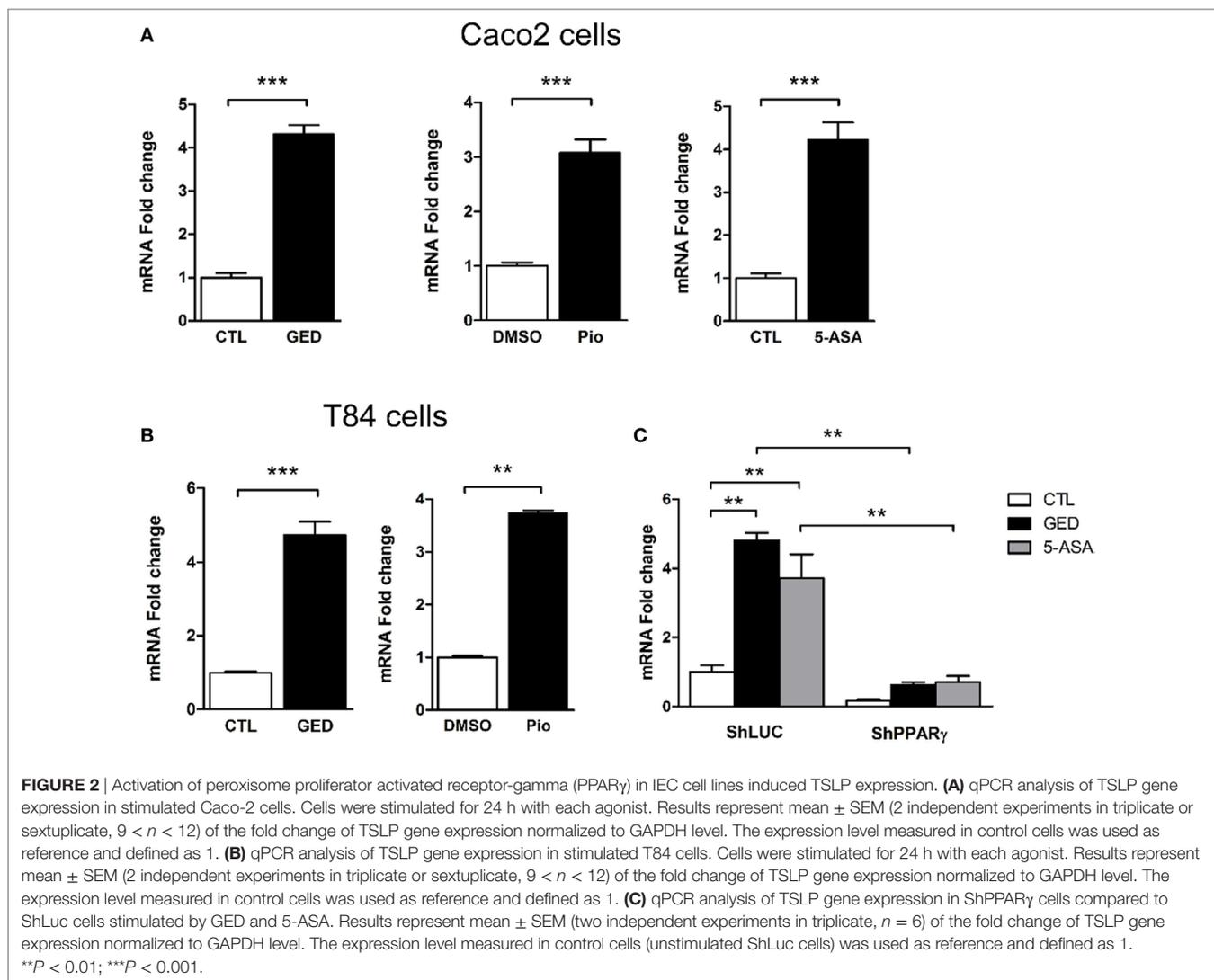
PPAR γ Regulates the Expression of the sTSLP

Three distinct mRNA variants can be transcribed from the human *tslp* gene, but only two encode functional protein. Variant 1 is composed of 4 exons which encode the long form TSLP (lTSLP) of 159 amino acids length. Variant 2 is composed of 2 exons which encode the sTSLP consisting in the 63 and/or 60 C terminal amino acids of long TSLP (Figure 3A). It is only very recently that studies have been interested in distinguishing these 2 isoforms and their possible distinct functions (25–27). It appears that lTSLP mRNA and protein are upregulated in response to inflammatory stimuli and therefore associated to inflammatory conditions such as asthma, dermatitis, or IBD (26). Conversely, sTSLP is constitutively expressed in healthy mucosa of the digestive tract as well as in the skin (25, 26). The cues that drive this distinct expression pattern are unknown. To test the implication of PPAR γ , we designed isoform-specific primers (Figure 3A; Table S1 in Supplementary Material for primer sequences) and assessed the transcription of the two TSLP variants. lTSLP transcript was amplified from bronchial epithelial cells (H292) stimulated with polyinosinic-polycytidylic acid (polyI:C) (Figure S1 in Supplementary Material), but no lTSLP transcript was detected in our quantitative RT-PCR experiments in Caco-2 and T84 cells, both at the basal level nor after stimulation with PPAR γ agonists. On the other hand, the three PPAR γ agonists were all able to significantly induce sTSLP mRNA expression, both in Caco-2 and T84 cells (Figures 3B,C). The Caco-2 cells have the



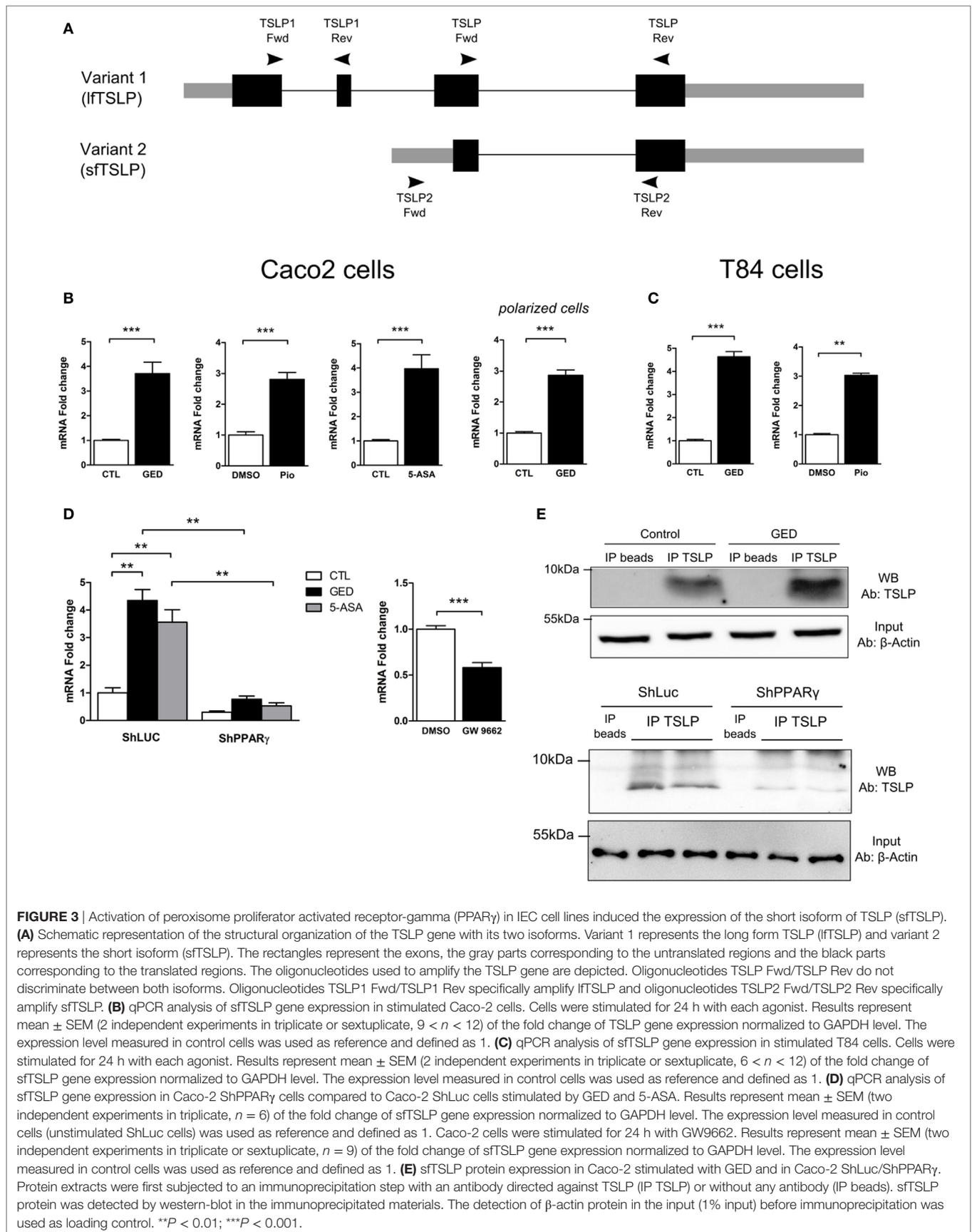
ability to spontaneously differentiate (when reaching confluence) into polarized IECs which express, for instance, tight junction proteins and develop transepithelial electric resistance. In order to test whether the induction of sTSLP by PPAR γ agonist was

maintained or modified by Caco-2 cells differentiation, we also assessed the induction of sTSLP by GED in polarized Caco-2 cells. To this end, Caco-2 cells were grown on transwell filters during 21 days to allow cell polarization, and then cells were stimulated



with PPAR γ agonist for 24 h. In these conditions, activation of PPAR γ with GED still resulted in a significant three-fold induction of sTSLP mRNA expression (**Figure 3B**). We next investigated the stimulation of sTSLP mRNA expression by PPAR γ ligands in PPAR γ knock-down Caco-2 cells. We found that 5-ASA and GED significantly induce sTSLP mRNA expression in Caco-2 ShLuc control cells (to the same extent to what we observed in **Figure 2C** with non-isoform specific primers), while in ShPPAR γ Caco-2 cells, both 5-ASA and GED-dependent expression of sTSLP mRNA were strongly compromised (**Figure 3D**). Treatment of Caco-2 cells with the PPAR γ antagonist GW9662, confirmed that blocking PPAR γ activity significantly impact the expression of sTSLP (**Figure 3D**). Finally, we tried to confirm these results by detecting and evaluating sTSLP protein level in stimulated Caco-2 cells. Given that the amino acid sequence of sTSLP perfectly overlap the C-terminal sequence of lTSLP, it is not possible to have an antibody specifically directed against sTSLP. However, by using an antibody raised against the C-terminal part common to both proteins, it should be possible to distinguish the two isoforms in western-blot analysis due to their different

molecular weights [apparent molecular weight of 23 kDa for lTSLP and 9 kDa for sTSLP (25)]. Nevertheless, we never succeeded to detect sTSLP in protein extracts from Caco-2 cells (stimulated or not) directly in western-blot analysis (not shown). We hypothesized that it could be due to a low level of sTSLP protein expression and, thus, in order to increase TSLP protein concentration, we first performed an immunoprecipitation step before western-blot analysis. This approach allowed us to detect sTSLP protein in Caco-2 stimulated with GED as well as in Caco-2 ShLuc/ShPPAR γ (**Figure 3E**). No signal was observed in the lanes corresponding to the immunoprecipitation control (**Figure 3E** “IP beads,” which correspond to the immunoprecipitation of protein extracts without any anti-TSLP antibody), whereas, we detected a band migrating just below 10 kDa in the lane corresponding to the proteins immunoprecipitated with anti-TSLP antibody (**Figure 3E**, “IP TSLP”). We concluded that this band corresponds to sTSLP protein. Moreover, we observed a stronger signal in the lane corresponding to proteins immunoprecipitated from Caco-2 cells stimulated with GED (**Figure 3E**, GED lane “IP TSLP”) compared to unstimulated Caco-2 cells (**Figure 3E**,



control lane “IP TSLP”) and fewer sFTSLP were immunoprecipitated from ShPPAR γ Caco-2 cells than from ShLuc Caco-2 cells (Figure 3E, ShPPAR γ “IP TSLP” lanes compared to ShLuc “IP TSLP” lanes). Given that equal amount of protein was engaged in each immunoprecipitation, we concluded from this experiment that modulation of PPAR γ in Caco-2 cells impacts sFTSLP protein expression. Taken together, the results of this part strongly suggest that PPAR γ is an important transcription factor controlling sFTSLP expression in CEC.

PPAR γ Functionally Binds to the Promoter of sFTSLP Gene

To further confirm the regulation of sFTSLP expression by PPAR γ in CECs, we next investigated the presence of PPAR response element (PPRE) sequences in the sFTSLP gene promoter. *In*

silico analysis of the 1,000 base pairs (bp) upstream from the transcription start site of the human sFTSLP gene revealed the presence of several potential PPRE (DR1, DR2, and DR3) by which PPAR γ may regulate sFTSLP gene expression (Figure 4A; Figure S2 in Supplementary Material). Chromatin immunoprecipitation analysis of these putative PPRE revealed notably that a DR3 located between -785 (bp) and -770 bp upstream of the transcription start site was bound by PPAR γ within the sFTSLP gene promoter in Caco-2 cells (both in control and stimulated cells) (Figure 4B). In order to test whether this binding was functional, we constructed two reporter plasmids: in the first one, the genomic fragment corresponding to the 1,000 bp upstream from the transcription start site of the human sFTSLP gene was cloned upstream to the luciferase reporter gene into a pGL4 vector (PromTSLP2-Luc construct; Figure 4C). In the second one, the genomic region containing the putative PPRE DR3 bound

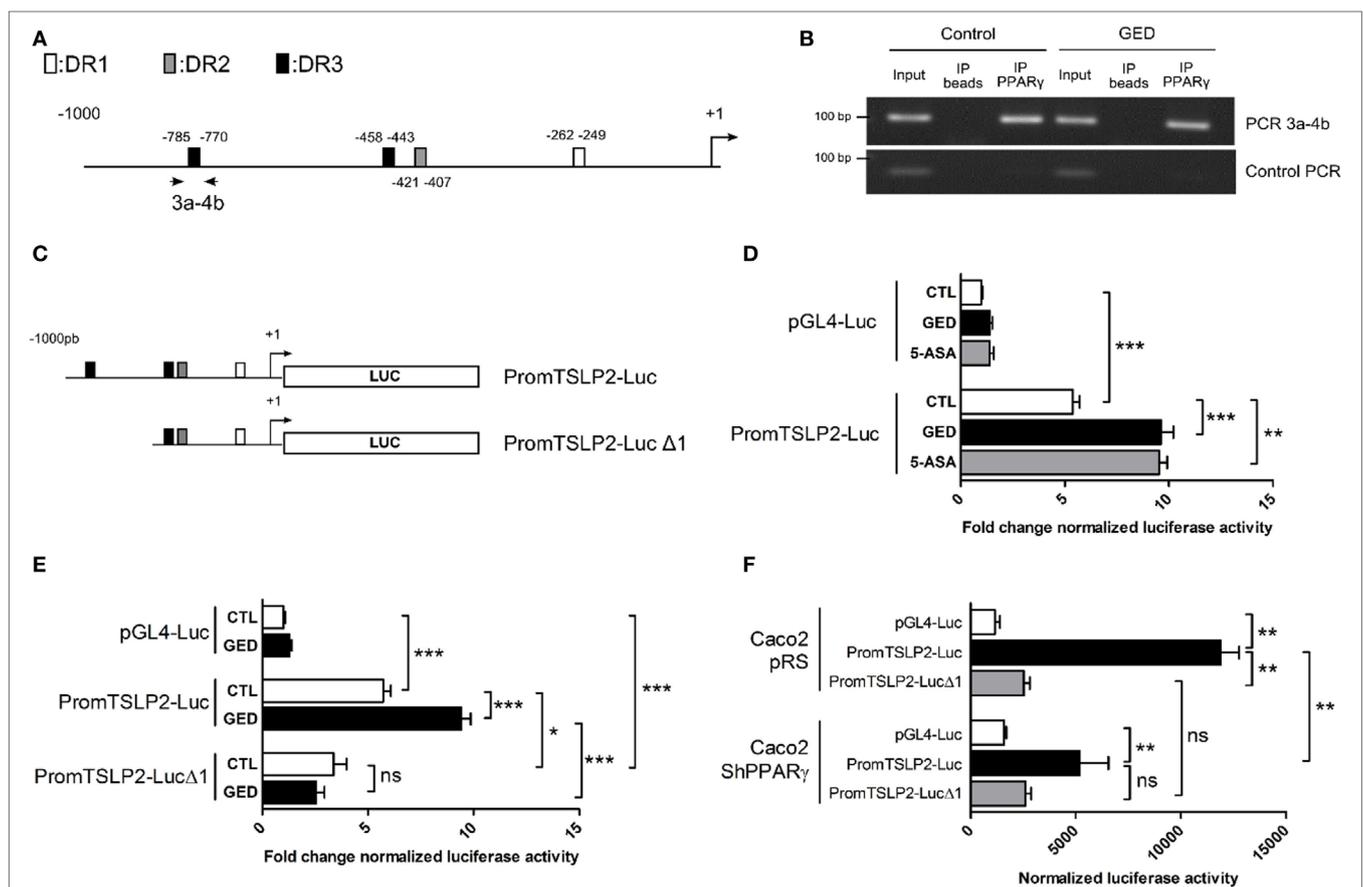


FIGURE 4 | Peroxisome proliferator activated receptor-gamma (PPAR γ) binds functionally to the promoter of sFTSLP gene. **(A)** Schematic representation of the promoter of the sFTSLP gene (1,000 base pairs upstream of the transcription start site) showing the potential presence of PPAR response element predicted by *in silico*. **(B)** Picture showing PCR amplification of the 3a-4b fragment in ChIP assay from Caco-2 cells. “Control PCR” corresponds to the amplification of a genomic region unrelated to sFTSLP promoter and serves as a control of specificity for the ChIP analysis. **(C)** Schematic representation of the two reporter constructions used in transient transfection and reporter gene assays. **(D,E)** Luciferase gene reporter assays in Caco-2 cells transfected with PromTSLP2-Luc and PromTSLP2-Luc Δ 1 reporter constructs. Cells transfected with empty pGL4Luc plasmid were used as control. Results represent the mean \pm SEM (3 independent experiments in triplicate or sextuplicate, $9 < n < 15$) of the fold change of luciferase activity normalized for protein content. The normalized Luc activity obtained in unstimulated Caco-2 cells transfected with empty pGL4Luc was used as reference and defined as 1. **(F)** Luciferase gene reporter assays in PPAR γ knock-down Caco-2 cells transfected with PromTSLP2-Luc and PromTSLP2-Luc Δ 1 reporter constructs. Cells transfected with empty pGL4Luc plasmid were used as control. Results represent the mean \pm SEM of luciferase activity normalized for protein content (two independent experiments in triplicate). * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; ns, not significant.

by PPAR γ was deleted, giving “promTSLP2-Luc Δ 1” construct (Figure 4C). Then, both plasmids were tested in transient reporter gene assays in Caco-2 cells (Figures 4D–F). First, in unstimulated conditions, the transfection of promTSLP2-Luc plasmids led to a significant five fold increase of luciferase activity compared to the transfection of empty pGL4-Luc vector, indicating that the portion of promoter present in the promTSLP2-Luc construct is active in Caco-2 cells (Figure 4D, compare white bar graphs). Stimulation of “promTSLP2-Luc”-transfected cells with GED or 5-ASA induced a significant additional increase of the reporter gene activity compared to unstimulated transfected cells, suggesting that putative functional PPAR γ -responsive sequences are present within the genomic fragment cloned in promTSLP2-Luc construct (Figure 4D, compare white, black, and gray bar graphs in promTSLP2-Luc transfected cells). In order to test whether the DR3 identified by the ChIP experiments could be this functional element, we tested the “promTSLP2-Luc Δ 1” reporter construct in transient transfection (Figure 4E). As expected, transfection of Caco-2 cells with “promTSLP2-Luc” construct gave the same luciferase induction as previously observed and, while in unstimulated conditions, the construction “promTSLP2-Luc Δ 1” retained some transcriptional activity (Figure 4E, compare white bar graphs), the inducing effect of the GED was totally lost (Figure 4E, compare white and black bar graphs in promTSLP2-Luc Δ 1 transfected cells). This result strongly suggests that the DR3 found to be bound by PPAR γ in ChIP experiment is functional in the sTSLP gene promoter. To irrevocably establish the involvement of PPAR γ in the regulation of the activity of the sTSLP promoter, we transfected PPAR γ knock-down Caco-2 cells with our two reporter plasmids (Figure 4F). In control cells (Caco-2 pRS cells, corresponding to Caco-2 cells having stably integrated the empty pRS vector used to express shRNA), the transfection of “promTSLP2-Luc” plasmid led to a significant increase of luciferase activity compared to cells transfected with the empty pGL4-Luc vector. The transfection of “promTSLP2-Luc Δ 1” provoked a dramatic decrease of luciferase activity compared to cells transfected with the “promTSLP2-Luc” plasmid, confirming our last result and the importance of the genomic region encompassing the DR3 element in the transcriptional activity of the sTSLP gene promoter. The knock-down of PPAR γ expression in Caco-2 shPPAR γ cells significantly decreased the luciferase activity of the “promTSLP2-Luc” construction (Figure 4F, compare black bar graphs), while it had no effect on the remaining transcriptional activity of the “promTSLP2-Luc Δ 1” construction (Figure 4F, compare gray bar graphs). These results clearly confirmed that PPAR γ is involved in the control of the activity of the sTSLP promoter. Taking together, our results demonstrate that PPAR γ binds functionally to the promoter of sTSLP and show that PPAR γ is a transcriptional regulator of sTSLP gene.

Decreased sTSLP Expression in Patients with UC Correlates with PPAR γ Expression

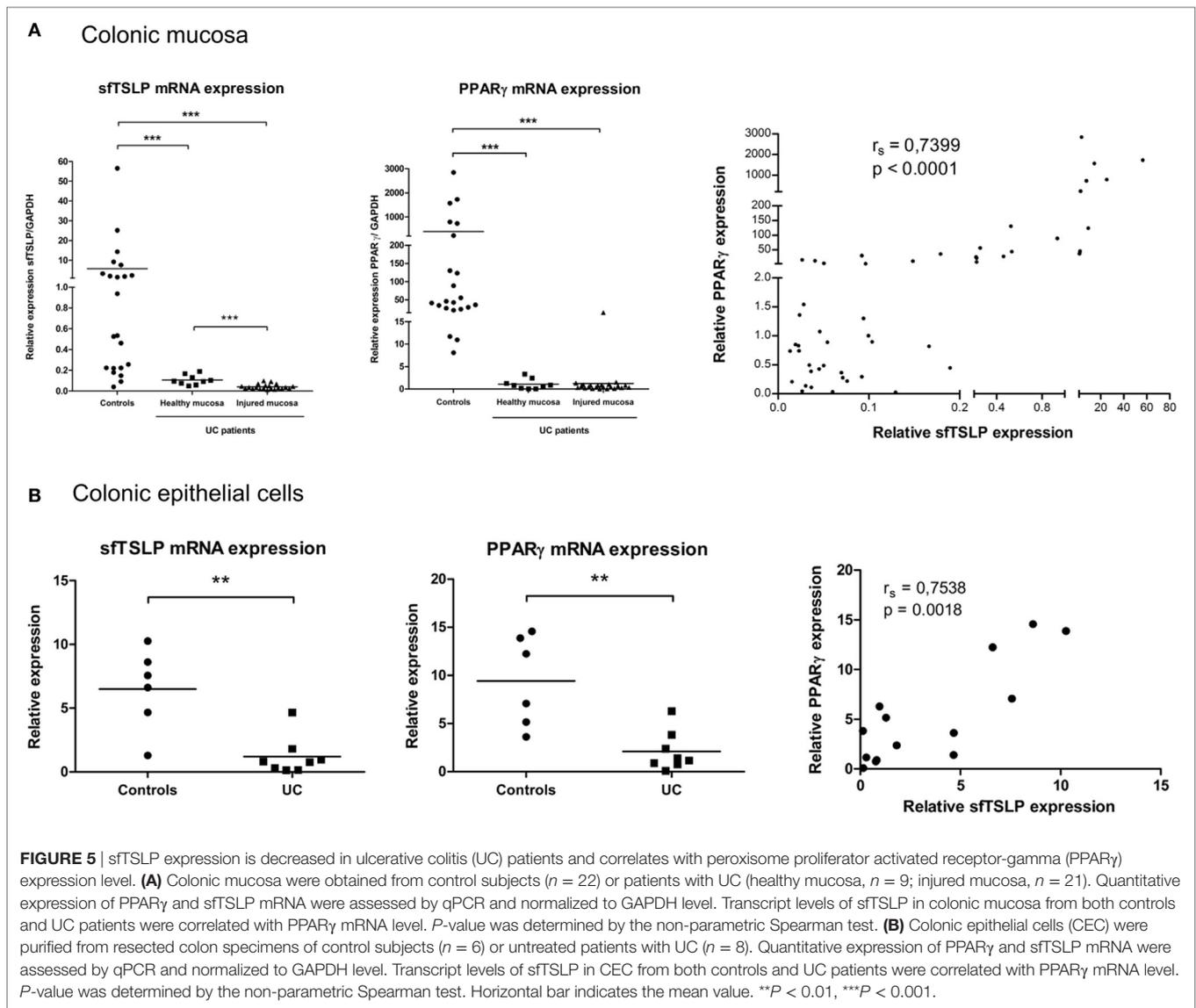
We and others have previously demonstrated that PPAR γ expression is impaired in the colonic mucosa and in the CEC of patients suffering from UC (9, 10, 28). Given our results demonstrating

the relationship between PPAR γ and sTSLP, we next assessed the expression level of TSLP isoforms in the colonic mucosa and CEC of UC patients and non-IBD controls. We first confirmed the significant impairment of PPAR γ mRNA expression, both in healthy and injured colonic mucosa from UC patients compared to control specimens (Figure 5A). We never obtained satisfactory lTSLP mRNA detection in our colonic patient samples, even when trying with different combinations of newly designed and already published PCR primers (26) (see Figure S1 in Supplementary Material). Interestingly, we observed that, in UC patients, the expression of sTSLP was significantly decreased in healthy and injured mucosa compared to controls specimens (Figure 5A), and we found a significant correlation between the expression level of PPAR γ and sTSLP in the colonic mucosa of all our patient samples (UC and controls) (Figure 5A). We finally assessed the expression levels of sTSLP and PPAR γ in CEC and found similar results: sTSLP mRNA expression level was significantly decreased specifically in purified CEC from patients with UC and correlated with PPAR γ mRNA expression level (Figure 5B). Taken together, these results therefore strengthen the relationship between PPAR γ and sTSLP and support the hypothesis that, in UC, the impaired expression of PPAR γ in colonic mucosa and CEC could play a role in the loss of sTSLP expression.

DISCUSSION

To our knowledge, the present study is the first to address the transcriptional regulation of the sTSLP, at least in the intestine. Our results identified the nuclear receptor PPAR γ as an important transcriptional regulator of sTSLP in CEC and strongly suggest that the lack of expression of sTSLP that we observed in the colonic mucosa and CEC of patients with UC is related to that of PPAR γ .

The distinction between the two isoforms of TSLP has only been made since very recently. It is now a fundamental aspect of the biology of this cytokine because the two isoforms possess distinct functions and expression profiles (26, 27, 29). *In vitro*, lTSLP expression is regulated by inflammatory stimuli. For instance, only the expression of lTSLP is induced in normal human bronchial epithelial cells after polyI:C exposure (30). In human keratinocytes, toll-like receptors ligands (polyI:C, FSL-1, and flagellin) as well as pro-inflammatory cytokines (IFN γ , TNF, IL-1 β) are potent inducers of lTSLP, but not of sTSLP (25, 31). Finally, exposure of Caco-2 cells to *Salmonella typhimurium* induces upregulation of lTSLP protein expression concomitantly to a downregulation of sTSLP expression (26). Consistent with these *in vitro* observations, lTSLP was found to be upregulated *in vivo* in inflammatory conditions such as IBD, asthma or atopic dermatitis and NF- κ B was identified as a critical factor for inflammation-induced expression of lTSLP (32, 33). In terms of function, although most of the work done so far has very rarely distinguished the two isoforms, the data suggest that lTSLP has proinflammatory properties (27, 29). Conversely, anti-inflammatory properties have been suggested, both *in vitro* and *in vivo*, for sTSLP which is the predominant form of TSLP, constitutively expressed at steady state (26, 29).



More interestingly, recent work has demonstrated that synthetic sFTSLP displays antimicrobial activity, suggesting that this isoform may act as an antimicrobial peptide (25, 27). In this context, our results appear to be highly relevant. Indeed, although the exact role of PPAR γ in the intestinal tract still remains to be characterized, it has been suggested that this factor plays a key role in gut homeostasis. Depletion of PPAR γ in mouse CEC increases susceptibility to experimental colitis, while activation of this receptor reduces intestinal inflammation and disease severity both in experimental models of colitis and in UC patients. One explanation of this positive role on gut homeostasis is the potential PPAR γ -dependent antimicrobial barrier function. Indeed, it has been suggested that PPAR γ mediates antimicrobial immunity in the colon, notably through the control of antimicrobial β -defensin-1 and mannose-binding lectin gene expression (34, 35). Therefore, the identification of the antimicrobial peptide sFTSLP as new PPAR γ -target gene in CEC broadens the role of this nuclear receptor both as an important antibacterial factor at

the gut mucosal surfaces as well as a key actor during intestinal homeostasis.

To date, only one study has investigated the differential expression of the TSLP isoforms in the intestine under normal conditions and in patients suffering from IBD (26). One of the main findings of this work was that the sFTSLP/lFTSLP ratio was altered (decreased) in patients with UC due to an upregulation of lFTSLP expression in colonic mucosa from UC patients compared to control subjects, whereas sFTSLP levels were unchanged. This perturbation could reflect the loss of intestinal homeostasis and could lead (or participate) to an uncontrolled inflammatory response. However, contrary to the study of Fornasa et al., we did not detect lFTSLP mRNA, either in Caco-2 or T84 cells nor in our human colonic samples (both from control and UC patients). We used different primer pairs (our own primers and already published primers) and, although they all allowed amplifying the long form from H292 bronchial epithelial cells, no primer pair efficiently amplified lFTSLP from CEC or colonic mucosa. Despite

this, we do believe that our results obtained on patient samples are not necessarily in conflict with those of Fornasa et al. (26), but could be rather in the same line. The decrease in expression of sTSLP that we observed, in association with an apparent unmodified level of lTSLP, results in a decrease in the sTSLP/lTSLP ratio as described in the work of Fornasa et al. On the other hand, the impaired expression of sTSLP in the colonic mucosa of our patients with UC is consistent with the suggested immunoregulatory/antimicrobial role of this isoform. This decreased pattern of expression of sTSLP in UC patients in our samples was confirmed by using the primer pair used to amplify sTSLP by Fornasa et al. (Figure S3 in Supplementary Material). *In vitro*, it has been suggested that the expression of the short form is inhibited by proinflammatory stimuli (26). Because we also observed lowered sTSLP expression in healthy mucosa from UC patients, it seems unlikely that the perturbed levels of sTSLP are related to the inflammatory process in our samples. In addition, we did not observe any correlation between the expression levels of sTSLP and IL-8 (Figure S4 in Supplementary Material). The significant correlation obtained between the expression levels of PPAR γ and sTSLP rather suggests that the defect of sTSLP is related to the loss of expression of PPAR γ typically observed in the CEC of UC patients. This correlation strengthens the functional link between these two factors. Moreover, these defects observed in inflamed and healthy colons of patients with UC could suggest a primary role of colonic PPAR γ and sTSLP impairment in the pathophysiology of UC. The discrepancy between the results published by Fornasa et al. and those of our study could be due to the nature of the colonic samples used. In the Fornasa study, it seems that most of the samples of UC patients came from the sigmoid colon, whereas in our work, very few samples are derived from sigmoidectomy but are rather more proximal colonic samples coming from partial or total colectomies (Table S2 in Supplementary Material). The expression profile of TSLP isoforms along the gut (proximal colon versus distal colon for example) is unknown, but it may exist some differences that could explain these divergent results. Thus, it would be important to precisely determine the expression profile of TSLP isoforms along the normal human gastrointestinal tract in order to better understand the biology of this cytokine.

It is important to note that the sTSLP does not exist in mouse and has been isolated only in humans. Consequently, the relevance of using mice models to study the sTSLP functions is limited. It has been suggested that intraperitoneal injection of sTSLP slightly improves DSS-induced colitis, but the underlying

mechanism was not investigated and thus remains unknown (26). Given the unique presence of sTSLP in humans, it would be more relevant to test the antimicrobial activity of sTSLP on commensal and pathological bacteria associated with human gut. The recent development of colonic organoids of human origin (36–38) should also make it possible to better understand the role of sTSLP during intestinal homeostasis.

In conclusion, the present study has identified a new PPAR γ target gene in CEC, confirming the key role of this nuclear receptor in antimicrobial immunity and during intestinal homeostasis. In addition, it suggests a new molecular mechanism involved in the loss of intestinal tolerance observed during UC.

ETHICS STATEMENT

A local ethics committee (Comité de Protection des Personnes Nord Ouest IV, CHRU Lille, France) approved the study and all subjects gave informed consent (No. DC-2008-642). Surgical colonic samples were obtained from patients with an established diagnosis of UC according to international criteria and from control patients (patients underwent surgery for colorectal cancer and patients for diverticulitis of the sigmoid). The detailed characteristics of patient cohort are given in Table S2 in Supplementary Material.

AUTHOR CONTRIBUTIONS

AMM, PD, LD, and BB designed the study. AMM, AL, SS, and BB performed experiments and analysis of the data. LS collected human colonic samples and managed clinical data. AMM, LD, and BB wrote the paper and participated to the critical reading of the manuscript.

ACKNOWLEDGMENTS

We acknowledge the support of the DigestScience Foundation. We deeply acknowledge Lennart Mars for the critical reading of the manuscript.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at <http://journal.frontiersin.org/article/10.3389/fimmu.2017.01052/full#supplementary-material>.

REFERENCES

1. Sansonetti PJ. War and peace at mucosal surfaces. *Nat Rev Immunol* (2004) 4(12):953–64. doi:10.1038/nri1499
2. Peterson LW, Artis D. Intestinal epithelial cells: regulators of barrier function and immune homeostasis. *Nat Rev Immunol* (2014) 14(3):141–53. doi:10.1038/nri3608
3. Rimoldi M, Chieppa M, Salucci V, Avogadri F, Sonzogni A, Sampietro GM, et al. Intestinal immune homeostasis is regulated by the crosstalk between epithelial cells and dendritic cells. *Nat Immunol* (2005) 6(5):507–14. doi:10.1038/nri1192
4. Iliev ID, Mileti E, Matteoli G, Chieppa M, Rescigno M. Intestinal epithelial cells promote colitis-protective regulatory T-cell differentiation through dendritic cell conditioning. *Mucosal Immunol* (2009) 2(4):340–50. doi:10.1038/mi.2009.13
5. Iliev ID, Spadoni I, Mileti E, Matteoli G, Sonzogni A, Sampietro GM, et al. Human intestinal epithelial cells promote the differentiation of tolerogenic dendritic cells. *Gut* (2009) 58(11):1481–9. doi:10.1136/gut.2008.175166
6. Anderson CA, Boucher G, Lees CW, Franke A, D'Amato M, Taylor KD, et al. Meta-analysis identifies 29 additional ulcerative colitis risk loci, increasing the number of confirmed associations to 47. *Nat Genet* (2011) 43(3):246–52. doi:10.1038/ng.764
7. Danese S, Fiocchi C. Ulcerative colitis. *N Engl J Med* (2011) 365(18):1713–25. doi:10.1056/NEJMra1102942
8. Abraham C, Cho JH. Inflammatory bowel disease. *N Engl J Med* (2009) 361(21):2066–78. doi:10.1056/NEJMra0804647

9. Dubuquoy L, Jansson EA, Deeb S, Rakotobe S, Karoui M, Colombel JF, et al. Impaired expression of peroxisome proliferator-activated receptor gamma in ulcerative colitis. *Gastroenterology* (2003) 124(5):1265–76. doi:10.1016/S0016-5085(03)00271-3
10. Bouguen G, Langlois A, Djouina M, Branche J, Koriche D, Dewaeles E, et al. Intestinal steroidogenesis controls PPARgamma expression in the colon and is impaired during ulcerative colitis. *Gut* (2015) 64(6):901–10. doi:10.1136/gutjnl-2014-307618
11. Tontonoz P, Spiegelman BM. Fat and beyond: the diverse biology of PPARgamma. *Annu Rev Biochem* (2008) 77:289–312. doi:10.1146/annurev.biochem.77.061307.091829
12. Bertin B, Dubuquoy L, Colombel JF, Desreumaux P. PPAR-gamma in ulcerative colitis: a novel target for intervention. *Curr Drug Targets* (2013) 14(12):1501–7. doi:10.2174/13894501113149990162
13. Adachi M, Kurotani R, Morimura K, Shah Y, Sanford M, Madison BB, et al. Peroxisome proliferator activated receptor gamma in colonic epithelial cells protects against experimental inflammatory bowel disease. *Gut* (2006) 55(8):1104–13. doi:10.1136/gut.2005.081745
14. Su CG, Wen X, Bailey ST, Jiang W, Rangwala SM, Keilbaugh SA, et al. A novel therapy for colitis utilizing PPAR-gamma ligands to inhibit the epithelial inflammatory response. *J Clin Invest* (1999) 104(4):383–9. doi:10.1172/JCI7145
15. Desreumaux P, Dubuquoy L, Nutten S, Peuchmaur M, Englaro W, Schoonjans K, et al. Attenuation of colon inflammation through activators of the retinoid X receptor (RXR)/peroxisome proliferator-activated receptor gamma (PPARgamma) heterodimer. A basis for new therapeutic strategies. *J Exp Med* (2001) 193(7):827–38. doi:10.1084/jem.193.7.827
16. Lewis JD, Lichtenstein GR, Deren JJ, Sands BE, Hanauer SB, Katz JA, et al. Rosiglitazone for active ulcerative colitis: a randomized placebo-controlled trial. *Gastroenterology* (2008) 134(3):688–95. doi:10.1053/j.gastro.2007.12.012
17. Fajas L, Auboeuf D, Raspe E, Schoonjans K, Lefebvre AM, Saladin R, et al. The organization, promoter analysis, and expression of the human PPARgamma gene. *J Biol Chem* (1997) 272(30):18779–89. doi:10.1074/jbc.272.30.18779
18. Dudakov JA, Hanash AM, van den Brink MR. Interleukin-22: immunobiology and pathology. *Annu Rev Immunol* (2015) 33:747–85. doi:10.1146/annurev-immunol-032414-112123
19. Lindemans CA, Calafiore M, Mertelmann AM, O'Connor MH, Dudakov JA, Jenq RR, et al. Interleukin-22 promotes intestinal-stem-cell-mediated epithelial regeneration. *Nature* (2015) 528(7583):560–4. doi:10.1038/nature16460
20. Podvynec M, Kaufmann MR, Handschin C, Meyer UA. NUBIScan, an in silico approach for prediction of nuclear receptor response elements. *Mol Endocrinol* (2002) 16(6):1269–79. doi:10.1210/mend.16.6.0851
21. Venkatachalam G, Kumar AP, Yue LS, Pervaiz S, Clement MV, Sakharkar MK. Computational identification and experimental validation of PPRE motifs in NHE1 and MnSOD genes of human. *BMC Genomics* (2009) 10(Suppl 3):S5. doi:10.1186/1471-2164-10-S3-S5
22. Lehmann JM, Moore LB, Smith-Oliver TA, Wilkison WO, Willson TM, Kliewer SA. An antidiabetic thiazolidinedione is a high affinity ligand for peroxisome proliferator-activated receptor gamma (PPAR gamma). *J Biol Chem* (1995) 270(22):12953–6. doi:10.1074/jbc.270.22.12953
23. Rousseaux C, Lefebvre B, Dubuquoy L, Lefebvre P, Romano O, Auwerx J, et al. Intestinal antiinflammatory effect of 5-aminosalicylic acid is dependent on peroxisome proliferator-activated receptor-gamma. *J Exp Med* (2005) 201(8):1205–15. doi:10.1084/jem.20041948
24. Mastromarino A, Kovacs D, Sarra M, Bastonini E, Cardinali G, Aspini N, et al. Preclinical studies of a specific PPARgamma modulator in the control of skin inflammation. *J Invest Dermatol* (2014) 134(4):1001–11. doi:10.1038/jid.2013.448
25. Bjerkan L, Schreurs O, Engen SA, Jahnsen FL, Baekkevold ES, Blix IJ, et al. The short form of TSLP is constitutively translated in human keratinocytes and has characteristics of an antimicrobial peptide. *Mucosal Immunol* (2015) 8(1):49–56. doi:10.1038/mi.2014.41
26. Fornasa G, Tsilingiri K, Caprioli F, Botti F, Mapelli M, Meller S, et al. Dichotomy of short and long thymic stromal lymphopoietin isoforms in inflammatory disorders of the bowel and skin. *J Allergy Clin Immunol* (2015) 136(2):413–22. doi:10.1016/j.jaci.2015.04.011
27. Bjerkan L, Sonesson A, Schenck K. Multiple functions of the new cytokine-based antimicrobial peptide thymic stromal lymphopoietin (TSLP). *Pharmaceuticals (Basel)* (2016) 9(3):41. doi:10.3390/ph9030041
28. Pedersen G, Brynskov J. Topical rosiglitazone treatment improves ulcerative colitis by restoring peroxisome proliferator-activated receptor-gamma activity. *Am J Gastroenterol* (2010) 105(7):1595–603. doi:10.1038/ajg.2009.749
29. Dong H, Hu Y, Liu L, Zou M, Huang C, Luo L, et al. Distinct roles of short and long thymic stromal lymphopoietin isoforms in house dust mite-induced asthmatic airway epithelial barrier disruption. *Sci Rep* (2016) 6:39559. doi:10.1038/srep39559
30. Harada M, Hirota T, Jodo AI, Doi S, Kameda M, Fujita K, et al. Functional analysis of the thymic stromal lymphopoietin variants in human bronchial epithelial cells. *Am J Respir Cell Mol Biol* (2009) 40(3):368–74. doi:10.1165/rcmb.2008-0041OC
31. Xie Y, Takai T, Chen X, Okumura K, Ogawa H. Long TSLP transcript expression and release of TSLP induced by TLR ligands and cytokines in human keratinocytes. *J Dermatol Sci* (2012) 66(3):233–7. doi:10.1016/j.jdermsci.2012.03.007
32. Lee HC, Ziegler SF. Inducible expression of the proallergic cytokine thymic stromal lymphopoietin in airway epithelial cells is controlled by NFkappaB. *Proc Natl Acad Sci U S A* (2007) 104(3):914–9. doi:10.1073/pnas.0607305104
33. Cultrone A, de Wouters T, Lakhdari O, Kelly D, Mulder I, Logan E, et al. The NF-kappaB binding site located in the proximal region of the TSLP promoter is critical for TSLP modulation in human intestinal epithelial cells. *Eur J Immunol* (2013) 43(4):1053–62. doi:10.1002/eji.201142340
34. Peyrin-Biroulet L, Beisner J, Wang G, Nuding S, Oommen ST, Kelly D, et al. Peroxisome proliferator-activated receptor gamma activation is required for maintenance of innate antimicrobial immunity in the colon. *Proc Natl Acad Sci U S A* (2010) 107(19):8772–7. doi:10.1073/pnas.0905745107
35. Choteau L, Parny M, Francois N, Bertin B, Fumery M, Dubuquoy L, et al. Role of mannose-binding lectin in intestinal homeostasis and fungal elimination. *Mucosal Immunol* (2016) 9(3):767–76. doi:10.1038/mi.2015.100
36. Miyoshi H, Stappenbeck TS. In vitro expansion and genetic modification of gastrointestinal stem cells in spheroid culture. *Nat Protoc* (2013) 8(12):2471–82. doi:10.1038/nprot.2013.153
37. VanDussen KL, Marinshaw JM, Shaikh N, Miyoshi H, Moon C, Tarr PI, et al. Development of an enhanced human gastrointestinal epithelial culture system to facilitate patient-based assays. *Gut* (2014) 64(6):911–20. doi:10.1136/gutjnl-2013-306651
38. Fatehullah A, Tan SH, Barker N. Organoids as an in vitro model of human development and disease. *Nat Cell Biol* (2016) 18(3):246–54. doi:10.1038/ncb3312

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Martin Mena, Langlois, Specia, Schneider, Desreumaux, Dubuquoy and Bertin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.