



**HAL**  
open science

## Marine-inspired enzymatic mineralization of dairy-derived whey protein isolate (wpi) hydrogels for bone tissue regeneration

Karl Norris, Magdalena Kocot, Anna M. Tryba, Feng Chai, Abdullah Talari, Lorna Ashton, Bogdan V. Parakhonskiy, Sangram Keshari Samal, Nicolas Blanchemain, Elzbieta Pamula, et al.

### ► To cite this version:

Karl Norris, Magdalena Kocot, Anna M. Tryba, Feng Chai, Abdullah Talari, et al.. Marine-inspired enzymatic mineralization of dairy-derived whey protein isolate (wpi) hydrogels for bone tissue regeneration. *Marine drugs*, 2020, 18 (6), pp.294. 10.3390/md18060294 . hal-03798594

**HAL Id: hal-03798594**

**<https://hal.univ-lille.fr/hal-03798594>**

Submitted on 5 Oct 2022

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Article

# Marine-Inspired Enzymatic Mineralization of Dairy-Derived Whey Protein Isolate (WPI) Hydrogels for Bone Tissue Regeneration

Karl Norris <sup>1,\*</sup>, Magdalena Kocot <sup>2</sup>, Anna M. Tryba <sup>2</sup>, Feng Chai <sup>3</sup>, Abdullah Talari <sup>1,4</sup>, Lorna Ashton <sup>4</sup>, Bogdan V. Parakhonskiy <sup>5,6</sup>, Sangram K. Samal <sup>7</sup>, Nicholas Blanchemain <sup>3</sup>, Elżbieta Pamuła <sup>2</sup> and Timothy E. L. Douglas <sup>1,8</sup>

<sup>1</sup> Engineering Department, Lancaster University, Lancaster LA1 4YW, UK; a.talari@lancaster.ac.uk (A.T.); t.douglas@lancaster.ac.uk (T.E.L.D.)

<sup>2</sup> Department of Biomaterials and Composites, Faculty of Materials Science and Ceramics, AGH University of Science and Technology, 30-962 Kraków, Poland; kocotmagda@gmail.com (M.K.); amtryba@agh.edu.pl (A.M.T.); epamuła@agh.edu.pl (E.P.)

<sup>3</sup> INSERM U1008-Controlled Drug Delivery Systems and Biomaterials, Université de Lille, 59006 Lille, France; fchai@univ-lille2.fr (F.C.); nicolas.blanchemain@univ-lille.fr (N.B.)

<sup>4</sup> Chemistry Department, Lancaster University, Lancaster LA1 4YW, UK; l.ashton@lancaster.ac.uk

<sup>5</sup> Department of Biotechnology, Ghent University, B-9000 Gent, Belgium; Bogdan.parakhonskiy@ugent.be

<sup>6</sup> Nanotechnology Department, Saratov State University, Saratov 410012, Russia

<sup>7</sup> Laboratory of Biomaterials and Regenerative Medicine for Advanced Therapies, Indian Council of Medical Research-Regional Medical Research Center, Bhubaneswar, Odisha 751023, India; sksamalrec@gmail.com

<sup>8</sup> Materials Science Institute (MSI), Lancaster University, Lancaster LA1 4YW, UK

\* Correspondence: hwbkn3@gmail.com; Tel.: +44-113-34-38217

Received: 27 April 2020; Accepted: 28 May 2020; Published: 2 June 2020



**Abstract:** Whey protein isolate (WPI) is a by-product from the production of cheese and Greek yoghurt comprising  $\beta$ -lactoglobulin ( $\beta$ -lg) (75%). Hydrogels can be produced from WPI solutions through heating; hydrogels can be sterilized by autoclaving. WPI hydrogels have shown cytocompatibility and ability to enhance proliferation and osteogenic differentiation of bone-forming cells. Hence, they have promise in the area of bone tissue regeneration. In contrast to commonly used ceramic minerals for bone regeneration, a major advantage of hydrogels is the ease of their modification by incorporating biologically active substances such as enzymes. Calcium carbonate ( $\text{CaCO}_3$ ) is the main inorganic component of the exoskeletons of marine invertebrates. Two polymorphs of  $\text{CaCO}_3$ , calcite and aragonite, have shown the ability to promote bone regeneration. Other authors have reported that the addition of magnesium to inorganic phases has a beneficial effect on bone-forming cell growth. In this study, we employed a biomimetic, marine-inspired approach to mineralize WPI hydrogels with an inorganic phase consisting of  $\text{CaCO}_3$  (mainly calcite) and  $\text{CaCO}_3$  enriched with magnesium using the calcifying enzyme urease. The novelty of this study lies in both the enzymatic mineralization of WPI hydrogels and enrichment of the mineral with magnesium. Calcium was incorporated into the mineral formed to a greater extent than magnesium. Increasing the concentration of magnesium in the mineralization medium led to a reduction in the amount and crystallinity of the mineral formed. Biological studies revealed that mineralized and unmineralized hydrogels were not cytotoxic and promoted cell viability to comparable extents (approximately 74% of standard tissue culture polystyrene). The presence of magnesium in the mineral formed had no adverse effect on cell viability. In short, WPI hydrogels, both unmineralized and mineralized with  $\text{CaCO}_3$  and magnesium-enriched  $\text{CaCO}_3$ , show potential as biomaterials for bone regeneration.

**Keywords:** hydrogel; composite; mineralization; enzyme; bioinspired; whey protein isolate

## 1. Introduction

Natural by-products from industrial processes have tremendous biomimetic properties and are inexpensive as they are infrequently utilized. Whey protein isolate (WPI) is a by-product from the production of cheese and Greek yoghurt comprising  $\beta$ -lactoglobulin ( $\beta$ -lg) (50%) and  $\alpha$ -lactalbumin ( $\alpha$ -la) (20%) [1]. WPI in solution has been shown to enhance osteogenic differentiation of bone-forming cells and promote cellular proliferation [1]. It was recently discovered that hydrogels can be produced from WPI solutions, whereby gelation is achieved through heating and sterilization is achieved by autoclaving [2–4].

With regards to mechanical properties, hydrogels are relatively weak due to the high water content. However, the incorporation of a mineral phase may improve the mechanical properties and therefore, promote cellular adhesion, proliferation and osteogenic differentiation [5]. In comparison to ceramic minerals, a major advantage of hydrogels is that they can be modified with ease by incorporating biologically active substances such as enzymes. Similarly, it is possible to modify and enhance properties of hydrogels by enriching the solution with a mineral phase either before or after gelation [6].

Marine invertebrates use the mineral calcium carbonate ( $\text{CaCO}_3$ ) in their exoskeletons.  $\text{CaCO}_3$  occurs naturally as three crystalline polymorphs known as calcite, aragonite and vaterite. Calcite occurs in bivalves and certain sponges, while aragonite occurs in coral and nacre, or “mother of pearl”. All three polymorphs of  $\text{CaCO}_3$  may exhibit beneficial biological properties. For example, calcite is a bioactive substance that can form direct connections with bone tissue *in vivo* and could potentially strengthen fractured bone [7]. Aragonite has been employed as a biomaterial for bone regeneration and outperformed calcium phosphates [8], and when added in particle form to WPI hydrogels, has improved mechanical properties and proliferation of osteoblast-like cells [4]. Vaterite coatings have been shown to stimulate the formation of apatite upon incubation in simulated body fluid (SBF) [9]. Hence, one may hypothesize that enzymatic mineralization of WPI hydrogels with polymorphs of  $\text{CaCO}_3$  will improve their biological performance *in vitro* and *in vivo*.

Several approaches to mineralize hydrogels have been tried (Gkioni et al—Reference 5). The most popular approach is the addition of pre-formed ceramic particles (e.g., hydroxyapatite, bioactive glasses) during hydrogel formation. Enzymatic mineralization has certain advantages over addition of pre-formed particles. Firstly, particles are prone to aggregation, resulting in inhomogeneous distribution, while enzymatic mineralization can potentially lead to a more homogeneous distribution of the mineral and better integration of the mineral with the hydrogel network. Secondly, the amount of pre-formed ceramic particles which can be incorporated is limited to approximately 30–50% [5], as the presence of too many particles may impede formation of the hydrogel. Using enzymatic mineralization, larger mineral contents can be achieved.

In previous work, hydrogels have been mineralized enzymatically with calcium phosphate (CaP) using alkaline phosphatase (ALP), the enzyme responsible for mineralization of bone tissue. One advantage of using urease instead of ALP is the superior thermal stability of urease at higher temperatures such as 70 °C, at which WPI hydrogels are formed.

The reaction steps of mineral precipitation using urease have been described previously [10]. Briefly, urea diffuses into the hydrogel, where it is converted under the action of urease into ammonia and bicarbonate ions. After dissociation of bicarbonate ions to hydrogen and carbonate ions, the carbonate ions react with calcium, which has also diffused into the hydrogel, to form  $\text{CaCO}_3$ . The hydrogen ions formed by the dissociation of the bicarbonate ions are neutralized by the ammonia, resulting in a sufficiently high pH to allow  $\text{CaCO}_3$  precipitation.

Another beneficial enriching agent is magnesium, which can be found in the calcareous exoskeletons of certain marine organisms and may provide benefits to mammals [11]. Previous work has shown that the presence of magnesium in CaP has promoted adhesion and proliferation of osteoblastic cell lines [12]. During enzymatic mineralization of hydrogels with  $\text{CaCO}_3$ , magnesium ions can be added to the mineralization medium which may lead to an increase in cell number [13,14]. In previous work, Gellan Gum (GG) hydrogels have been mineralized with  $\text{CaCO}_3$  and/or calcium magnesium carbonate

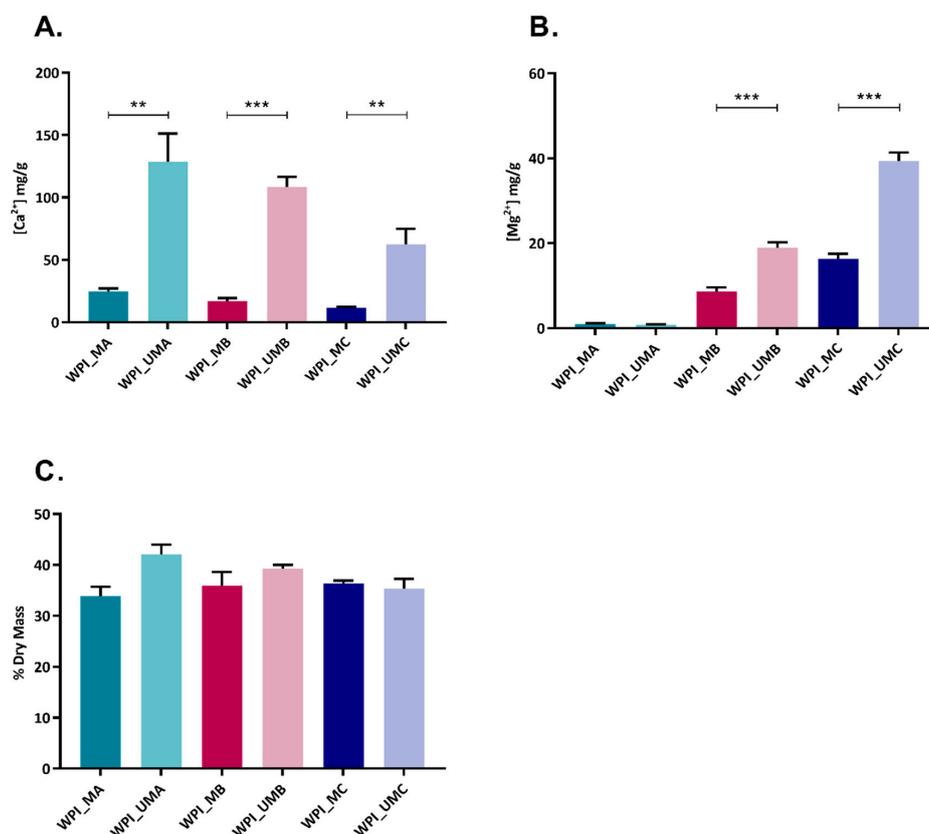
by incorporation of the enzyme urease into the hydrogel network followed by incubation in a solution containing the enzyme substrate, urea, and calcium and magnesium ions [14,15].

In the following study, the enzymatic mineralization approach was extended to WPI hydrogels. WPI hydrogels were mineralized with  $\text{CaCO}_3$  or magnesium-enriched  $\text{CaCO}_3$  via enzymatic mineralization with urease and incubation in solutions containing calcium and magnesium ions. The novelty of this study lies in both the enzymatic mineralization of WPI hydrogels and enrichment of the mineral with magnesium. It was thus demonstrated that urease retained activity after incorporation into WPI hydrogels, which involved gelation at 70 °C. Three different Ca:Mg concentration ratios were compared. The resulting hydrogels were subjected to physicochemical, morphological and biological characterization to examine the influence of Ca:Mg ratio.

## 2. Results

### 2.1. Influence of Mineralization Medium on Extent and Elemental Composition of Mineral Formed

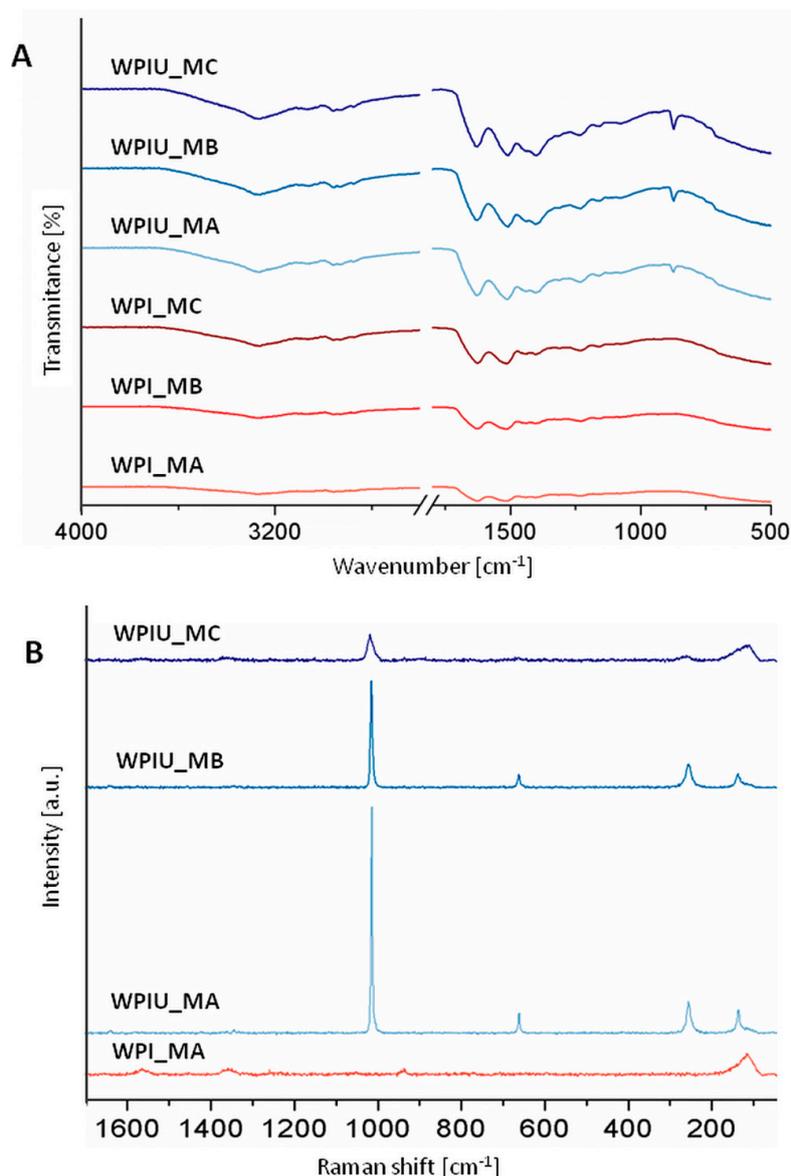
We aimed to enhance the properties of WPI hydrogels by mineralization with  $\text{CaCO}_3$  or magnesium-enriched  $\text{CaCO}_3$ . The effects of  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  content of the mineralization media on extent of mineral formed were investigated. As expected, a dose dependent effect was observed depending in which medium the urease WPI hydrogel was incubated (Table 1, Figure 1A,B). Notably, WPI hydrogels containing urease were able to retain higher amounts of  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  than control hydrogels, suggesting a greater extent of mineralization. When WPI hydrogels containing urease were incubated with calcium and magnesium in equimolar concentrations (WPI\_U\_MC), calcium content was five-fold that of magnesium, indicating that calcium was preferentially incorporated into the hydrated polymer. The increase in dry mass as a result of enzymatic mineralization was highest for hydrogels incubated in calcium only (WPI\_U\_MA) (Figure 1C).



**Figure 1.** Mass of elemental calcium (A) and magnesium (B) per unit mass of hydrogel and dry mass percentage (C) of WPI hydrogels without urease (WPI) and with urease (WPI\_U) incubated in different media: MA, MB, MC,  $n = 3$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ .

## 2.2. Physicochemical Characterization of Mineral Formed by FTIR, Raman, XRD and SEM

FTIR spectra (Figure 2A) of urease-free hydrogels indicated characteristic bands for WPI at 1650, 1570 and 1350–1200  $\text{cm}^{-1}$  [16]. In contrast, hydrogels containing urease showed bands characteristic of calcium carbonate. There was a broad band at 1400  $\text{cm}^{-1}$ , which is indicative of calcite and corresponds to  $\nu_3$  antisymmetric stretching of the carbonate group [17]. Moreover, a band at approximately 1500  $\text{cm}^{-1}$  was detected, which might indicate the presence of vaterite. This band was more intense in WPI\_U\_MA than in samples incubated in MB and MC. A band at approximately 1080  $\text{cm}^{-1}$  was also observed in all hydrogels with urease and might indicate that vaterite was formed within these hydrogels [18]. In all hydrogels with urease, bands at 870 and 715  $\text{cm}^{-1}$  correspond to  $\nu_2$  out-of-plane bending and  $\nu_4$  in-plane bending, respectively. These bands are characteristic for calcite [17]. These results show strong evidence that, in WPI hydrogels containing urease, the minerals calcite and vaterite were formed.



**Figure 2.** FTIR (A) and Raman spectra (B) of WPI hydrogels without and with urease WPI\_U incubated in different media: MA, MB, MC.

Raman spectra of control WPI\_MA hydrogels (Figure 2B) showed bands at 1658 and 1453  $\text{cm}^{-1}$ , which are associated with amide I and  $\text{CH}_2$  bending, respectively [19]. There were also bands at approximately 1000  $\text{cm}^{-1}$  relating to phenylalanine [20,21]. Bands typical for  $\text{CaCO}_3$  were not observed.

In contrast, WPI\_U\_MA and WPI\_U\_MB showed intense bands at  $1085\text{ cm}^{-1}$ , which are typical for  $\text{CaCO}_3$  and correspond to  $\nu_1$  symmetric stretching [22]. Raman bands were also observed at  $711\text{ cm}^{-1}$  relating to  $\nu_4$  in-plane stretching and both hydrogels showed bands at  $281$  and  $154\text{ cm}^{-1}$  consistent with lattice vibration modes [22]. Sharp peaks in WPI\_U\_MA and WPI\_U\_MB were indicative of crystalline structure. In WPI\_U\_MC, Raman bands at  $1085\text{ cm}^{-1}$  were broader and less intense, suggesting increased amorphicity, and thus, incorporation of magnesium in the calcite lattice. It was found that increased magnesium concentration reduced the crystallinity of  $\text{CaCO}_3$ . This observation was confirmed by two other bands typical for calcite (at  $281$  and  $154\text{ cm}^{-1}$ ), which were also broader in WPI\_U\_MC than in WPI\_U\_MA and WPI\_U\_MB [17,23,24]. The decrease in intensity and band sharpness in the order WPIU\_MA > WPIU\_MB > WPIU\_MC suggests a decrease in crystallinity in the same order.

XRD diffractograms of control samples (urease-free hydrogels) displayed a peak at a  $2\theta$  value of approximately  $19.5$ , which is a characteristic peak for WPI (Figure 3) [20]. However, in WPI hydrogels with urease (WPI\_U), this peak was less intense, which might suggest that the proportion of mineral present in this sample was higher than the proportion of WPI. Minerals formed in all WPI hydrogels containing urease, displaying peaks at  $23.2, 29.5, 36.1, 39.5, 43.3, 47.7, 48.6$  and  $57.5$ , which are characteristic for calcite [13,25]. In addition, WPI\_U\_MA hydrogels displayed peaks characteristic for vaterite at  $24.9, 27.0$  and  $32.8$  [26]. A peak near  $30$  degrees in WPI\_U\_MC hydrogels may indicate the presence of calcium magnesium carbonate. XRD may be also used to identify the degree of crystallinity of material. Crystalline materials show sharp peaks whereas amorphous produce a broad background pattern. Therefore, it could be concluded that WPI hydrogels with urease incubated in media MA and MB (WPIU\_MA and WPIU\_MB) are crystalline, as indicated by sharp peaks [13,25]. The lower intensity and sharpness of peaks of hydrogels incubated in medium MC (WPI\_U\_MC) suggest lower crystallinity.

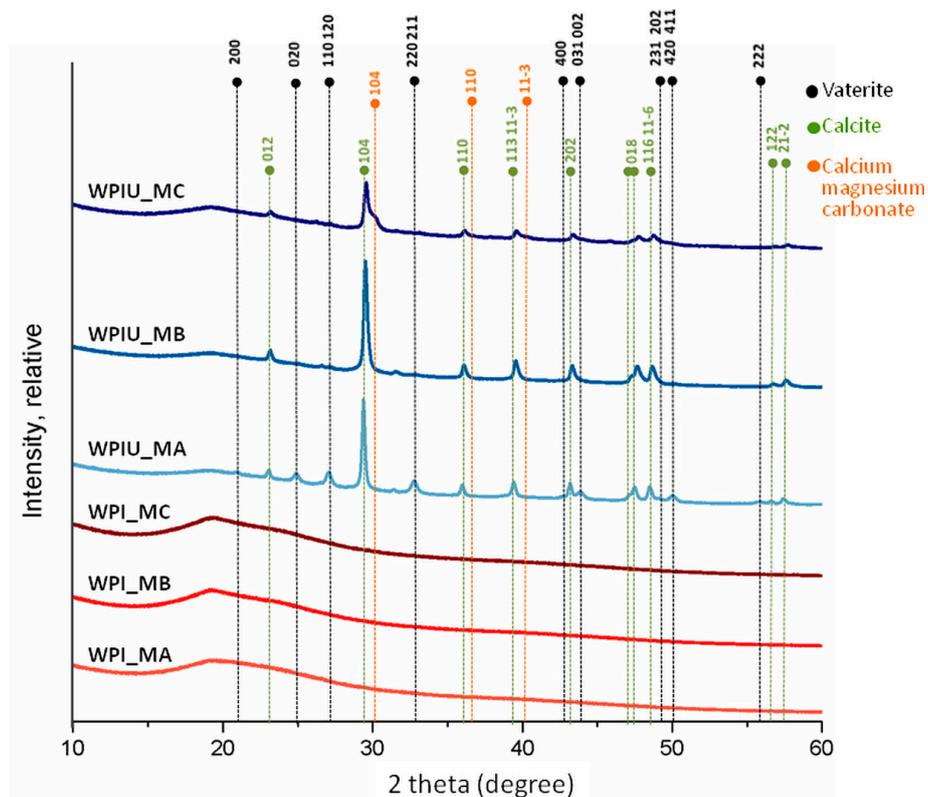
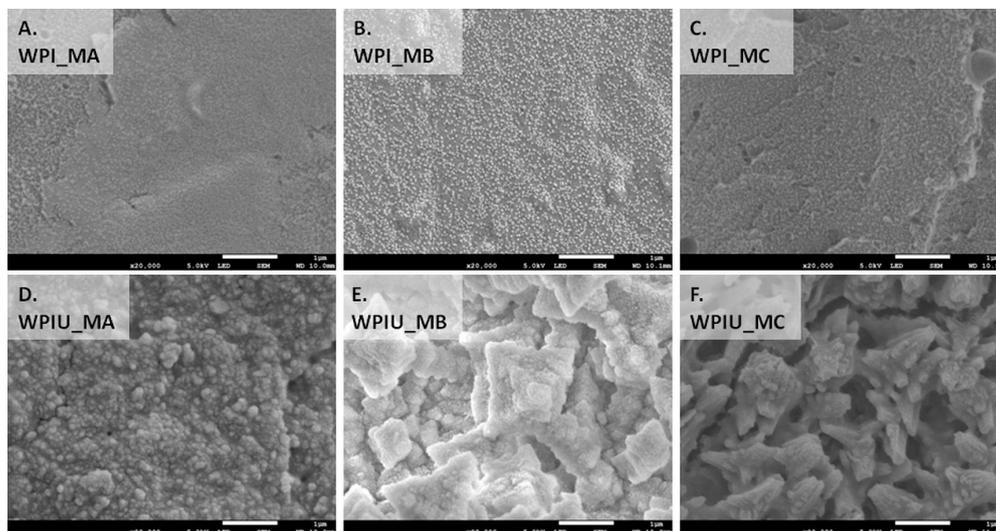


Figure 3. XRD diffractograms of WPI hydrogels incubated in different media MA, MB and MC.

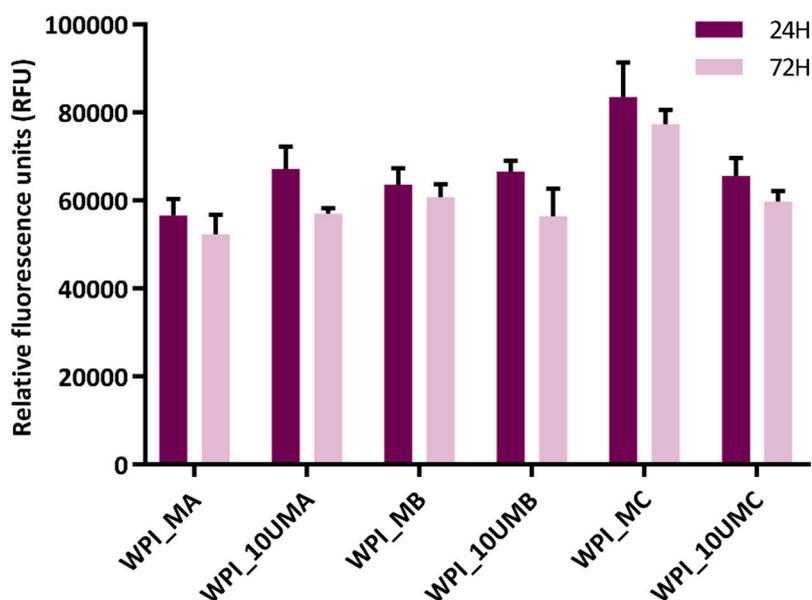
The morphology of the different WPI hydrogels was studied by SEM (Figure 4). As expected, WPI hydrogels without urease displayed surfaces devoid of mineral deposits (Figure 4A–C). The surface of hydrogels incubated in medium MA was smooth, whereas hydrogels incubated in media MB and MC had some deposits on the surface, which might have been polymer residues. In contrast, WPI hydrogels containing urease displayed microstructures rich in mineral deposits (Figure 4D–L). In mineralized hydrogels incubated in medium MA (WPI\_U\_MA), some porous cube-like deposits typical for calcite and spherical deposits typical for vaterite were observed [27,28]. Moreover, some undefined elongated, rod-like deposits were also detected on the surface of hydrogels incubated in medium MA. Cuboid deposits typical for calcite were also observed in WPI hydrogels incubated in media containing magnesium (MB and MC). However, there were also some undefined deposits in samples incubated in medium with the highest concentration of magnesium (MC), which were not described in previous studies [14,27]. This finding suggests a lower degree of crystallinity of the mineral formed in such samples (WPI\_U\_MC). In mineralized hydrogels incubated in medium MB (WPI\_U\_MB), besides rhombohedral crystals of calcite, there were also rod-like crystals covered with spherical deposits. Plate-like deposits, which were reported in a previous work on GG hydrogels and are characteristic of hydromagnesite, were not observed in WPI hydrogels [14].



**Figure 4.** SEM images of WPI and WPI\_U hydrogels incubated in media MA (A,D), MB (B,E) and MC (C,F). Scale bar is representative of 1 µm (A,B,C) and 10 µm (D,E,F).

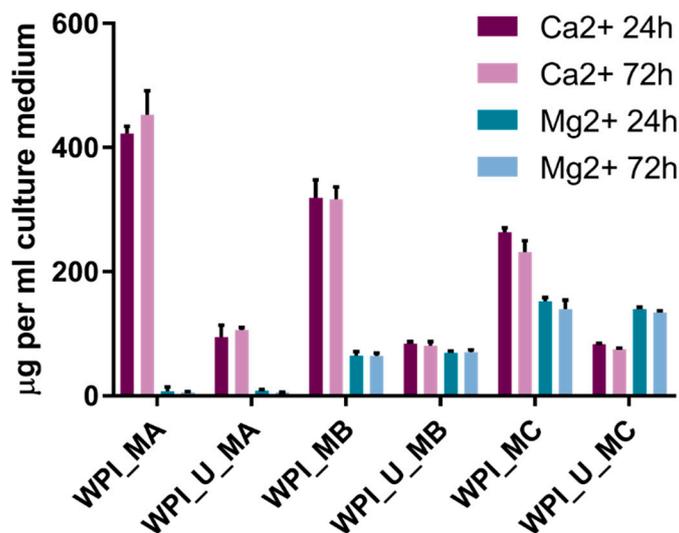
### 2.3. Cytotoxicity Studies and Release of Ca and Mg into Cell Culture Medium

Cell viability experiments utilizing the AlamarBlue assay (Figure 5) indicated that none of the hydrogels were immediately cytotoxic. Cells were viable following 24 h incubation on the WPI hydrogels. Interestingly, cell viability increased on WPI hydrogels which lacked urease where the calcium and magnesium concentrations decreased and increased, respectively (WPI\_MA vs. WPI\_MB vs. WPI\_MC). In contrast, viability remained similar for all WPI hydrogels mineralized with urease, independent of the mineralization medium used. For the most part, cell viability remained comparable after 72 h, indicating that the cells were able to adhere and remain viable.



**Figure 5.** Cell viability of mouse osteoblast (MC3T3-E1 cells) seeded onto hydrogels over 24 h and 72 h. Background fluorescence of AlamarBlue and cell culture media was subtracted from each experimental hydrogel.  $n = 3$ , error bars are representative of SD.

The results of measurements of elemental Ca and Mg in extraction medium (Figure 6) revealed that considerably more Ca and Mg were released from hydrogels which did not contain urease, i.e., which were unmineralized. One explanation may be the presence of residual mineralization medium containing Ca and Mg in the hydrogels, allowing these ions to diffuse out more easily from unmineralized samples. In mineralized samples, taking into account the lower amount of Mg present compared to Ca (see methods and materials; Table 1), Mg was preferentially released. It was noticeable that the concentrations present in the extraction medium were similar after 24 h and 72 h. This suggests that practically all Ca and Mg release takes place within the first 24 h.



**Figure 6.** Concentrations of elemental Ca and Mg in hydrogel extracts after 24 h and 72 h.  $n = 3$ , error bars show standard deviation.

### 3. Discussion

Enzymatic mineralization of WPI hydrogels was demonstrated directly by FTIR and Raman spectra (Figure 2), XRD diffractograms (Figure 3), SEM images (Figure 4), and indirectly by ICP-OES quantification of elemental magnesium and calcium and measurement of dry mass percentage (Figure 1).

The amount of mineral formed was highest when the mineralization medium contained only calcium, i.e., in sample group WPI\_U\_MA. Furthermore, when both calcium and magnesium were present in the mineralization medium, calcium was preferentially incorporated into mineral formed. These results were consistent with previous studies on urease-mediated mineralization of GG hydrogels [14]. This effect was also observed for CaP formation and can be explained by the mechanism proposed by Martin and Brown [29]. Accordingly, magnesium ions in solution are more hydrated than calcium ions, thus undergoing dehydration more slowly, which results in their adsorption on CaCO<sub>3</sub>. Subsequently, they are not included in the carbonate mineral and remain on the surface. As a result, the total amount of magnesium the mineral formed is lower.

Marine research on exoskeletons of marine invertebrates mineralized with CaCO<sub>3</sub> has revealed similar effects. In studies in which the effect of the Mg:Ca elemental ratio in seawater on the incorporation of magnesium into marine invertebrate exoskeletons and non-skeletal precipitation was investigated [30,31], preferential incorporation of calcium was reported.

A decrease in crystallinity in the order WPIU\_MA > WPIU\_MB > WPIU\_MC was observed using Raman spectroscopy (Figure 2B), XRD (Figure 3) and SEM (Figure 4). This can also be explained by the mechanism proposed by Martin and Brown (see above), whereby magnesium ions dehydrate more slowly, and hence, are not included in the carbonate mineral and remain on the surface [29]. XRD results suggested that small amounts of calcium magnesium carbonate may have formed in WPIU\_MC samples (Figure 3). Raz et al. (2000) proposed that calcium magnesium carbonate forms via an amorphous precursor phase, into which hydrated magnesium ions can be more easily incorporated in the amorphous precursor phase, and hence, are more likely to be incorporated into the final, more crystalline calcium magnesium carbonate phase [32]. Furthermore, magnesium may stabilize the amorphous precursor phase; magnesium has been reported to 'poison' calcite formation by binding to the surface of calcite nuclei and inhibiting further crystal growth as a result of its hydration [33].

SEM images (Figure 4) revealed differences in the morphologies of mineral deposits formed in WPI hydrogels in this study and those formed in GG hydrogels in previous work [14]. Hydrogels are 3D crosslinked polymer networks with entrapped water. The mechanism of WPI hydrogel formation at elevated temperature and pressure involves denaturation of its main component  $\beta$ -lg, leading to unfolding of the molecule and the formation of disulphide bonds between  $\beta$ -lg molecules and the formation of a 3D crosslinked network [34].

Differences between the structures of WPI hydrogels and GG hydrogels may influence the type of mineral formed. The concentration of polymer in WPI hydrogels in this study (50% (*w/v*)) is much higher than in the GG hydrogels used in previous works (0.7% (*w/v*)) [10,14,15]. A higher concentration of polymer would lead to smaller pores in the polymer network. In turn, this would hinder diffusion of mineralization medium through the hydrogel, which would lower the amount of mineral formed. Another consequence of small pores is hindrance of the formation of large crystals. Different degrees of compactness of the mineralized hydrogels were observed (Figure 4D–F), which may be due to the differences in the sizes of the mineral deposits. A higher concentration of polymer might however promote mineral formation; diffusion of enzyme out of the hydrogel would be impeded, and also, more polymer chains mean more potential binding sites for calcium or carbonate ions, which may serve as nucleation sites for crystal growth.

There is no obvious correlation between amount of Ca and Mg released (Figure 6) and cell proliferation (Figure 5). This demonstrates that the amounts of Ca and Mg released are non-cytotoxic and may be considered harmless. Ca concentrations of 10 mM and above have been reported to be toxic for osteoblasts, while concentrations in the range 2–4 mM have been reported to be beneficial for proliferation [35]. The Ca (and Mg) concentrations in cell culture media are in the range 0–10 mM

(Figure 6), however, no obvious positive effect was observed. It is to be expected that protein is released from WPI hydrogels; one may speculate that the released protein may be binding Ca and Mg and hindering any stimulatory effect of these ions.

Previous work on GG hydrogels mineralized with calcium and magnesium carbonates by enzymatic mineralization demonstrated that hydrogels mineralized with calcite improved the adhesion and proliferation of osteoblast-like MC3T3-E1 cells, and that hydrogels mineralized with calcite containing small amounts of magnesium had no appreciable negative effect [14]. Similarly, a previous study on GG hydrogels mineralized with calcium and magnesium carbonates by alternate soaking in solutions of calcium/magnesium and carbonate ions also demonstrated comparable proliferation and differentiation of osteoblast-like MC3T3-E1 cells on hydrogels mineralized with predominantly calcite and vaterite and those mineralized with predominantly calcite and vaterite containing small amounts of magnesium [15].

The results of this study suggest that magnesium as a dopant of CaCO<sub>3</sub> does not provide an appreciable positive biological effect on cell viability, in contrast to reports of a positive effect of magnesium as a dopant of CaP. The reasons for this remain unclear and are outside the scope of this work. In this study, no appreciable effect of mineralization on swelling was observed. However, such effects are worthy of investigation in future work.

#### 4. Materials and Methods

##### 4.1. Production of Urease WPI Hydrogels Containing Urease

WPI hydrogels were produced as previously described [2–4]. Briefly, WPI powder (Davisco, Le Sueur, MN, USA) was added to ddH<sub>2</sub>O for a final concentration of 50% (*w/v*). The solution was incubated in an ultrasonic bath for 30 min to ensure the powder had fully dissolved. Urease extracted from *Canavalia ensiformis* (Sigma Aldrich, U1500, Poole, UK) was added to WPI solution at a concentration of 10 mg urease/mL solution prior to the solution being heated to 70 °C for 8 min for gelation.

##### 4.2. Mineralization of Urease WPI Hydrogels

The urease WPI hydrogels were mineralized using three different media at room temperature over 7 days. In these media, the concentration of urea (0.17 M) was kept constant whilst the ratio of CaCl<sub>2</sub>:MgCl<sub>2</sub> was varied (Table 1). Unmineralized control samples were prepared in a similar fashion by incubating urease-free WPI hydrogels in the mineralization media.

**Table 1.** Mineralization media composition

Medium	Concentration (mol/dm <sup>3</sup> )			Ratio
	CaCl <sub>2</sub>	MgCl <sub>2</sub>	Urea	
MA	0.2700	0	0.1700	1:0
MB	0.2025	0.0675	0.1700	0.75:0.25
MC	0.1350	0.1350	0.1700	0.5:0.5

##### 4.3. Determination of Mineral Formation and Elemental Composition

To assess the extent of mineral formation, the samples were dried at 60 °C for 48 h. Samples were weighed before and after the drying process to calculate the dry mass percentage, or (weight after drying/weight before drying) × 100%, which the mass percentage is attributable to polymer and mineral and not water and serves as a measure of extent of mineralization.

Inductively coupled plasma optical emission spectroscopy (ICP-OES) was used to determine the calcium and magnesium content of samples using a 5100 Synchronous Vertical Dual View spectrometer (Agilent, Cheadle, UK). Prior to analysis, solid samples were dried and ground before being diluted in

1% nitric acid and diluted 10-fold. No additional steps were necessary for liquid samples (cell culture medium), which were diluted in 1% nitric acid immediately.

#### 4.4. Physicochemical and Morphological Characterisation: Raman, XRD, SEM, FTIR

Samples were dried at 60 °C for 48 h prior to analysis. Raman spectra were collected using a confocal Raman system (InVia, Renishaw plc, Wotton-under-Edge, UK) equipped with a near infrared laser. A laser power of ~15 mW was used to prevent any damage to the samples. Spectral collection exposure time was set to 1 s with one acquisition. Spectra were collected from each sample over the spectral range of 60–1800  $\text{cm}^{-1}$ . Pre-processing, such as baseline correction, was carried out using Wire 4.0 software (Renishaw plc, Wotton-under-Edge, UK).

X-ray diffraction (XRD) measurements were performed with a Rigaku SmartLab 9 kW with a DteX-250 detector and a Cu rotating anode source (Rigaku, Tokyo, Japan). Diffractometer was set to 45 kV and 200 mA. Five degree soller slits and 2 theta scans in Bragg–Brentano configuration were used.

Morphological characterization of hydrogels was performed using scanning electron microscopy (SEM). Firstly, samples were coated with a 9 nm layer of gold for 3 min at 20 mA and  $1 \times 10^{-1}$  mBar using a Quorum Q150RES sputter coater (Quorum Technologies Ltd, Lewes, UK). SEM analysis was performed using a secondary electron detector JSM-7800F (Jeol UK Ltd., Welwyn Garden City, UK). Images were acquired with an accelerating voltage of 5 kV at a working distance of about 10 mm.

The chemical structure of samples was examined using Fourier transform infrared spectroscopy (FTIR) (Agilent Technology, Cheadle, UK) in Attenuated Total Reflectance (ATR) mode. Spectra were collected in the 500–4000  $\text{cm}^{-1}$  spectral range with a resolution of 4  $\text{cm}^{-1}$  and an average of 8 scans.

#### 4.5. Cell Culture and Cell Viability and Release of Calcium and Magnesium from Hydrogels into Cell Culture Medium

The mouse osteoblast MC3T3-E1 cell line (ATCC<sup>®</sup> CRL-2594<sup>™</sup>, Gaithersburg, MD, USA) was routinely cultured in MEM- $\alpha$  medium supplemented with 10% FBS and incubated in a humidified 5% CO<sub>2</sub> environment at 37 °C. Once 80% confluent, cells were trypsinized and used for cytotoxicity testing. Cell tests were carried out in accordance with the International Organization for Standardization (ISO) norm ISO 10993-5.

In 96 well polystyrene plates,  $4 \times 10^3$  MC3T3-E1 cells were seeded on WPI hydrogels and incubated for 24 h to allow the cells to adhere. MC3T3-E1 seeded WPI hydrogels were washed with PBS (pH 7.4) before AlamarBlue (10% in media) was added and allowed to incubate for 2 h. Fluorescence analysis was performed using an excitation wavelength of 530 nm and an emission wavelength of 590 nm. Controls lacking cells were used to determine background fluorescence which was subsequently subtracted from cell viability results. Cell viability was measured after 24 h and 72 h.

To study release of calcium and magnesium, WPI hydrogels were incubated for 24 h or 72 h in cell culture medium under sterile conditions in the absence of cells. Calcium and magnesium concentrations in media were determined using ICP-OES, as described above

#### 4.6. Statistical Analyses

Student's *t*-test was applied to determine statistical significance using Excel software. The results of average weight, dry mass percentage, calcium and magnesium concentration and biological tests were analyzed. A two-tailed unpaired *t*-test with 95%, 99% and 99.9% confidence intervals was considered statistically significant if  $p < 0.05$  (\*),  $p < 0.01$  (\*\*) and  $p < 0.001$  (\*\*\*)

## 5. Conclusions

WPI hydrogels were enzymatically mineralized with an inorganic phase consisting of CaCO<sub>3</sub> or magnesium-enriched CaCO<sub>3</sub>. Calcium was incorporated into the mineral formed to a greater extent than magnesium. Increasing the concentration of magnesium in the mineralization medium led to a reduction in the mineral formed. These observations were confirmed by dry mass percentage and

ICP-OES. Moreover, increasing the amount of magnesium in medium resulted in a less crystalline structure of the mineral formed in hydrogels as shown by XRD and Raman spectroscopy results. The type of the carbonate phase detected in all hydrogels with urease was mainly calcite, which was confirmed by SEM morphology observation, XRD and FTIR analysis. There were also some vaterite deposits detected in all hydrogels mineralized by urease. XRD analysis showed that in hydrogels incubated in mineralization medium MC with equimolar concentrations of calcium and magnesium, some calcium magnesium carbonate was also present. Biological studies revealed that hydrogels were not cytotoxic. Hydrogels mineralized by urease had similar cell viability, which amounted to 74%. The presence of magnesium in the mineral formed did not promote or inhibit cell metabolic activity.

Further work should focus on biological studies concerning cell differentiation and in vivo implantation in order to determine the influence of CaCO<sub>3</sub> and magnesium-enriched CaCO<sub>3</sub> on bone tissue regeneration.

**Author Contributions:** Conceptualization, T.E.L.D., E.P., and A.M.T.; formal analysis, T.E.L.D., M.K., A.T., L.A., F.C., N.B., B.V.P., S.K.S., K.N.; investigation, T.E.L.D., M.K., A.T., L.A., F.C., N.B., B.V.P., S.K.S.; writing—original draft preparation, T.E.L.D., M.K., K.N.; writing—review and editing, T.E.L.D., K.N.; project administration, T.E.L.D.; funding acquisition, T.E.L.D. All authors have read and agree to the published version of the manuscript.

**Funding:** This research was funded by N8 Agrifood pump-priming grant “Food2Bone” (K.N., T.E.L.D.). The British Council is thanked for financial support in the framework of the grant “Advanced biomaterials to combat cancer” (2019-RLWK11-10353).

**Acknowledgments:** Sara Baldock and David Rochester (both Lancaster University, United Kingdom) are thanked for their excellent technical assistance.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- Douglas, T.E.L.; Vandrovцова, M.; Krocilova, N.; Keppler, J.K.; Zarubova, J.; Skirtach, A.G.; Bacakova, L. Application of whey protein isolate in bone regeneration: Effects on growth and osteogenic differentiation of bone-forming cells. *J. Dairy Sci.* **2018**, *101*, 28–36. [[CrossRef](#)] [[PubMed](#)]
- Dziadek, M.; Kudlackova, R.; Zima, A.; Slosarczyk, A.; Ziabka, M.; Jelen, P.; Shkarina, S.; Cecilia, A.; Zuber, M.; Baumbach, T. Novel multicomponent organic–inorganic WPI/gelatin/CaP hydrogel composites for bone tissue engineering. *J. Biomed. Mater. Res. Part A* **2019**, *107*, 2479–2491. [[CrossRef](#)] [[PubMed](#)]
- Dziadek, M.; Douglas, T.E.; Dziadek, K.; Zagrajczuk, B.; Serafim, A.; Stancu, I.-C.; Cholewa-Kowalska, K. Novel whey protein isolate-based highly porous scaffolds modified with therapeutic ion-releasing bioactive glasses. *Mater. Lett.* **2020**, *261*, 127115. [[CrossRef](#)]
- Gupta, D.; Kocot, M.; Tryba, A.M.; Serafim, A.; Stancu, I.C.; Jaegermann, Z.; Pamula, E.; Reilly, G.C.; Douglas, T.E. Novel naturally derived whey protein isolate and aragonite biocomposite hydrogels have potential for bone regeneration. *Mater. Design* **2020**, *188*, 108408. [[CrossRef](#)]
- Gkioni, K.; Leeuwenburgh, S.C.; Douglas, T.E.; Mikos, A.G.; Jansen, J.A. Mineralization of hydrogels for bone regeneration. *Tissue Eng. Part B Rev.* **2010**, *16*, 577–585. [[CrossRef](#)]
- Douglas, T.E.; Wlodarczyk, M.; Pamula, E.; Declercq, H.A.; de Mulder, E.L.; Bucko, M.M.; Balcaen, L.; Vanhaecke, F.; Cornelissen, R.; Dubruel, P.; et al. Enzymatic mineralization of gellan gum hydrogel for bone tissue-engineering applications and its enhancement by polydopamine. *J. Tissue Eng. Regen. Med.* **2014**, *8*, 906–918. [[CrossRef](#)]
- Fujita, Y.; Yamamuro, T.; Nakamura, T.; Kotani, S.; Ohtsuki, C.; Kokubo, T. The bonding behavior of calcite to bone. *J. Biomed. Mater. Res.* **1991**, *25*, 991–1003. [[CrossRef](#)]
- Viateau, V.; Manassero, M.; Sensebe, L.; Langonne, A.; Marchat, D.; Logeart-Avramoglou, D.; Petite, H.; Bensidhoum, M. Comparative study of the osteogenic ability of four different ceramic constructs in an ectopic large animal model. *J. Tissue Eng. Regen. Med.* **2016**, *10*, E177–E187. [[CrossRef](#)]
- Maeda, H.; Maquet, V.; Kasuga, T.; Chen, Q.Z.; Roether, J.A.; Boccaccini, A.R. Vaterite deposition on biodegradable polymer foam scaffolds for inducing bone-like hydroxycarbonate apatite coatings. *J. Mater. Sci. Mater. Med.* **2007**, *18*, 2269–2273. [[CrossRef](#)]

10. Lopez-Heredia, M.A.; Łapa, A.; Mendes, A.C.; Balcaen, L.; Samal, S.K.; Chai, F.; Van der Voort, P.; Stevens, C.V.; Parakhonskiy, B.V.; Chronakis, I.S. Bioinspired, biomimetic, double-enzymatic mineralization of hydrogels for bone regeneration with calcium carbonate. *Mater. Lett.* **2017**, *190*, 13–16. [[CrossRef](#)]
11. Borzęcka-Prokop, B.; Weselucha-Birczyńska, A.; Koszowska, E. MicroRaman, PXRD, EDS and microscopic investigation of magnesium calcite biomineral phases. The case of sea urchin biominerals. *J. Mol. Struct.* **2007**, *828*, 80–90. [[CrossRef](#)]
12. Douglas, T.E.; Krawczyk, G.; Pamula, E.; Declercq, H.A.; Schaubroeck, D.; Bucko, M.M.; Balcaen, L.; Van Der Voort, P.; Bliznuk, V.; van den Vreken, N.M.; et al. Generation of composites for bone tissue-engineering applications consisting of gellan gum hydrogels mineralized with calcium and magnesium phosphate phases by enzymatic means. *J. Tissue Eng. Regen. Med.* **2016**, *10*, 938–954. [[CrossRef](#)] [[PubMed](#)]
13. Douglas, T.E.; Lapa, A.; Reczynska, K.; Krok-Borkowicz, M.; Pietryga, K.; Samal, S.K.; Declercq, H.A.; Schaubroeck, D.; Boone, M.; Van der Voort, P.; et al. Novel injectable, self-gelling hydrogel-microparticle composites for bone regeneration consisting of gellan gum and calcium and magnesium carbonate microparticles. *Biomed. Mater.* **2016**, *11*, 065011. [[CrossRef](#)]
14. Douglas, T.E.L.; Lapa, A.; Samal, S.K.; Declercq, H.A.; Schaubroeck, D.; Mendes, A.C.; der Voort, P.V.; Dokupil, A.; Plis, A.; De Schampelaere, K.; et al. Enzymatic, urease-mediated mineralization of gellan gum hydrogel with calcium carbonate, magnesium-enriched calcium carbonate and magnesium carbonate for bone regeneration applications. *J. Tissue Eng. Regen. Med.* **2017**, *11*, 3556–3566. [[CrossRef](#)] [[PubMed](#)]
15. Lopez-Heredia, M.A.; Lapa, A.; Reczynska, K.; Pietryga, K.; Balcaen, L.; Mendes, A.C.; Schaubroeck, D.; Van Der Voort, P.; Dokupil, A.; Plis, A.; et al. Mineralization of gellan gum hydrogels with calcium and magnesium carbonates by alternate soaking in solutions of calcium/magnesium and carbonate ion solutions. *J. Tissue Eng. Regen. Med.* **2018**, *12*, 1825–1834. [[CrossRef](#)] [[PubMed](#)]
16. Geara, C. Study of the Gelation of Whey Protein Isolate by FTIR Spectroscopy and Rheological Measurements. Master Thesis, McGill University, Montreal, Canada, March 1999.
17. Gunasekaran, S.; Anbalagan, G.; Pandi, S. Raman and infrared spectra of carbonates of calcite structure. *J. Raman Spectrosc.* **2006**, *37*, 892–899. [[CrossRef](#)]
18. Sato, M.; Matsuda, S. Structure of vaterite and infrared spectra. *Z. Kristallogr. Cryst. Mater.* **1969**, *129*, 405–410. [[CrossRef](#)]
19. Alizadeh-Pasdar, N.; Nakai, S.; Li-Chan, E.C. Principal component similarity analysis of Raman spectra to study the effects of pH, heating, and  $\kappa$ -carrageenan on whey protein structure. *J. Agric. Food Chem.* **2002**, *50*, 6042–6052. [[CrossRef](#)]
20. Zhang, S.; Zhang, Z.; Lin, M.; Vardhanabhuti, B. Raman spectroscopic characterization of structural changes in heated whey protein isolate upon soluble complex formation with pectin at near neutral pH. *J. Agric. Food Chem.* **2012**, *60*, 12029–12035. [[CrossRef](#)]
21. Nonaka, M.; Li-Chan, E.; Nakai, S. Raman spectroscopic study of thermally induced gelation of whey proteins. *J. Agric. Food Chem.* **1993**, *41*, 1176–1181. [[CrossRef](#)]
22. Farmer, V.C. *Infrared Spectra of Minerals*; Mineralogical Society of Great Britain and Ireland: London, UK, 1974.
23. Douglas, T.E.; Sobczyk, K.; Łapa, A.; Włodarczyk, K.; Brackman, G.; Vidiashva, I.; Reczyńska, K.; Pietryga, K.; Schaubroeck, D.; Bliznuk, V. Ca:Mg:Zn:CO<sub>3</sub> and Ca:Mg:CO<sub>3</sub>—tri- and bi-elemental carbonate microparticles for novel injectable self-gelling hydrogel-microparticle composites for tissue regeneration. *Biomed. Mater.* **2017**, *12*, 025015. [[CrossRef](#)] [[PubMed](#)]
24. Bischoff, W.D.; Sharma, S.K.; MacKenzie, F.T. Carbonate ion disorder in synthetic and biogenic magnesian calcites: A Raman spectral study. *Am. Mineral.* **1985**, *70*, 581–589.
25. Landi, E.; Logroscino, G.; Proietti, L.; Tampieri, A.; Sandri, M.; Sprio, S. Biomimetic Mg-substituted hydroxyapatite: From synthesis to in vivo behaviour. *J. Mater. Sci. Mater. Med.* **2008**, *19*, 239–247. [[CrossRef](#)] [[PubMed](#)]
26. Chen, Z.; Xin, M.; Li, M.; Xu, J.; Li, X.; Chen, X. Biomimetic synthesis of coexistence of vaterite-calcite phases controlled by histidine-grafted-chitosan. *J. Cryst. Growth* **2014**, *404*, 107–115. [[CrossRef](#)]
27. Bermanec, V.; Posilović, H.; Žigovečki Gobac, Ž. Identification of biogenetic calcite and aragonite using SEM. *Geol. Croat.* **2009**, *62*, 201–206.
28. Yang, D.; Qi, L.; Ma, J. Well-defined star-shaped calcite crystals formed in agarose gels. *Chem. Commun.* **2003**, 1180–1181. [[CrossRef](#)] [[PubMed](#)]

29. Martin, R.; Brown, P. The effects of magnesium on hydroxyapatite formation in vitro from  $\text{CaHPO}_4$  and  $\text{Ca}_4(\text{PO}_4)_2\text{O}$  at 37.4 °C. *Calcif. Tissue Int.* **1997**, *60*, 538–546. [[CrossRef](#)] [[PubMed](#)]
30. Stanley, S.M.; Ries, J.B.; Hardie, L.A. Low-magnesium calcite produced by coralline algae in seawater of Late Cretaceous composition. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 15323–15326. [[CrossRef](#)]
31. Ries, J.B. Effect of ambient Mg/Ca ratio on Mg fractionation in calcareous marine invertebrates: A record of the oceanic Mg/Ca ratio over the Phanerozoic. *Geology* **2004**, *32*, 981–984. [[CrossRef](#)]
32. Raz, S.; Testeniere, O.; Hecker, A.; Weiner, S.; Luquet, G. Stable amorphous calcium carbonate is the main component of the calcium storage structures of the crustacean *Orchestia cavimana*. *Biol. Bull.* **2002**, *203*, 269–274. [[CrossRef](#)]
33. Folk, R.L. The natural history of crystalline calcium carbonate; effect of magnesium content and salinity. *J. Sediment. Res.* **1974**, *44*, 40–53.
34. Orlien, V. Utilizing High Pressure Processing to Induce Structural Changes in Dairy and Meat Products. In *Reference Module in Food Science*; Elsevier: Amsterdam, The Netherlands, 2017.
35. Maeno, S.; Niki, Y.; Matsumoto, H.; Morioka, H.; Yatabe, T.; Funayama, A.; Toyama, Y.; Taguchi, T.; Tanaka, J. The effect of calcium ion concentration on osteoblast viability, proliferation and differentiation in monolayer and 3D culture. *Biomaterials* **2005**, *26*, 4847–4855. [[CrossRef](#)] [[PubMed](#)]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).