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A real-world comparison of tisagenlecleucel and axicabtagene ciloleucel CAR T cells in relapsed or refractory diffuse large B cell lymphoma

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Axicabtagene ciloleucel (axi-cel) and tisagenlecleucel (tisa-cel) have both demonstrated impressive clinical activity in relapsed/refractory (R/R) diffuse large B cell lymphoma (DLBCL). In this study, we analyzed the outcome of 809 patients with R/R DLBCL after two or more previous lines of treatment who had a commercial chimeric antigen receptor (CAR) T cells order for axi-cel or tisa-cel and were registered in the retrospective French DESCAR-T registry study (NCT04328298). After 1:1 propensity score matching ($n = 418$), the best overall response rate/complete response rate (ORR/CRR) was 80%/60% versus 66%/42% for patients treated with axi-cel compared to tisa-cel, respectively ($P < 0.001$ for both ORR and CRR comparisons). After a median follow-up of 11.7 months, the 1-year progression-free survival was 46.6% for axi-cel and 33.2% for tisa-cel (hazard ratio (HR) = 0.61; 95% confidence interval (CI), 0.46–0.79; $P = 0.0003$). Overall survival (OS) was also significantly improved after axi-cel infusion compared to after tisa-cel infusion (1-year OS 63.5% versus 48.8%; HR = 0.63; 95% CI, 0.45–0.88; $P = 0.0072$). Similar findings were observed using the inverse probability of treatment weighting statistical approach. Grade 1–2 cytokine release syndrome was significantly more frequent with axi-cel than with tisa-cel, but no significant difference was observed for grade ≥ 3 . Regarding immune effector cell-associated neurotoxicity syndrome (ICANS), both grade 1–2 and grade ≥ 3 ICANS were significantly more frequent with axi-cel than with tisa-cel. In conclusion, our matched comparison study supports a higher efficacy and also a higher toxicity of axi-cel compared to tisa-cel in the third or more treatment line for R/R DLBCL.

DLBCL is the most common lymphoma subtype, accounting for about 40% of all non-Hodgkin lymphomas¹. CAR T cell therapies targeting CD19 have shown impressive efficacy and manageable toxicity for the treatment of various lymphoma histology subtypes, such as mantle cell lymphoma, follicular lymphoma and DLBCL^{2–7}. Tisagenlecleucel (tisa-cel) and axicabtagene ciloleucel (axi-cel) are two CAR T products that were initially

approved for the treatment of DLBCL in the third or subsequent line of treatment. Tisa-cel is a 4-1BB co-stimulatory domain-based second-generation CAR T, whereas axi-cel is CD28 based. Approvals were granted after the results of the JULIET and ZUMA-1 pivotal studies demonstrating best ORR/CRR of 52%/40% and 82%/58% for tisa-cel and axi-cel, respectively^{5,6,8,9}. The recent updated follow-up of ZUMA-1 after 5 years suggested that ~40% of patients might be

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cured with CAR T in this setting¹⁰. In the last 2 years, many publications based on real-life data from various countries worldwide have confirmed the high response rates, prolonged response duration and survival achieved with CAR T in DLBCL^{11–15}. Strikingly, and despite stringent patient selection in clinical trials, efficacy in the non-trial setting seems to parallel results obtained in pivotal studies, and toxicity appears significantly lower in real life due to the earlier mitigating strategy with anti-interleukin-6 and steroids use^{16,17}. A multitude of parameters can impact efficacy and safety of CAR T, such as, among many others, the use of a bridging therapy to control for disease progression during product manufacturing, the tumor bulk or the delay between leukapheresis and infusion^{18,19}. Therefore, the need for real-world evidence (RWE) studies to apprehend this fast-moving field has never been so high.

Crude response rates and safety reports from clinical trials suggest higher efficacy and toxicity associated with the use of axi-cel compared to tisa-cel^{5,6}. However, these conclusions might be misleading due to large differences between study designs: (1) patients with primary mediastinal B cell lymphoma (PMBCL) were enrolled in ZUMA-1 but not in JULIET; (2) the doses of fludarabine and cyclophosphamide as conditioning regimen were higher in ZUMA-1; and (3) bridging chemotherapy to control for disease progression during the CAR T manufacturing process was allowed in JULIET but not in ZUMA-1 (refs. 5,6). The latter introduced a major bias precluding any possible direct comparison between studies because patients with more aggressive lymphomas cannot usually be spared from bridging therapy between leukapheresis and lymphodepletion.

Several matching-adjusted indirect comparisons (MAICs) have been attempted to compare different CAR T products^{20,21}. MAIC uses individual patient data (IPD) from one study and trial-level data from another to form a population-adjusted indirect comparison between treatments. One of these recently reported MAICs suggests that axi-cel is superior to tisa-cel for disease control but is associated with significantly more toxicity²⁰. In addition, despite increasing popularity, many biases remain with such statistical methods^{22–24}.

Since 2019, the French Health Authorities have required extensive data collection for each patient with a theoretical indication of CAR T treatment. Reimbursement is conditional on data comprehensive completion by the local investigator. The DESCAR-T registry has been set up by the Lymphoma Study Association (LYSA) and the Lymphoma Academic Research Organization (LYSARC) to fulfil this regulatory request and to allow for comprehensive RWE studies.

Given the lack of an adequate comparison for efficacy and safety between tisa-cel and axi-cel, we embarked on an IPD-based matched comparison considering all French patients with DLBCL treated with commercial CAR T and included in the DESCAR-T registry.

Results

Patient characteristics and outcome. Between December 2019 and October 2021, 809 patients from 23 French centers with R/R DLBCL after at least two lines of previous therapy had a commercial CAR T order for axi-cel or tisa-cel and were registered in DESCAR-T (Fig. 1). Patient characteristics are presented in Table 1. Median age was 63 years (range, 19–81 years), and 61% of patients were male. Median number of prior lines of treatment was three (range, 2–10), and 21% of patients had received a prior stem cell transplant (SCT). The median time between the end date of last treatment and CAR T order was 35 days (Q1;Q3, 15;78 days). Most patients ($n=604$, 75%) had DLBCL not otherwise specified (NOS) or high-grade B cell lymphoma (HGBCL); 127 patients (16%) had transformed follicular lymphoma (tFL); 35 patients (4%) had primary mediastinal large B cell lymphoma (PMBCL); and 24 patients (3%) had transformed marginal zone lymphoma (tMZL). Few patients ($n=19$; 2%) had

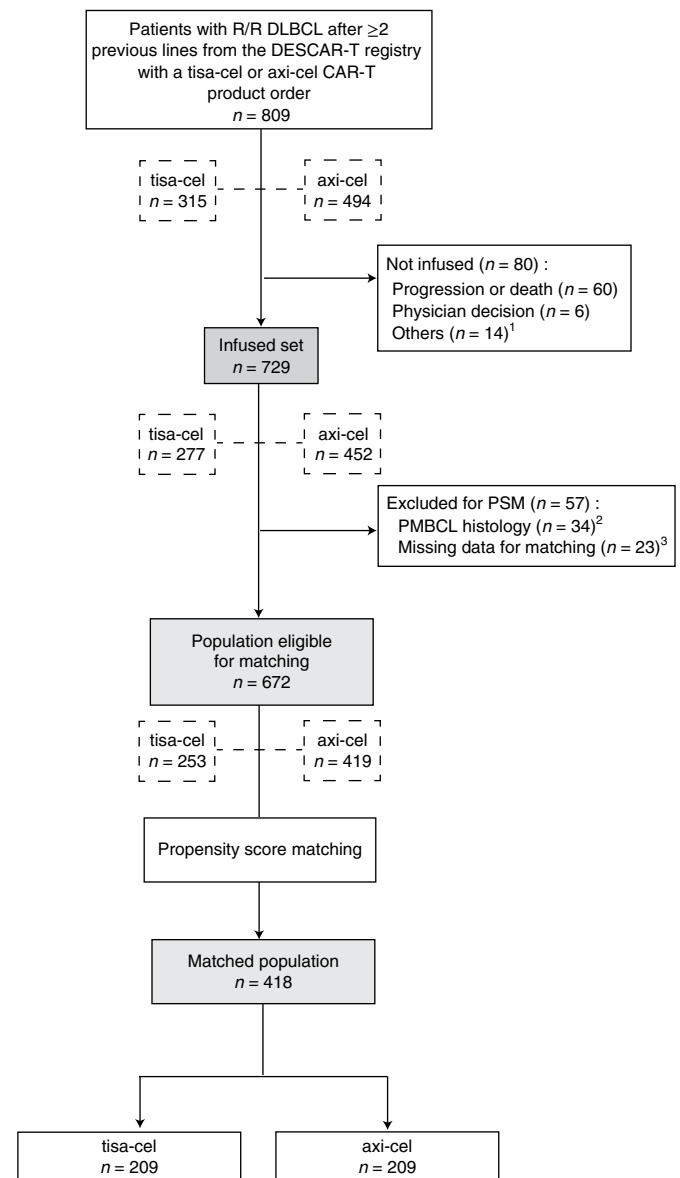


Fig. 1 | Patient flow diagram for PSM analysis. ¹Manufacturing failure ($n=3$), uncontrolled infection ($n=3$), waiting for infusion ($n=3$), patient decision ($n=1$), leukapheresis failure ($n=1$), acute coronary syndrome ($n=1$), concomitant malignancy ($n=1$) and progression of another malignancy ($n=1$). ²Patients with PMBCL histology were excluded because tisa-cel has no approval for this histology. ³Patients with $\geq 25\%$ of missing data for matching covariates were removed from the matching step.

other histologies (T cell/histiocyte-rich large B cell lymphoma (T/HRLBCL) in 11 patients; systemic relapse of primary central nervous system lymphoma (PCNSL) in four patients; and DLBCL, leg type, in four patients) (Table 1). With a median follow-up of 13 months (95% CI, 12.1–13.5 months), projected median OS was 17 months (95% CI, 13.3–21.1 months) from CAR T order (Fig. 2a).

Sixty patients out of 809 with a CAR T product order progressed or died between leukapheresis and lymphodepletion, and 20 did not proceed to lymphodepletion after physician decision or for other reasons (Fig. 1). Of these 80 patients with a CAR T product order who did not proceed until an infusion ($n=38$ for tisa-cel and $n=42$ for axi-cel), OS was expectedly very poor and similar according to CAR T product (axi-cel or tisa-cel) ($P=0.48$; Extended Data Fig. 1). Finally, 729 patients proceeded to lymphodepletion and

Table 1 | Patient characteristics

	All patients*				Before PSM [‡]				After PSM [‡]			
	Order set		Infusion set		axi-cel		tisa-cel		axi-cel		tisa-cel	
	n = 809		n = 729		n = 419		n = 253		n = 209		n = 209	
Age at time of CAR T order (years)												
Median (min;max)	63 (19;81)		63 (19;81)		63 (19;79)		64 (20;81)		62 (20;79)		64 (20;81)	
Missing	1		1		0		0		0		0	
Sex												
Male	490	(60.6%)	437	(59.9%)	251	(59.9%)	157	(62.1%)	121	(57.9%)	126	(60.3%)
Female	318	(39.3%)	291	(39.9%)	168	(40.1%)	96	(37.9%)	88	(42.1%)	83	(39.7%)
Missing	1	(0.1%)	1	(0.1%)	0	(0.0%)	0	(0.0%)	0	(0.0%)	0	(0.0%)
aalPI												
0	54	(6.7%)	52	(7.1%)	31	(7.4%)	18	(7.1%)	17	(8.1%)	16	(7.7%)
1	237	(29.3%)	224	(30.7%)	135	(32.2%)	69	(27.3%)	71	(34.0%)	56	(26.8%)
2	373	(46.1%)	336	(46.1%)	190	(45.3%)	126	(49.8%)	89	(42.6%)	105	(50.2%)
3	58	(7.2%)	40	(5.5%)	19	(4.5%)	20	(7.9%)	11	(5.3%)	16	(7.7%)
Missing	87	(10.8%)	77	(10.6%)	44	(10.5%)	20	(7.9%)	21	(10.0%)	16	(7.7%)
ECOG PS												
0-1	665	(82.2%)	613	(84.1%)	361	(86.2%)	208	(82.2%)	178	(85.2%)	173	(82.8%)
≥2	97	(12.0%)	75	(10.3%)	39	(9.3%)	33	(13.0%)	20	(9.6%)	27	(12.9%)
Missing	47	(5.8%)	41	(5.6%)	19	(4.5%)	12	(4.7%)	11	(5.3%)	9	(4.3%)
CRP [†]												
≤30 mg L ⁻¹	-		521	(71.5%)	313	(74.7%)	175	(69.2%)	150	(71.8%)	147	(70.3%)
>30 mg L ⁻¹			165	(22.6%)	92	(22.0%)	65	(25.7%)	49	(23.4%)	55	(26.3%)
Missing			43	(5.9%)	14	(3.3%)	13	(5.1%)	10	(4.8%)	7	(3.3%)
LDH [†]												
≤ULN	-		311	(42.7%)	174	(41.5%)	116	(45.8%)	85	(40.7%)	83	(39.7%)
[ULN; 2× ULN]			286	(39.2%)	177	(42.2%)	96	(37.9%)	85	(40.7%)	88	(42.1%)
>2× ULN			87	(11.9%)	50	(11.9%)	30	(11.9%)	30	(14.4%)	29	(13.9%)
Missing			45	(6.2%)	18	(4.3%)	11	(4.3%)	9	(4.3%)	9	(4.3%)
Bulk (with a cutoff at 5 cm) [†]												
No	-		551	(75.6%)	326	(77.8%)	198	(78.3%)	168	(80.4%)	160	(76.6%)
Yes			150	(20.6%)	85	(20.3%)	51	(20.2%)	39	(18.7%)	45	(21.5%)
Missing			28	(3.8%)	8	(1.9%)	4	(1.6%)	2	(1.0%)	4	(1.9%)
Ann Arbor stage												
I	57	(7.0%)	55	(7.5%)	31	(7.4%)	16	(6.3%)	18	(8.6%)	16	(7.7%)
II	90	(11.1%)	85	(11.7%)	51	(12.2%)	25	(9.9%)	26	(12.4%)	22	(10.5%)
III	100	(12.4%)	92	(12.6%)	63	(15.0%)	25	(9.9%)	29	(13.9%)	24	(11.5%)
IV	513	(63.4%)	453	(62.1%)	249	(59.4%)	180	(71.1%)	126	(60.3%)	140	(67.0%)
Missing	49	(6.1%)	44	(6.0%)	25	(6.0%)	7	(2.8%)	10	(4.8%)	7	(3.3%)
Number of prior treatment lines												
Median (min;max)	3 (2;10)		3 (2;10)		3 (2;9)		3 (2;10)		2 (2;8)		2 (2;10)	
Missing	10		9		0		0		0		0	
At least one prior transplant												
No	640	(79.1%)	567	(77.8%)	332	(79.2%)	187	(73.9%)	160	(76.6%)	163	(78.0%)
Yes	169	(20.9%)	162	(22.2%)	87	(20.8%)	66	(26.1%)	49	(23.4%)	46	(22.0%)
Missing	0		0		0		0		0		0	
Time between first CAR T order of center and CAR T order of patient (days) [‡]												
Median (Q1;Q3)	446 (214;681)		446 (206;671)		420 (169;681)		485 (316;662)		517 (174;724)		495 (317;664)	
Missing	0		0		0		0		0		0	

Continued

Table 1 | Patient characteristics (continued)

	All patients ^a				Before PSM ^a				After PSM ^a			
	Order set		Infusion set		axi-cel		tisa-cel		axi-cel		tisa-cel	
	n = 809	n = 729	n = 419	n = 253	n = 209	n = 209						
Time between end of last treatment and CAR T infusion (days) ^b												
Median (Q1;Q3)	35 (15;78)	87 (66;138)	90 (68;146)	87 (66;133)	91 (71;132)	92 (68;147)						
Missing	17	16	0	0	0	0						
Bridging and response to bridging												
No bridging	NA	126 (17.3%)	76 (18.1%)	35 (13.8%)	26 (12.4%)	29 (13.9%)						
Response to bridging (PR or CR)		188 (25.8%)	105 (25.1%)	72 (28.5%)	65 (31.1%)	57 (27.3%)						
No response to bridging (SD or PD)		386 (52.9%)	221 (52.7%)	138 (54.5%)	111 (53.1%)	117 (56.0%)						
Missing		29 (4.0%)	17 (4.1%)	8 (3.2%)	7 (3.3%)	6 (2.9%)						
Histological diagnosis												
DLBCL NOS or HGBCL	604 (74.7%)	542 (74.3%)	328 (78.2%)	193 (76.3%)	165 (78.9%)	166 (79.4%)						
T/HRLBCL	11 (1.3%)	10 (1.4%)	7 (1.7%)	3 (1.2%)	1 (0.5%)	2 (1.0%)						
DLBCL after PCNSL	4 (0.5%)	4 (0.5%)	1 (0.2%)	3 (1.2%)	0 (0.0%)	1 (0.5%)						
DLBCL, leg type	4 (0.5%)	4 (0.5%)	2 (0.5%)	2 (0.8%)	1 (0.5%)	0 (0.0%)						
PMBCL ^c	35 (4.3%)	34 (4.7%)	NA	NA	NA	NA						
tFL	127 (15.7%)	117 (16.0%)	71 (16.9%)	44 (17.4%)	37 (17.7%)	33 (15.8%)						
tMZL	24 (3.0%)	18 (2.5%)	10 (2.4%)	8 (3.2%)	5 (2.4%)	7 (3.3%)						
Missing	0	0	0	0	0	0						

^aSum may not equal 100% because of rounding. ^bCRP, LDH and bulk were assessed at time of lymphodepletion. ^cExcept for the order set where time between the last treatment and the CAR T order was considered. ^dPMBCL was not considered for PSM because tisa-cel is not approved for this histology. ^eTime between first CAR T order of center and CAR T order of patient was used as a surrogate for center experience for CAR T therapy for each given patient. ^fPatients from the infusion set with more than 25% of missing data and with PMBCL were excluded for the matching procedure. CR, complete response; NA, not applicable; Q1, first quartile; Q3, third quartile; PD, progressive disease; PR, partial response; SD, stable disease.

CAR T infusion. Characteristics of this patient subpopulation are presented in Table 1. Median OS from infusion was 19.0 months (95% CI, 15.2–not reached), and median progression-free survival (PFS) was 5.6 months (95% CI, 4.1–7.5 months) (Fig. 2a,b).

Propensity score matching. A propensity score is the conditional probability that a patient receives one treatment or another given a set of observed covariates. The aim of propensity score matching (PSM) was to balance covariates between axi-cel and tisa-cel groups to account for all possible measured confounding variables (that is, variables that have a causal relationship with both the measured outcome and the CAR T product used) (Fig. 3a). For PSM, of 729 patients infused with a CAR T product, 34 patients with PMBCL (for which tisa-cel is not approved) and 23 patients with more than 25% of missing data for matching variables were removed before matching (Fig. 1). The final population for matching comprised 253 patients treated with tisa-cel and 419 patients treated with axi-cel. Patient characteristics according to CAR T product are detailed in Table 1. Univariate prognostic analyses for PFS and OS confirmed that many patient characteristics were significantly associated with outcome and were potential confounders when comparing efficacy of CAR T products (Extended Data Fig. 2). After stringent PSM on 14 parameters (Extended Data Fig. 3a), absolute values of the standardized mean differences (SMDs) were less than 0.1 for almost all matching covariates (Extended Data Fig. 3b,c). PSM resulted in a much balanced distribution of CAR T product use across centers (Extended Data Fig. 3d) and according to individual covariates (Extended Data Fig. 3e,f). However, disease severity was still slightly higher for patients treated with tisa-cel than with axi-cel, as exemplified by a higher age-adjusted international prognostic index

(aaIPI) score of 2 or 3 (57.9% versus 47.9%). In the 1:1 matched population ($n=418$; 209 patients treated with axi-cel and 209 patients treated with tisa-cel), the best ORR/CRR was 80.4%/60.3% versus 66.0%/42.1% for patients treated with axi-cel compared to tisa-cel, respectively ($P<0.001$ for both ORR and CRR comparisons; Table 2). After a median follow-up of 11.7 months (95% CI, 10.5–12.0 months), the duration of response (DOR) was not significantly different between axi-cel and tisa-cel (1-year DOR 53.8% for axi-cel compared to 41.8% for tisa-cel, $P=0.106$; Fig. 3b). There was no further significant difference in DOR according to the quality of response (complete versus partial) (Fig. 3c). However, the 1-year PFS was 46.6% for axi-cel and 33.2% for tisa-cel (HR=0.61; 95% CI, 0.46–0.79; $P=0.0003$; Fig. 3d and Table 2). OS was also significantly improved after axi-cel infusion compared to after tisa-cel infusion (1-year OS 63.5% versus 48.8%; HR=0.63; 95% CI, 0.45–0.88; $P=0.0072$; Fig. 3e and Table 2).

Inverse probability of treatment weighting. Inverse probability of treatment weighting (IPTW) is another method where weights are assigned to patients based on the inverse probability of receiving one treatment or the other as estimated by the propensity score. IPTW results in a pseudo-population that is balanced regarding the distribution of patient covariates in each treatment group. After IPTW, absolute values of the SMDs were less than 0.1 for almost all matching covariates (Extended Data Fig. 3g,h). IPTW was used to support the findings of PSM analysis and to allow for proper comparison between the two populations of patients treated with axi-cel or tisa-cel. Using this statistical approach, significantly higher ORR/CRR and longer PFS and OS with axi-cel compared to tisa-cel were observed (Table 2 and

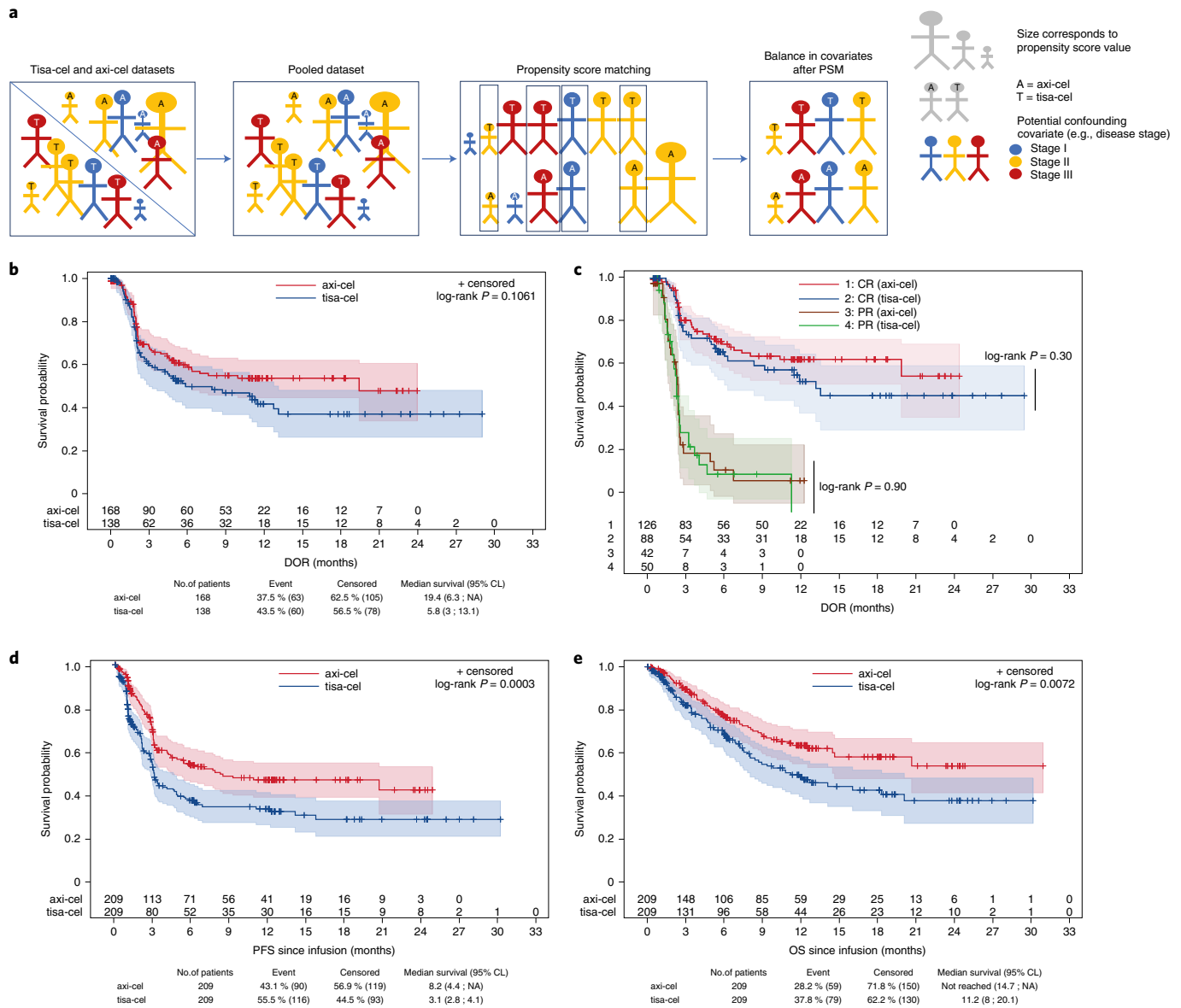


Fig. 3 | Survival according to CAR T product after PSM. a, Propensity score reflects the probability of receiving tisa-cel or axi-cel conditional on an exhaustive list of 14 pre-infusion covariates. PSM is based on matching patients with similar propensity score. Comparability according to each covariate (for instance, disease stage depicted here) of the resulting two sub-cohorts of patients receiving one CAR T or the other is checked using SMDs (Extended Data Fig. 3). **b**, DOR according to CAR T product (axi-cel, $n=168$, red line; tisa-cel, $n=138$, blue line) ($P=0.11$). **c**, DOR according to CAR T product and response quality (complete response (CR) versus partial response (PR); $P=0.30$ and $P=0.90$, respectively) (axi-cel and CR, $n=126$, red line; tisa-cel and CR, $n=88$, blue line; axi-cel and PR, $n=42$, brown line; tisa-cel and PR, $n=50$, green line). **d**, PFS according to CAR T product ($P=0.0003$). **e**, OS according to CAR T product ($P=0.0072$). P values were calculated using two-sided log-rank tests. No adjustment was made for multiple comparisons. Shaded areas correspond to the 95% confidence bands using the Hall-Wellner method. CL, confidence limit; NA, not assessable.

to 19.1% after tisa-cel infusion ($P<0.001$). Twenty-nine patients (13.9%) presented a grade ≥ 3 ICANS with axi-cel compared to only six (2.9%) with tisa-cel ($P<0.001$).

Hematological toxicity was also significantly more frequent and more severe with axi-cel than with tisa-cel (Table 3). Any grade cytopenia at 1 month after CAR T infusion was observed in 64.6% of patients compared to 39.2% and grade ≥ 3 cytopenia in 34.0% compared to 12.4% with axi-cel and tisa-cel, respectively (Table 3). Significantly higher hematological toxicity after axi-cel infusion compared to after tisa-cel infusion was consistent across all hematological lineages (that is, neutropenia, anemia and thrombocytopenia; Table 3). The same held true for prolonged cytopenias observed at 3 months after CAR T infusion (Table 3). Notably, no significant

difference in cytopenias was observed before lymphodepletion between patients treated with axi-cel or with tisa-cel, meaning that the observed higher hematological toxicity with axi-cel was not attributable to significant baseline differences.

No grade 5 CRS deemed related to axi-cel was noted compared to two with tisa-cel in the matched-population. One grade 5 ICANS was reported with axi-cel but none with tisa-cel. No other grade 5 adverse event directly associated with CAR T infusion was reported in the matched populations.

Subgroup analyses. Two subgroup analyses were originally planned. First, outcome according to age category (that is, ≤ 70 years and >70 years) was assessed in the PSM population. PFS was

Table 2 | Response rates and survival according to CAR T product in matched populations using PSM and IPTW approaches^a

	PSM			IPTW		
	axi-cel <i>n</i> = 209	tisa-cel <i>n</i> = 209	<i>P</i>	axi-cel	tisa-cel	<i>P</i>
Response rate						
ORR% (95% CI)	80.4 (74.3–85.5)	66.0 (59.2–72.4)	<0.001	78.5 (75.3–81.6)	62.8 (59.2–66.3)	<0.001
CRR% (95% CI)	60.3 (53.3–67.0)	42.1 (35.3–49.1)	<0.001	60.1 (56.4–63.8)	42.0 (38.3–45.6)	<0.001
Survival						
PFS% at 1 year (95% CI)	46.6 (38.5–54.3)	33.2 (25.7–40.8)	0.0003	44.5 (38.7–50.1)	34.7 (26.2–43.3)	0.0005
HR (95% CI)	0.61 (0.46–0.79)	1 (ref)		0.66 (0.53–0.82)	1 (ref)	
DOR% at 1 year (95% CI)	53.8 (44.7–62.1)	41.8 (31.3–51.9)	0.106	51.0 (44.4–57.1)	46.0 (33.0–58.0)	0.482
HR (95% CI)	0.75 (0.53–1.06)	1 (ref)		0.81 (0.60–1.08)	1 (ref)	
OS% at 1 year (95% CI)	63.5 (55.0–70.8)	48.8 (39.7–57.2)	0.0072	61.2 (55.1–66.6)	48.3 (37.1–58.5)	0.011
HR (95% CI)	0.63 (0.45–0.88)	1 (ref)		0.71 (0.54–0.93)	1 (ref)	

^aThe response was assessed according to the local investigators per Lugano 2014 criteria, and the best response throughout patient follow-up was reported.

significantly longer after axi-cel infusion than after tisa-cel infusion both in patients aged 70 years or younger and in patients older than 70 years. Median PFS was 5.9 months compared to 3.1 months for axi-cel and tisa-cel, respectively, for patients ≤ 70 years ($P=0.0128$) and was not reached compared to 3 months, respectively, for >70 years ($P=0.0026$) (Extended Data Fig. 5a,b). For OS, survival was longer with axi-cel compared to tisa-cel in both age categories similarly, although statistical significance was not reached in patients ≤ 70 years ($P=0.0779$ in the ≤ 70 -years group and $P=0.0167$ in the >70 -years group, respectively) (Extended Data Fig. 5c,d). Second, because CAR T potency in the case of high tumor bulk could depend on the type of co-stimulatory domain, efficacy was evaluated according to the longest diameter of the largest node or extranodal mass taken as a correlate of the tumor bulk (that is, ≤ 5 cm or >5 cm). PFS was significantly longer regardless of tumor bulk after axi-cel infusion compared to after tisa-cel infusion. In the absence of a bulky mass, median PFS was 7.9 months with axi-cel and 3.5 months with tisa-cel ($P=0.0164$). In the presence of a bulky disease, median PFS was 8.2 months with axi-cel and 2.1 months with tisa-cel ($P=0.0023$) (Extended Data Fig. 5e,f). Better outcome with axi-cel than with tisa-cel regardless of tumor bulk held true for OS (Extended Data Fig. 5g,h).

Sensitivity analyses. To ensure the robustness of comparison of results, several sensitivity analyses were performed. First, PSM and efficacy analyses were carried out on a subpopulation of patients with no missing data for any matching parameter. In total, 174 patients treated with axi-cel were 1:1 matched with 174 patients treated with tisa-cel (Extended Data Table 1). Similar results were found regarding ORR/CRR (Extended Data Table 2), DOR, PFS and OS (Extended Data Fig. 6a–f), with a superior efficacy of axi-cel compared to tisa-cel using both PSM and IPTW approaches. Apart from considering missing data as a category (missing indicator method) or from removing missing data (complete case analysis), multiple imputation approach on ten simulated datasets was also used and found similarly that patients treated with axi-cel experienced significantly prolonged PFS (HR=0.64; 95% CI, 0.49–0.83) and OS (HR=0.70; 95% CI, 0.51–0.97) (Extended Data File 1). Furthermore, PSM and IPTW comparisons for OS were performed from CAR T order instead of CAR T infusion to avoid biases due to the manufacturing process. OS from CAR T order was significantly longer with axi-cel than with tisa-cel using both PSM or IPTW ($P=0.038$ and $P=0.012$, respectively; Extended Data Fig. 7a,b). Because a residual imbalance of adverse prognosis factors remained for patient treated with tisa-cel after stringent matching on 14

parameters, bivariate Cox analyses with CAR T product and aaPI as explanatory variables were performed. Significantly prolonged PFS and OS were still associated with axi-cel compared to tisa-cel (HR=0.64 and $P=0.012$ for PFS and HR=0.61 and $P<0.001$ for OS, respectively).

Despite exhaustive matching on known and measured confounding factors, an unmeasured confounder can still lead to erroneous conclusions. For PFS and OS, the E-values were 2.18 (lower limit (LL) of the CI, 1.6) and 2.09 (LL of the CI, 1.39), respectively, meaning that the observed difference for PFS and OS between axi-cel and tisa-cel could be explained away only by an unmeasured confounder that was associated with both CAR T products and PFS (or OS) by a risk ratio of more than 2.18-fold each for PFS (or 2.09-fold each for OS).

Discussion

In the present study, 809 patients for whom a CAR T order was obtained outside of a trial setting for DLBCL in second or subsequent relapse were analyzed. Median OS from CAR T order and CAR T infusion was 17 months and 19 months, respectively, for the whole cohort of patients. Strikingly, in the 1:1 matched population of 418 patients considered after the stringent PSM statistical approach, ORR/CRR were 66%/42% for tisa-cel and 80%/60% for axi-cel, which mirror response rates in the two pivotal clinical trials: JULIET and ZUMA-1 (52%/40% and 82%/58%, respectively)^{5,6,8,9}. Similarly, median OS was 11.2 months with tisa-cel, whereas median OS was not reached with axi-cel, echoing the 11.1 months and 25.8 months of median OS in the recent updates of the JULIET and ZUMA-1 trials, respectively.

RWE studies are of utmost importance to assess if trial conclusions are reproducible in routine practice and if they can be applied to a more diverse patient population than the one strictly limited to pivotal trial enrollment criteria. Furthermore, RWE studies provide a critical basis from which to conduct cross-comparison analyses based on IPD. PSM and IPTW are increasingly used to address confounding by indication in RWE studies. The objectives of these statistical approaches are to balance out differences between patient groups that can be substantial and that preclude drawing firm conclusions when comparing outcome measurements. Subtle differences exist between the two methods that have been extensively reviewed elsewhere²⁵. In the present study, both techniques similarly concluded that axi-cel provides better disease control than tisa-cel in R/R DLBCL after two lines of previous therapy.

After stringent matching to control for slightly more aggressive disease features in the patient population treated with tisa-cel (more

Table 3 | Toxicity after CAR T infusion according to CAR T product in the PSM cohorts

	axi-cel		tisa-cel		P
	n = 209		n = 209		
CRS of any grade	180	(86.1%)	158	(75.6%)	0.006
Grade 1–2	169	(80.9%)	139	(66.5%)	<0.001
Grade ≥3	11	(5.3%)	19	(9.1%)	0.130
ICANS of any grade	102	(48.8%)	46	(22.0%)	<0.001
Grade 1–2	73	(34.9%)	40	(19.1%)	<0.001
Grade ≥3	29	(13.9%)	6	(2.9%)	<0.001
Cytopenia of any grade at M1	135	(64.6%)	82	(39.2%)	<0.001
Grade 1–2	64	(30.6%)	56	(26.8%)	0.387
Grade ≥3	71	(34.0%)	26	(12.4%)	<0.001
Neutropenia of any grade at M1	124	(59.3%)	57	(27.3%)	<0.001
Grade 1–2	71	(34.0%)	37	(17.7%)	<0.001
Grade ≥3	53	(25.4%)	20	(9.6%)	<0.001
Anemia of any grade at M1	94	(45.0%)	58	(27.8%)	<0.001
Grade 1–2	90	(43.1%)	58	(27.8%)	0.001
Grade ≥3	4	(1.9%)	0	(0.0%)	0.044
Thrombocytopenia of any grade at M1	116	(55.5%)	62	(29.7%)	<0.001
Grade 1–2	70	(33.5%)	43	(20.6%)	0.003
Grade ≥3	46	(22.0%)	19	(9.1%)	<0.001
Cytopenia of any grade at M3	75	(35.9%)	29	(13.9%)	<0.001
Grade 1–2	51	(24.4%)	21	(10.0%)	<0.001
Grade ≥3	24	(11.5%)	8	(3.8%)	0.003
Neutropenia of any grade at M3	62	(29.7%)	22	(10.5%)	<0.001
Grade 1–2	44	(21.1%)	16	(7.7%)	<0.001
Grade ≥3	18	(8.6%)	6	(2.9%)	0.012
Anemia of any grade at M3	52	(24.9%)	15	(7.2%)	<0.001
Grade 1–2	51	(24.4%)	13	(6.2%)	<0.001
Grade ≥3	1	(0.5%)	2	(1.0%)	0.562
Thrombocytopenia of any grade at M3	58	(27.8%)	20	(9.6%)	<0.001
Grade 1–2	40	(19.1%)	13	(7.7%)	<0.001
Grade ≥3	18	(8.6%)	4	(1.9%)	0.002

Toxicities were graded according to CTCAE version 5.0 for cytopenias and according to the consensus grading from the ASTCT for CRS and ICANS. Only patients who experienced at least grade ≥1 toxicity are reported in the table. M1, 1 month; M3, 3 months.

frequent stage IV disease, older age and poorer performance status), ORR, CRR, PFS and OS were all significantly higher or longer after axi-cel infusion than after tisa-cel infusion. All sensitivity analyses, by considering time from CAR T order instead of time from infusion, by performing complete case analysis, by adjusting for residual imbalance in aaIPI or by using multiple imputation, led to the same exact conclusions.

Regarding toxicity, axi-cel was associated with significantly more frequent low-grade CRS and, more importantly, with significantly

more frequent grade ≥3 ICANS. The rate of grade ≥3 ICANS reported here is low, with 13.9% and 2.9% for axi-cel and tisa-cel, respectively, in the matched population. In ZUMA-1, grade ≥3 ICANS was 31% for axi-cel and 12% for tisa-cel in the JULIET trial. In RWE for patients treated with axi-cel, Nastoupil et al.¹² reported on 31% of grade ≥3 ICANS, whereas Jacobson et al.¹¹ reported on 35%. Underreporting of severe neurotoxicity in the DESCAR-T registry cannot be excluded. However, it is well-known that new mitigation strategies for CRS and ICANS management have led to much lower rates of severe CRS or ICANS. For instance, recent data on prospective evaluation of early use of dexamethasone after axi-cel infusion demonstrated 17% of grade ≥3 ICANS²⁶. In the study from the German group, grade ≥3 ICANS was 16% for axi-cel, quite similar to our data, and 7% for tisa-cel, slightly higher than what is reported here²⁷. Moreover, marked and prolonged hematological toxicity was frequently observed after axi-cel infusion compared to after tisa-cel infusion. However, no significant difference was observed with regard to grade 5 adverse events. Therefore, even if higher efficacy with axi-cel comes at the cost of higher toxicity, the latter does not undermine the significantly better outcome. Because toxicity might be of greater concern in elderly patients and could counterbalance axi-cel's higher efficacy, we undertook a planned subgroup analysis in patients aged 70 years and younger and those older than 70 years. Higher efficacy of axi-cel was still observed across age categories both for PFS and OS.

Interestingly, no significant difference was observed in DOR after PSM, whereas PFS was significantly longer with axi-cel. In fact, much of the PFS difference was related to the proportion of patients reaching a response after axi-cel as opposed to patients treated with tisa-cel and especially a complete response (60% versus 42% in the matched population). A 4-1BB-based autologous anti-CD19 CAR T product like tisa-cel is known to lead to longer persistence of the CAR T in vivo, but a CD28 co-stimulatory domain has been shown to lead to higher and faster proliferation^{28,29}. Our findings provide strong clinical support to how these bio-cellular characteristics might translate into different disease controls. Recent data have suggested that a potential dose–response relationship exists between tumor burden before infusion and subsequent disease control with tisa-cel, suggesting that tisa-cel might be more potent in case of a lower tumor burden in DLBCL³⁰. However, in a subgroup analysis, no difference in efficacy was further observed between tisa-cel and axi-cel in patients with or without a bulky disease at lymphodepletion assessed by a longest diameter of the largest node or mass >5 cm. Further correlations using total metabolic tumor volume (TMTV) or total lesion glycolysis (TLG), not readily available in DESCAR-T, will be of highest interest because it allows for a more accurate tumor bulk assessment.

Our study has limitations. First, most patient data were retrospectively collected. However, DESCAR-T is a monitored registry with high quality control. Second, a substantial amount of data were missing for PSM, and, if missing for important parameters, this may have allowed the introduction of significant residual uncontrolled bias. However, sensitivity analyses using a complete case analysis (instead of a missing indicator method), or using a multiple imputation approach, led to similar conclusions. Third, at only 11.7 months in the matched cohort, the median follow-up was short but was sufficient to reveal an OS difference between the two CAR T products. Recently reported survival curves with long follow-up of pivotal trials indicate that most deaths occur before 1 year after CAR T infusion, explaining why this short follow-up is sufficient to demonstrate significant statistical survival differences. Fourth, precise evaluation of HGBCL exhibiting double-hit or triple-hit chromosomal rearrangement by fluorescence in situ hybridization was not possible using histological data available in the registry and would require further queries or biomolecular testing. Nonetheless, almost all known confounding factors for efficacy after therapy with CAR T

were taken into account in the PSM and ITPW approaches to ensure robust and balanced comparison between the two groups of patients treated with axi-cel or tisa-cel. Although the potential influence of unmeasured confounders may undermine the validity of causal conclusions, the magnitude of the observed outcome difference makes it unlikely as demonstrated by the high E-values above 2 found for both PFS and OS. This means that a relatively strong unmeasured confounding association (for instance, as strong as a poor performance status for which HR is 2.09 for PFS) would be needed to completely explain away the poorer outcome associated with tisa-cel.

At the end of last year, the ZUMA-7 randomized phase 3 trial comparing a standard of care (SOC) strategy (salvage regimen followed by ASCT) with axi-cel in second-line DLBCL demonstrated a significantly prolonged event-free survival (EFS) associated with axi-cel³¹. Conversely, the BELINDA trial, comparing tisa-cel to SOC in second-line DLBCL as well found no difference in EFS between the two randomized strategies³⁰. Many design differences impaired a straight comparison between the two trials and their opposite conclusions. First, no bridging therapy was allowed before lymphodepletion in the ZUMA-7 trial except steroid use, as opposed to the BELINDA trial where bridging with chemotherapy was permitted. Second, early salvage regimen switching in the BELINDA trial was not considered an EFS event compared to the ZUMA-7 trial. Our RWE data suggest that, beyond extensive trial dissimilarities, a true efficacy difference between axi-cel and tisa-cel also probably substantiates the outcome divergence.

Furthermore, two RWE studies, partly addressing the same question using adjustment instead of matching, were recently reported^{27,32}. In the first one, 356 patients treated with CAR T in Germany were considered (173 treated with axi-cel and 183 treated with tisa-cel). After adjusting for six parameters in a Cox model, PFS was significantly longer after axi-cel infusion than after tisa-cel infusion. No significant difference was observed for OS. In the second one, 68 patients from the United States treated with axi-cel were compared to 31 patients treated with tisa-cel, showing higher response rate after axi-cel infusion. With 809 patients analyzed, a comprehensive matching approach encompassing most of known confounding factors, multiple sensitivity analyses and a sufficient follow-up showing, to our knowledge for the first time, an OS advantage associated with axi-cel compared to tisa-cel, our study is one of the most mature to date.

In conclusion, although only a randomized study could allow for an undisputable comparison between the two CAR T products, our study is in favor of a higher efficacy but also a higher toxicity of axi-cel compared to tisa-cel in ≥ 3 rd line of treatment for R/R DLBCL. These results need to be confirmed by other large RWE studies with similar statistical methods to account for imbalance between patient characteristics. Our findings could help in refining the choice of CAR T product for a specific patient based on the tradeoff between safety and efficacy.

Online content

Any methods, additional references, Nature Research reporting summaries, source data, extended data, supplementary information, acknowledgements, peer review information; details of author contributions and competing interests; and statements of data and code availability are available at <https://doi.org/10.1038/s41591-022-01969-y>.

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References

1. World Health Organization Classification of Tumors of Haematopoietic and Lymphoid Tissues (eds Swerdlow, S. et al) (IARC Publications, 2008).
2. Wang, M. et al. KTE-X19 CAR T-cell therapy in relapsed or refractory mantle-cell lymphoma. *N. Engl. J. Med.* **382**, 1331–1342 (2020).

3. Fowler, N. H. et al. Tisagenlecleucel in adult relapsed or refractory follicular lymphoma: the phase 2 ELARA trial. *Nat. Med.* **28**, 325–332 (2022).
4. Jacobson, C. A. et al. Axicabtagene ciloleucel in relapsed or refractory indolent non-Hodgkin lymphoma (ZUMA-5): a single-arm, multicentre, phase 2 trial. *Lancet Oncol.* **23**, 91–103 (2022).
5. Neelapu, S. S. et al. Axicabtagene ciloleucel CAR T-cell therapy in refractory large B-cell lymphoma. *N. Engl. J. Med.* **377**, 2531–2544 (2017).
6. Schuster, S. J. et al. Tisagenlecleucel in adult relapsed or refractory diffuse large B-cell lymphoma. *N. Engl. J. Med.* **380**, 45–56 (2019).
7. Abramson, J. S. et al. Lisocabtagene maraleucel for patients with relapsed or refractory large B-cell lymphomas (TRANSCEND NHL 001): a multicentre seamless design study. *Lancet* **396**, 839–852 (2020).
8. Locke, F. L. et al. Long-term safety and activity of axicabtagene ciloleucel in refractory large B-cell lymphoma (ZUMA-1): a single-arm, multicentre, phase 1–2 trial. *Lancet Oncol.* **20**, 31–42 (2019).
9. Schuster, S. J. et al. Long-term clinical outcomes of tisagenlecleucel in patients with relapsed or refractory aggressive B-cell lymphomas (JULIET): a multicentre, open-label, single-arm, phase 2 study. *Lancet Oncol.* **22**, 1403–1415 (2021).
10. Jacobson, C. et al. Long-term (≥ 4 year and ≥ 5 year) overall survival (OS) by 12- and 24-month event-free survival (EFS): an updated analysis of ZUMA-1, the pivotal study of axicabtagene ciloleucel (axi-cel) in patients (pts) with refractory large B-cell lymphoma (LBCL). *Blood* **138**, 1764–1764 (2021).
11. Jacobson, C. A. et al. Axicabtagene ciloleucel in the non-trial setting: outcomes and correlates of response, resistance, and toxicity. *J. Clin. Oncol.* **38**, 3095–3106 (2020).
12. Nastoupil, L. J. et al. Standard-of-care axicabtagene ciloleucel for relapsed or refractory large B-cell lymphoma: results from the US Lymphoma CAR T Consortium. *J. Clin. Oncol.* **38**, 3119–3128 (2020).
13. Pasquini, M. C. et al. Real-world evidence of tisagenlecleucel for pediatric acute lymphoblastic leukemia and non-Hodgkin lymphoma. *Blood Adv.* **4**, 5414–5424 (2020).
14. Sesques, P. et al. Commercial anti-CD19 CAR T cell therapy for patients with relapsed/refractory aggressive B cell lymphoma in a European center. *Am. J. Hematol.* **95**, 1324–1333 (2020).
15. Vercellino, L. et al. Predictive factors of early progression after CAR T-cell therapy in relapsed/refractory diffuse large B-cell lymphoma. *Blood Adv.* **4**, 5607–5615 (2020).
16. Neelapu, S. S. et al. Chimeric antigen receptor T-cell therapy—assessment and management of toxicities. *Nat. Rev. Clin. Oncol.* **15**, 47–62 (2018).
17. Oluwole, O. O. et al. Prophylactic corticosteroid use in patients receiving axicabtagene ciloleucel for large B-cell lymphoma. *Br. J. Haematol.* **194**, 690–700 (2021).
18. Locke, F. L. et al. Tumor burden, inflammation, and product attributes determine outcomes of axicabtagene ciloleucel in large B-cell lymphoma. *Blood Adv.* **4**, 4898–4911 (2020).
19. Pinnix, C. C. et al. Bridging therapy prior to axicabtagene ciloleucel for relapsed/refractory large B-cell lymphoma. *Blood Adv.* **4**, 2871–2883 (2020).
20. Oluwole, O. O. et al. Comparing efficacy, safety, and preinfusion period of axicabtagene ciloleucel versus tisagenlecleucel in relapsed/refractory large B cell lymphoma. *Biol. Blood Marrow Transplant.* **26**, 1581–1588 (2020).
21. Maloney, D. G. et al. Matching-adjusted indirect treatment comparison of liso-cel versus axi-cel in relapsed or refractory large B cell lymphoma. *J. Hematol. Oncol.* **14**, 140 (2021).
22. Zhang, J. et al. Letter to the editor regarding ‘Comparing efficacy, safety, and preinfusion period of axicabtagene ciloleucel versus tisagenlecleucel in relapsed/refractory large B cell lymphoma’. *Biol. Blood Marrow Transplant.* **26**, e333–e334 (2020).
23. Zhang, J. et al. A review of two regulatory approved anti-CD19 CAR T-cell therapies in diffuse large B-cell lymphoma: why are indirect treatment comparisons not feasible? *Adv. Ther.* **37**, 3040–3058 (2020).
24. Signorovitch, J. E. et al. Matching-adjusted indirect comparisons: a new tool for timely comparative effectiveness research. *Value Health* **15**, 940–947 (2012).
25. Allan, V. et al. Propensity score matching and inverse probability of treatment weighting to address confounding by indication in comparative effectiveness research of oral anticoagulants. *J. Comp. Eff. Res.* **9**, 603–614 (2020).
26. Topp, M. S. et al. Earlier corticosteroid use for adverse event management in patients receiving axicabtagene ciloleucel for large B-cell lymphoma. *Br. J. Haematol.* **195**, 388–398 (2021).
27. Bethge, W. A. et al. GLA/DRST real-world outcome analysis of CAR-T cell therapies for large B-cell lymphoma in Germany. *Blood.* **140**, 349–358 (2022).
28. Cappell, K. M. & Kochenderfer, J. N. A comparison of chimeric antigen receptors containing CD28 versus 4-1BB costimulatory domains. *Nat. Rev. Clin. Oncol.* **18**, 715–727 (2021).
29. Kawalekar, O. U. et al. Distinct signaling of coreceptors regulates specific metabolism pathways and impacts memory development in CAR T cells. *Immunity* **44**, 380–390 (2016).
30. Bishop, M. R. et al. Second-line tisagenlecleucel or standard care in aggressive B-cell lymphoma. *N. Engl. J. Med.* **386**, 629–639 (2022).

31. Locke, F. L. et al. Axicabtagene ciloleucel as second-line therapy for large B-cell lymphoma. *N. Engl. J. Med.* **386**, 640–654 (2022).
32. Gauthier, J. et al. Impact of CD19 CAR T-cell product type on outcomes in relapsed or refractory aggressive B-NHL. *Blood.* **139**, 3722–3731 (2022).

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Methods

Study design and patients. All patients treated in France with axi-cel or tisa-cel from December 2019 to October 2021 and retrospectively included in the DESCAR-T registry sponsored by the LYSARC were considered. Data export from the registry was set on 18 October 2021. All patients with DLBCL for whom a CAR T therapy with tisa-cel or axi-cel was ordered in the setting of the European Medicines Agency approval label (that is, after at least two prior lines of treatment) were considered. Patients could be treated (1) under French Temporary Authorization for Use (ATU); (2) under post-ATU authorization; or (3) under Market Authorization covered by the French health insurance system in an approved center. All patients received a non-opposition notice letter before enrollment, according to French laws. The protocol was approved by national ethics committees and the data protection agency, and the study was undertaken in accordance with the Declaration of Helsinki. DESCAR-T is registered under the ClinicalTrials.gov identifier [NCT04328298](https://clinicaltrials.gov/ct2/show/study/NCT04328298).

Outcomes. Primary outcome was PFS according to local investigator. Secondary outcomes were OS, best ORR and CRR, DOR and safety. Response was assessed according to the Lugano 2014 criteria, based on ¹⁸fluoro-deoxyglucose positron emission tomography (FDG-PET) at the approximate following timepoints: 1 month, 3 months, 6 months and 12 months after CAR T infusion³³. Best response rate was considered. For all survival endpoints, survival was calculated from the date of CAR T infusion unless otherwise specified (that is, survival from CAR T order). PFS was defined from the date of CAR T infusion to the date of first documented relapse, progressive disease, date of last follow-up or death from any cause, whichever came first. OS was defined from the date of CAR T infusion or CAR T order to the date of death from any cause or the date of last follow-up. DOR was defined from the date of first response (partial or complete) to the date of first documented relapse, date of last follow-up or death from any cause, whichever came first. Hematological toxicity was graded according to the National Cancer Institute Common Terminology Criteria for Adverse Events (CTCAE, version 5.0). Hematological toxicity was reported in patients without initiation of a new treatment for progression or relapse after CAR T infusion. CRS and ICANS were graded according to the consensus criteria from the American Society for Transplantation and Cellular Therapy (ASTCT)³⁴.

Matching procedures. PSM was used to create a balanced covariate distribution between a cohort of patients treated with axi-cel and a cohort of patients treated with tisa-cel. Propensity scores were estimated using a multivariate logistic regression model with CAR T type (axi-cel versus tisa-cel) as the dependent variable. An exhaustive list of covariates was used for PSM: age (as a continuous parameter), sex, lactate dehydrogenase (LDH) level (normal versus between the upper limit of normal (ULN) and 2× ULN versus >2× ULN), C reactive protein (CRP) (dichotomized with a cutoff set at 30 mg L⁻¹), time between last treatment and infusion (continuous), Eastern Cooperative Oncology Group (ECOG) performance status (PS) (0–1 or ≥2), Ann Arbor stage (I versus II versus III versus IV), number of prior lines of treatment before CAR T (2–4 versus >4), bridging and response to bridging (no bridge versus bridging and response (partial or complete) to bridging versus bridging and no response (stable or progressive disease)), prior SCT either autologous or allogeneic (yes versus no), bulk assessed at lymphodepletion (dichotomized with a cutoff set at 5 cm), center (all centers with fewer than 20 patients were grouped into one category) and diagnosis (DLBCL NOS or HGBCL versus transformed indolent lymphoma (tFL or tMZL)). To account for a given center experience in CAR T procedure implementation and improvement of toxicity management over time that might impact outcome (especially because some centers had access to one CAR T before the other), time between first CAR T order for that center and CAR T infusion for a given patient was also considered for PSM (as a continuous parameter). For all matching parameters except continuous variables (no missing value could be used for continuous parameters in PSM), missing data were considered as one distinct category for PSM. Of note, when survival was assessed from CAR T order instead of CAR T infusion, time intervals were calculated until or from CAR T order instead of CAR T infusion. Matching parameters are detailed in Extended Data Table 3.

Matching was performed considering a 1:1 ratio without replacement and with optimal matching applying a caliper width of the propensity score set at 0.1. Basically, a patient treated with tisa-cel was selected and then matched with a patient treated with axi-cel given the constraint that the difference between the logit (that is, the logarithm of the odds of the logistic regression that models the probability of receiving tisa-cel or axi-cel) was less than a pre-specified maximum (that is, the caliper distance).

IPTW was used as another statistical approach to allow for outcome comparison between patients treated with axi-cel and patients treated with tisa-cel. In the IPTW method, the weight for each patient is calculated by inverting the probability of receiving the treatment the patient actually receives. PSM and IPTW rely on different statistical matching approaches, provide different information and should be interpreted differently. The first one (PSM) allows for assessing average treatment effect for the treated (ATT), whereas the other (IPTW with the weighting technique used here) provides estimation of the average treatment

effect (ATE). The first gives the average effect of treatment on those patients who ultimately received one CAR T versus the other, whereas the second provides the average effect of theoretically moving the entire population from receiving one CAR T to the other. For IPTW, the exact same covariates as for PSM were used for the logistic regression model to calculate the propensity of receiving one of the CAR T products versus the other. Methodology underlying propensity-score-based matched comparisons and differences with adjustment approaches have been reviewed elsewhere³⁵.

Sensitivity analyses. Several sensitivity analyses were conducted. First, all patients with at least one missing value for at least one matching variable were removed from PSM analysis (complete case analysis). Second, a multiple imputation approach was performed using the fully conditional specification (FCS) method, allowing for different distributions across variables. Continuous variables were imputed using linear regression, whereas categorical parameters were imputed using logistic regression. All propensity score covariates and outcome (OS) were used for imputation. Ten imputed datasets were generated. A treatment effect was estimated within each imputed dataset using PSM. Estimated treatment effects from each imputed dataset were then combined into a single treatment effect using Rubin's rule (within method). Third, a Cox bivariate model adjusting for residual aalPI imbalance after matching was used to assess association between CAR T product and outcome (PFS and OS). Fourth, PSM was performed with a time of origin for OS set at the time of CAR T order instead of the time of CAR T infusion. Finally, to assess how robust the association between CAR T product and outcome was to potential unmeasured or uncontrolled confounding, E-value was computed³⁶. It represents the minimum strength of association that a unique (or a set of) unmeasured confounder would need to have with both the treatment and the outcome conditional on the measured covariates to fully explain away the association between treatment (here, the CAR T product) and the outcome (here, PFS or OS). Therefore, the higher the E-value, the stronger the confounder associations must be to explain away the effect.

Statistical analysis. Survival distributions were compared using the log-rank test. Response rates were compared using the χ^2 test. A two-sided *P* value of less than 0.05 was considered significant. No adjustment was performed for multiple testing. Two subgroup analyses according to age (≤ 70 years and > 70 years) and tumor bulk (≤ 5 cm and > 5 cm) were pre-planned in the statistical analysis plan. Survival curves were generated using the Kaplan–Meier estimation method. Statistical analyses were performed using SAS software version 9.3 and R version 4.2.0. The MATCH macro for PSM and the MI and MIANALYZE procedures for multiple imputation were used with SAS. The E-value package was used with R.

Reporting summary. Further information on research design is available in the Nature Research Reporting Summary linked to this article.

Data availability

Data from the DESCAR-T registry are subject to controlled access by the LYSARC owing to privacy and legal requirements and to proprietary reasons. Anonymized IPD requests will be promptly reviewed by the corresponding author (E.B.) and the scientific committee of the DESCAR-T registry. Individual de-identified participant data will be made available for replication and validation purposes from the present study only. For any other reason, an agreement for data sharing will depend on the nature of the request, the intended use of the data and their availability, as well as the merit of the research project. Agreement will be made after the DESCAR-T scientific committee decision, and a data sharing agreement will have to be signed before any data transfer. All requests should be addressed to descar-t@lysarc.org. A reply will be provided within 1 month after the data request.

References

- Cheson, B. D. et al. Recommendations for initial evaluation, staging, and response assessment of Hodgkin and non-Hodgkin lymphoma: the Lugano classification. *J. Clin. Oncol.* **32**, 3059–3068 (2014).
- Lee, D. W. et al. ASTCT consensus grading for cytokine release syndrome and neurologic toxicity associated with immune effector cells. *Biol. Blood Marrow Transplant.* **25**, 625–638 (2019).
- Austin, P. C. The use of propensity score methods with survival or time-to-event outcomes: reporting measures of effect similar to those used in randomized experiments. *Stat. Med.* **33**, 1242–1258 (2014).
- VanderWeele, T. J. & Ding, P. Sensitivity analysis in observational research: introducing the E-value. *Ann. Intern. Med.* **167**, 268–274 (2017).

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Author contributions

E.B., S.L.G., R.H., F.M., E.G. and F.B. contributed to study design. E.B., S.L.G., R.D.B., P.S., G.M., G.C., D.B., L.R., F.X.G., M.T.R., P.B., J.O.B., C.C.L., S.C., R.O.C., M.M., S.G., M.J., M.L., S.C., J.A., A.C., L.D.L.R., B.D.F., O.H., T.G., J.J.T., C.T., R.H. and F.M. enrolled and treated patients. E.G. performed statistical analyses. F.B. directed the registry database organization for the LYSARC. All authors analyzed and interpreted the data. All authors contributed to the writing of the manuscript and approved its final version.

Competing interests

The authors report the following competing interests: E.B.: consulting fees or honoraria from Novartis, Kite/Gilead, Roche, Takeda and Incyte; research funding (paid to institution) from Amgen; and travel and personal fees from Roche and Incyte. S.L.G.: honoraria from Janssen-Cilag, Kite/Gilead and Novartis. P.S.: honoraria or consultancy from Chugai, Bristol Myers Squibb, Novartis and Kite/Gilead. T.G.: honoraria from Kite/Gilead, Pfizer and Takeda. S.G.: honoraria from Kite/Gilead, Incyte, Takeda and Janssen. P.B.: honoraria from Bristol Myers Squibb, Kite/Gilead, Novartis and Abbvie. R.H.: honoraria from Bristol Myers Squibb, Kite/Gilead, Incyte, Janssen, Merck Sharp & Dohme, Takeda, Novartis and Roche. F.M.: consulting fees or honoraria from Genmab, Novartis, Kite/Gilead, Bristol Myers Squibb, AstraZeneca, Epizyme, Roche, Abbvie, Chugai, Janssen, Incyte, Kymera, Miltenyi and Roche; and expert testimony for Roche. O.H.: consultancy for AB Science and Inaterys; and research funding (paid to institution) from Bristol Myers Squibb and Alexion. G.C.: consulting fees and honoraria from Roche, Bristol Myers Squibb, Onwards Therapeutics, MedxCell, EmerCell, MabQ, Sanofi, Abbvie, Takeda, Roche, Janssen, Roche, Novartis and Myltenyi. M.L.:

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Additional information

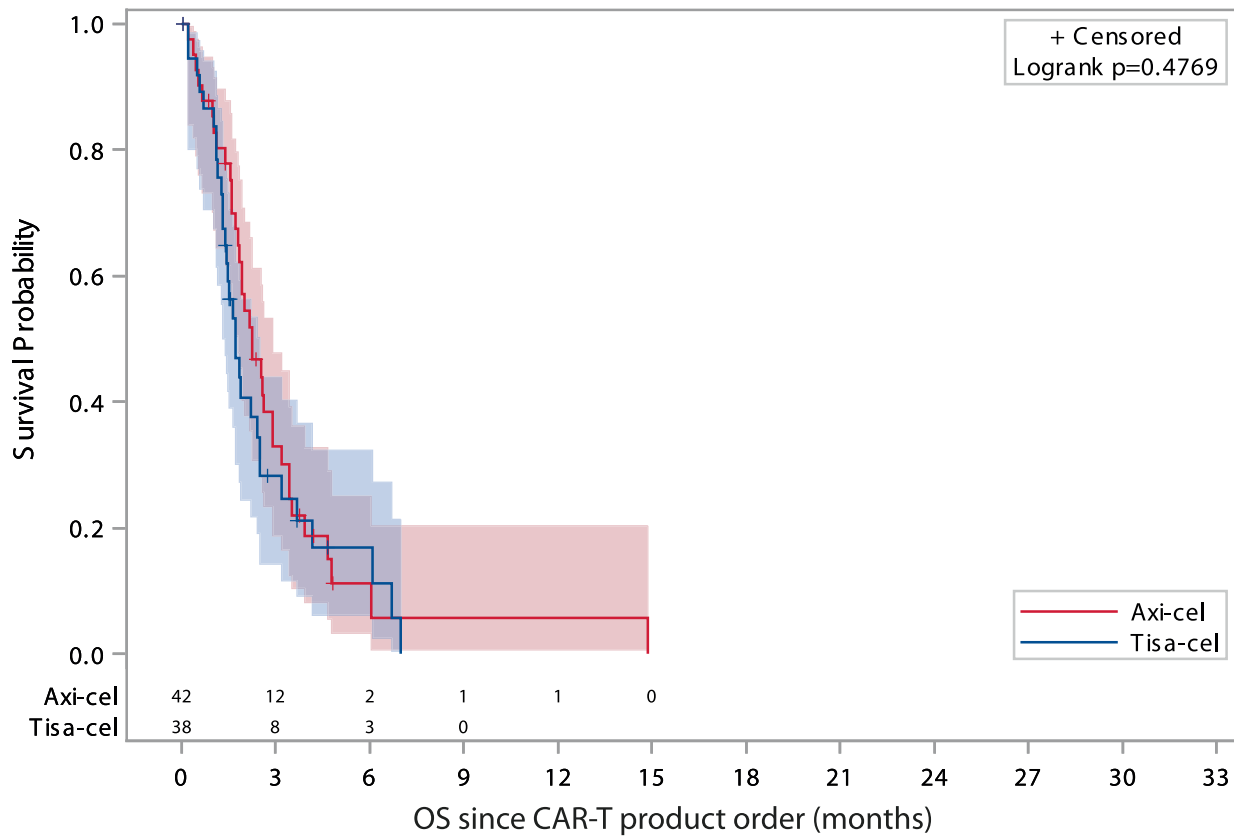
Extended data is available for this paper at <https://doi.org/10.1038/s41591-022-01969-y>.

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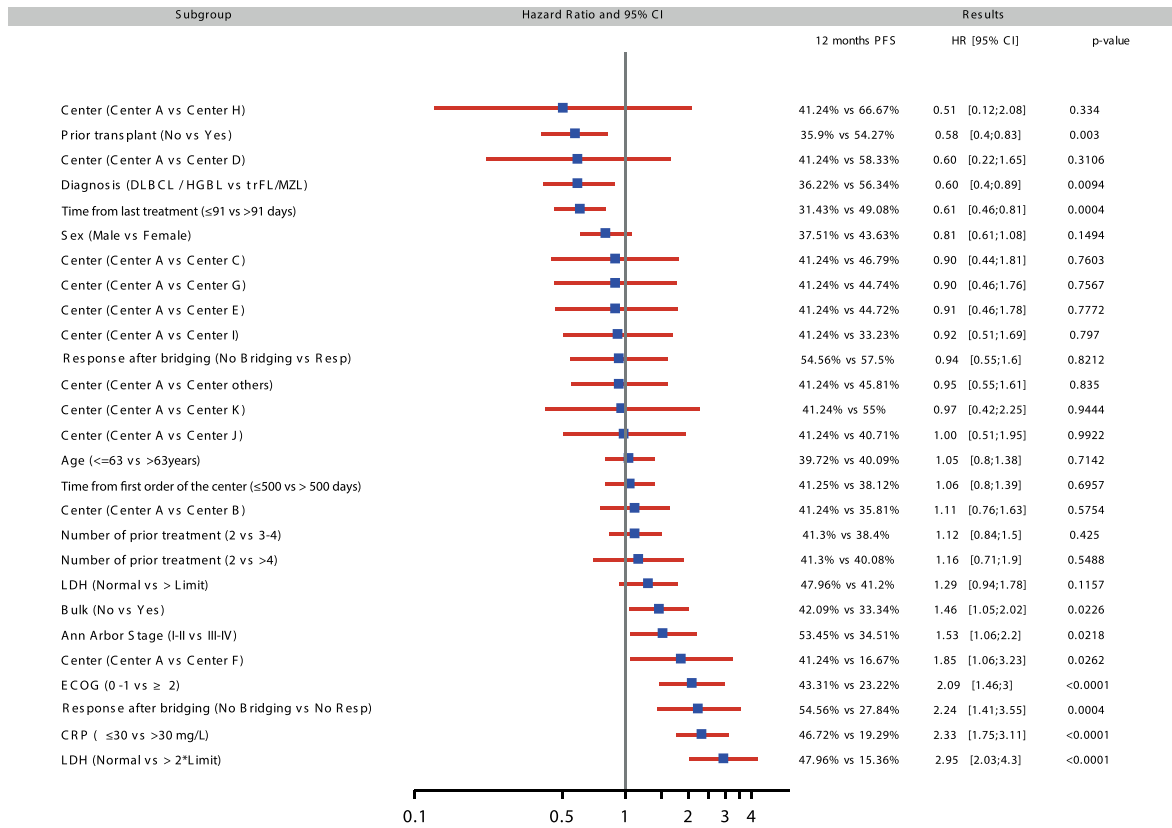
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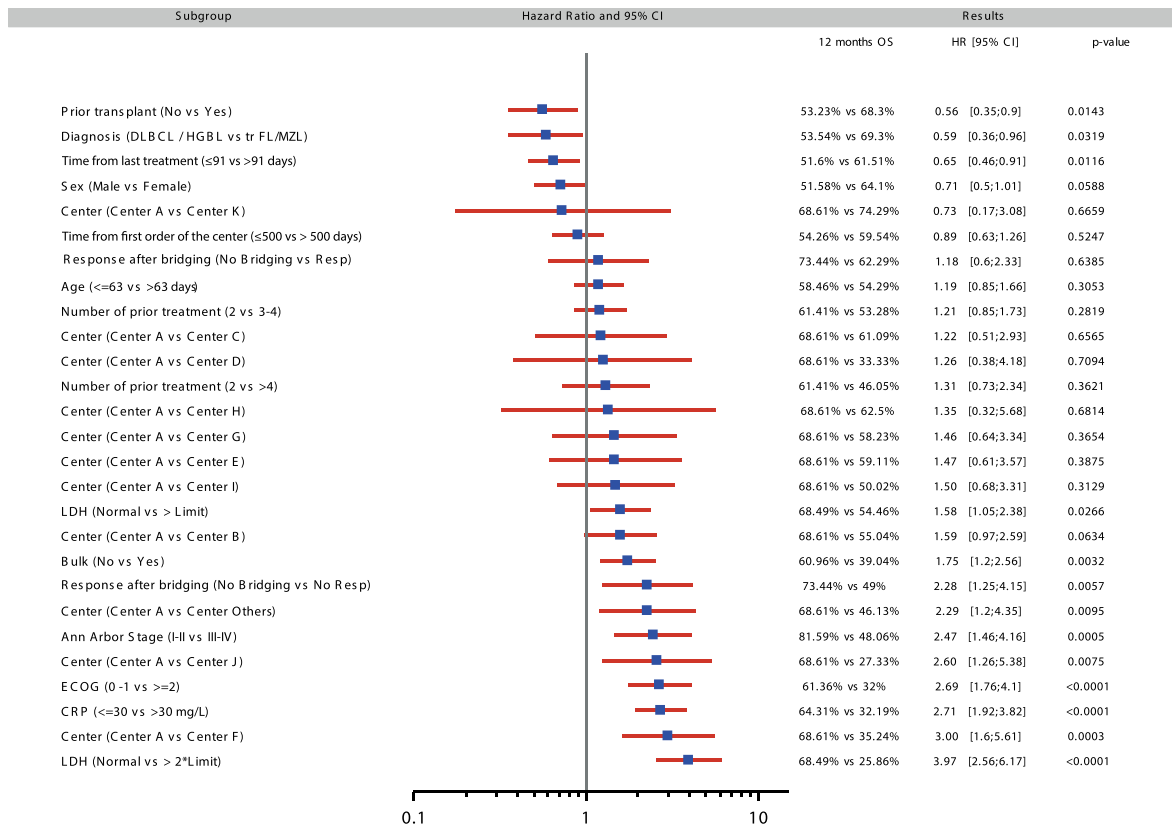


Extended Data Fig. 1 | Overall survival (OS) for patients with a CAR-T product order who did not proceed until infusion according to CAR-T type. Extended Data Fig. 1 For 80 patients, a CAR-T product was ordered but was never infused ($n=42$ for axi-cel, $n=38$ for tisa-cel) due to disease progression ($n=60$), physician decision ($n=6$) and other reasons detailed in the patient flow diagram in Fig. 1 ($n=14$). Shaded areas correspond to the 95% confidence bands using Hall-Wellner method. P value was calculated using a two-sided logrank test.

a

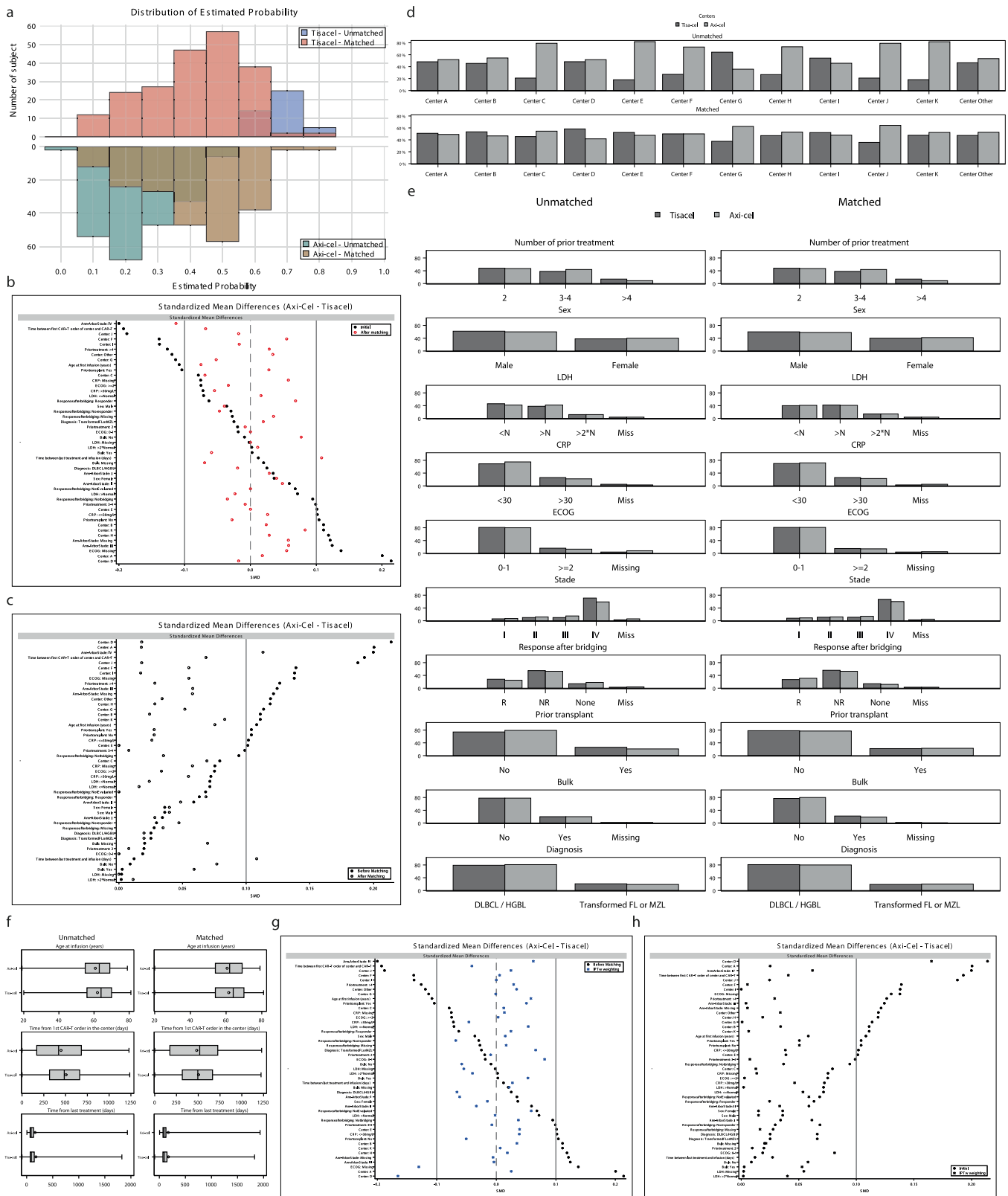


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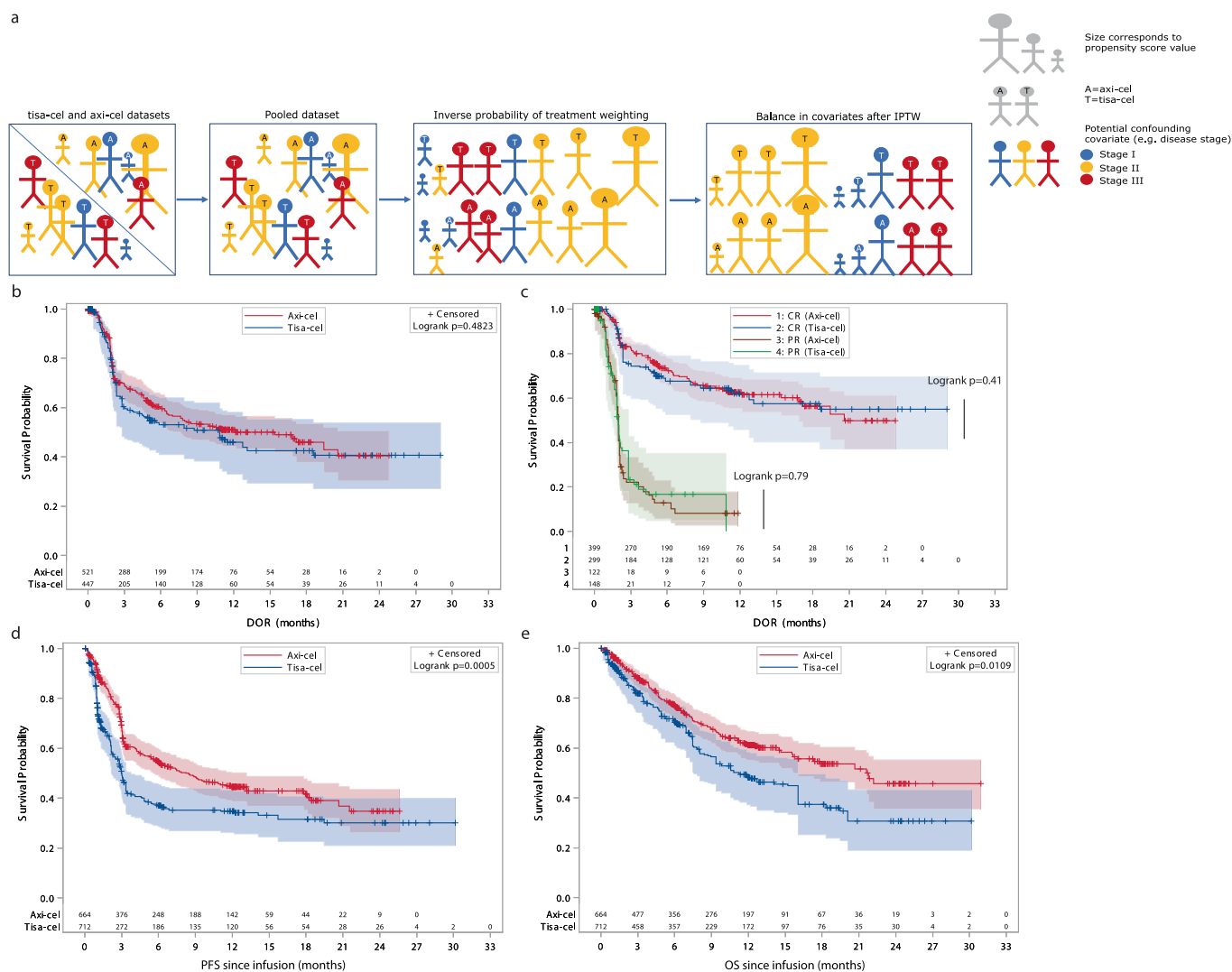


Extended Data Fig. 2 | See next page for caption.

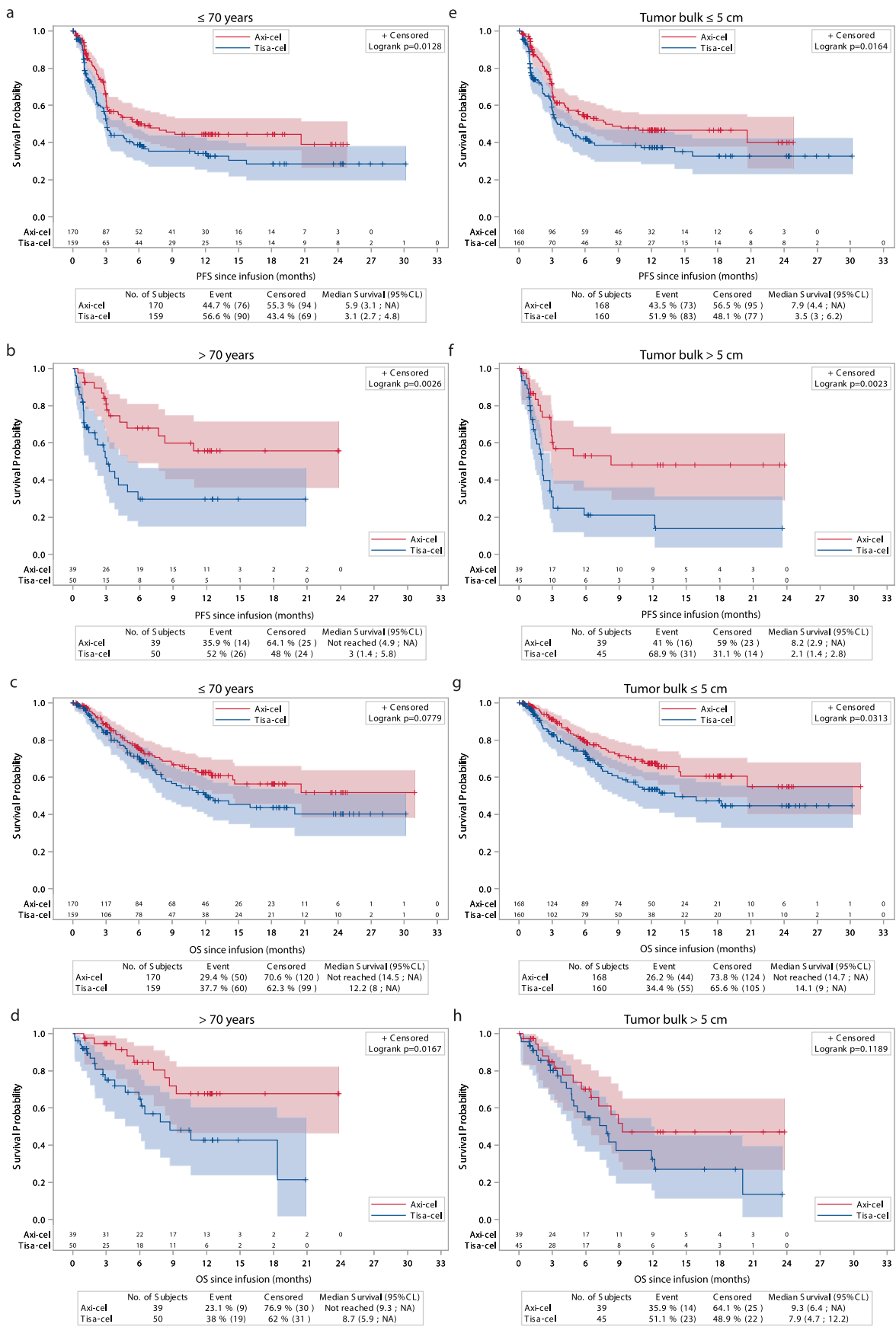
Extended Data Fig. 2 | Univariate prognostic analysis. **a**, univariate analysis for progression-free survival (PFS). **b**, univariate analysis for overall survival (OS). Blue point represents the value of the hazard ratio (HR) and red segment the value of the 95% confidence interval (CI). The first category in parentheses is taken as the reference category for comparison and HR computation. For instance, for PFS analysis HR is 2.95 for patients with a LDH level twice above the upper limit of the normal (ULN) compared to patients with a normal value. A $HR < 1$ represents a prognosis factor associated with a prolonged survival while a $HR > 1$ represents a prognosis factor associated with a shorter survival. A prognostic factor is statistically significant if the 95% CI does not contain 1. Cox univariate model was used for calculating HR and associated two-sided *P* value. No adjustment was performed for multiple comparisons. All centers were anonymized. Time from last treatment, age and time to first order of the center were dichotomized according to the median value of data distribution. Number of prior treatment, LDH level and bridging were divided into 3 categories (2 vs 3-4 vs >4 prior lines; normal LDH, LDH between 1 and 2 times the ULN and LDH above 2 times the ULN; no bridging vs response to bridging vs no response to bridging, respectively). *Time from last treatment* represents the time from the start of the last treatment to the time of CAR T infusion. *Time from first order of the center* represents the time from the order of the first CAR-T in the center to the time of infusion of the CAR T for the patient (as a surrogate of the “center experience” for CAR-T therapy for a given patient). DLBCL, diffuse large B-cell lymphoma; HGBL, high grade B-cell lymphoma; trFL/MZL, transformed follicular lymphoma or marginal zone lymphoma; Resp; complete or partial response to bridging; No Resp, no response (that is stable or progressive disease) to bridging; ECOG, Eastern Collaborative Oncology Group Performance Status; CRP, C reactive protein; LDH, lactate dehydrogenase.



Extended Data Fig. 3 | Balance assessment before and after matching. **a**, Propensity score (PS) distribution before and after PSM. **b**, Standardized Mean Differences (SMD) of covariates categories before and after PSM. **c**, Absolute SMD before and after PSM. **d**, CAR T products distribution across centers from the DESCAR-T registry before and after PSM (light grey= axi-cel; dark grey=tisa-cel). **e**, CAR T products distribution according to categorical covariates before and after PSM (light grey= axi-cel; dark grey=tisa-cel). **f**, Balance assessment according to CAR T product for continuous covariates before and after PSM. Box plot represents 1st quartile and 3rd quartile. Line in the middle of the box represents the median. Round symbol represents the mean. Left segment represents distribution from the minimal value. Right segment represents distribution to the maximal value. **g**, SMD of covariates categories before and after IPTW. **h**, absolute SMD before and after IPTW.

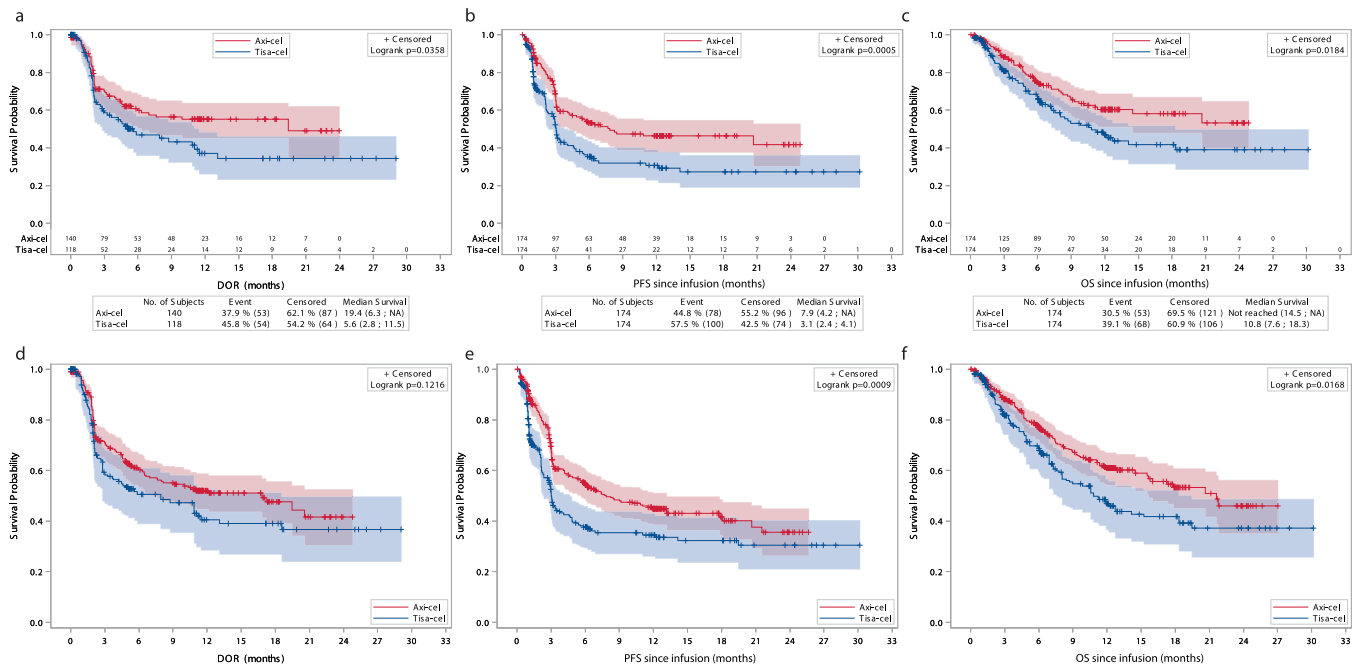


Extended Data Fig. 4 | Response rates and survival according to CAR-T type after inverse probability of treatment weighting. **a**, In IPTW, weight of each individual patient is calculated as the inverse of their probability of receiving tisa-cel or axi-cel, assessed by their propensity score. Compared to PSM, it creates a pseudo-population of patients in which patients with a lower likelihood of receiving one CAR-T is over-weighted in the final population. As for PSM, comparability according to each covariate of the resulting 2 pseudo-cohorts of patients receiving one CAR-T or the other is checked using standardized mean differences (SMD, Extended Data Fig. 3). **b**, DOR according to CAR-T. **c**, DOR according to CAR-T and response quality. **d**, PFS according to CAR-T. **e**, OS according to CAR-T. Shaded areas correspond to the 95% confidence bands using Hall-Wellner method. *P* values were calculated using two-sided logrank tests. No adjustment was made for multiple comparisons.

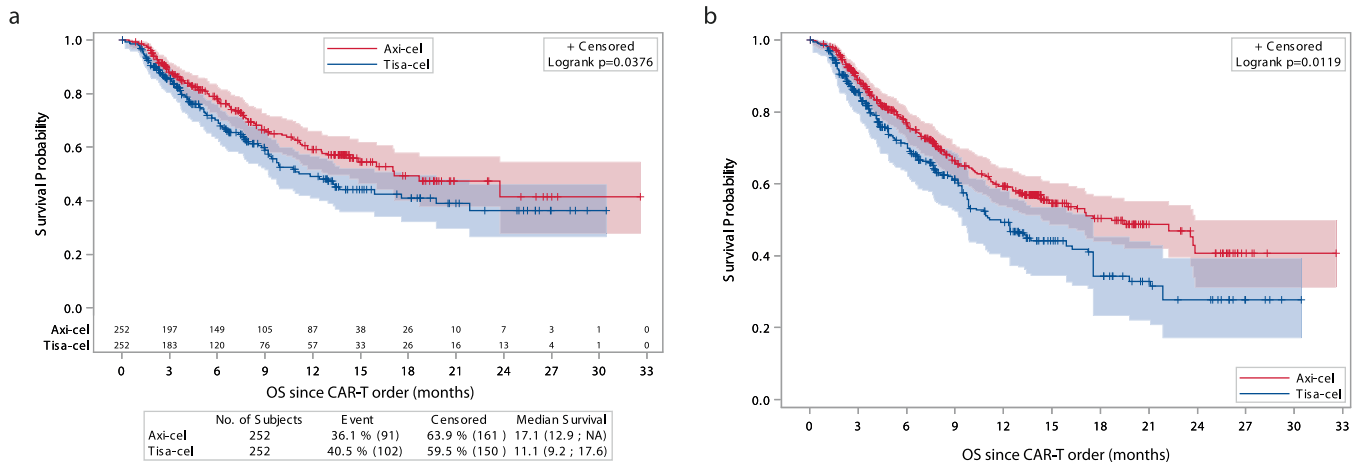


Extended Data Fig. 5 | See next page for caption.

Extended Data Fig. 5 | Planned subgroup analyses according to age and tumor bulk. a, PFS according to CAR T product in patients ≤ 70 years. **b**, PFS according to CAR T product in patients > 70 years. **c**, OS according to CAR T product in patients ≤ 70 years. **d**, OS according to CAR T product in patients > 70 years. **e**, PFS according to CAR T product in patients with ≤ 5 cm tumor bulk. **f**, PFS according to CAR T product in patients with > 5 cm tumor bulk. **g**, OS according to CAR T product in patients with ≤ 5 cm tumor bulk. **h**, OS according to CAR-T in patients with > 5 cm tumor bulk. Shaded areas correspond to the 95% confidence bands using Hall-Wellner method. P values were calculated using two-sided logrank tests. No adjustment was made for multiple comparisons.



Extended Data Fig. 6 | Survival according to CART product after PSM or IPTW in the complete case analysis (that is, where all cases with at least one missing value in matching covariates have been removed). **a**, DOR according to CART product after PSM. **b**, PFS according to CART product after PSM. **c**, OS according to CART product after PSM. **d**, DOR according to CART product after IPTW. **e**, PFS according to CART product after IPTW. **f**, OS according to CART product after IPTW. Shaded areas correspond to the 95% confidence bands using Hall-Wellner method. P values were calculated using two-sided logrank tests. No adjustment was made for multiple comparisons.



Extended Data Fig. 7 | Overall survival from CAR-T order (instead of from infusion) according to CAR-T type after PSM or IPTW. a, OS according to CAR-T after PSM. **b,** OS according to CAR-T after IPTW. Shaded areas correspond to the 95% confidence bands using Hall-Wellner method. *P* values were calculated using two-sided logrank tests. No adjustment was made for multiple comparisons.

Extended Data Table 1 | Patient characteristics in the matched cohorts without any missing data for matching covariates (that is, complete case analysis)

	Treatment		Matched set N=348
	Tisa-cel N=174	Axi-cel N=174	
Age at First Infusion (Years)			
Median	64.00	63.00	63.00
Min ; Max	25;81	20;79	20;81
Sex			
Male	104 (59.8%)	99 (56.9%)	203 (58.3%)
Female	70 (40.2%)	75 (43.1%)	145 (41.7%)
LDH at Lymphodepletion			
<= Normal	72 (41.4%)	77 (44.3%)	149 (42.8%)
> Normal	77 (44.3%)	71 (40.8%)	148 (42.5%)
> 2*Normal	25 (14.4%)	26 (14.9%)	51 (14.7%)
CRP at Lymphodepletion			
<= 30 mg/L	131 (75.3%)	128 (73.6%)	259 (74.4%)
> 30 mg/L	43 (24.7%)	46 (26.4%)	89 (25.6%)
Time between Last Treatment and Infusion (Days)			
Median	90.50	90.50	90.50
Q1 ; Q3	70;138	71;136	70;136
Performance Status (ECOG Scale) at Injection			
0-1	143 (82.2%)	149 (85.6%)	292 (83.9%)
>=2	31 (17.8%)	25 (14.4%)	56 (16.1%)
Ann Arbor Stage			
I	13 (7.5%)	19 (10.9%)	32 (9.2%)
II	17 (9.8%)	21 (12.1%)	38 (10.9%)
III	20 (11.5%)	22 (12.6%)	42 (12.1%)
IV	124 (71.3%)	112 (64.4%)	236 (67.8%)
Number of Prior Treatment Lines			
Median	3.00	3.00	3.00
Min;Max	2;10	2;7	2;10
Number of Prior Treatment Lines in Class			
2	85 (48.9%)	86 (49.4%)	171 (49.1%)
3-4	78 (44.8%)	71 (40.8%)	149 (42.8%)
>4	11 (6.3%)	17 (9.8%)	28 (8.0%)
Response After Last Bridging Therapy			
Responder	48 (27.6%)	52 (29.9%)	100 (28.7%)
Not Responder	101 (58.0%)	98 (56.3%)	199 (57.2%)
Not Evaluated	1 (0.6%)	2 (1.1%)	3 (0.9%)
No Bridging	24 (13.8%)	22 (12.6%)	46 (13.2%)
At Least One Prior Transplant			
No	139 (79.9%)	131 (75.3%)	270 (77.6%)
Yes	35 (20.1%)	43 (24.7%)	78 (22.4%)
Bulk			
No	137 (78.7%)	136 (78.2%)	273 (78.4%)
Yes	37 (21.3%)	38 (21.8%)	75 (21.6%)
Time between First CAR-T Order of Center and CAR-T Order of Patient (Days)			
Median	504.00	555.00	521.50
Q1;Q3	326;676	222;762	303;722
Diagnosis			
DLBCL / HGBL	140 (80.5%)	138 (79.3%)	278 (79.9%)
Transformed FL or MZL	34 (19.5%)	36 (20.7%)	70 (20.1%)

Extended Data Table 2 | Response rates in the matched cohorts without any missing data for matching covariates. Patients without response assessment (due to whatever reason) are considered as non-responders

	Treatment		Matched Datasets N=348	<i>P</i>
	Tisa-Cel N=174	Axi-Cel N=174		
Complete Response (CR)				
Patients with CR	73 (42.0%)	107 (61.5%)	180 (51.7%)	<0.001
IC 95% For CR Rate	[34.5% ; 49.7%]	[53.8% ; 68.8%]	[46.3% ; 57.1%]	
Overall Response (ORR)				
Patients with Overall Response	118 (67.8%)	140 (80.5%)	258 (74.1%)	0.007
IC 95% For CR Rate	[60.3% ; 74.7%]	[73.8% ; 86.1%]	[69.2% ; 78.7%]	

Extended Data Table 3 | List of covariates used for propensity score calculation

Variable	Type	List of covariates and time of parameter evaluation in the analysis since order	List of covariates and time of parameter evaluation in the analysis since infusion
Age	Continuous	At CAR T product order	At CAR T product infusion
Sex	Male vs Female	X	X
LDH	Normal vs]ULN-2*ULN] vs >2*ULN	Not available	At lymphodepletion or at infusion if value at lymphodepletion is missing
CRP	≤ 30 mg/L vs > 30 mg/L	Not available	At lymphodepletion or at infusion if value at lymphodepletion is missing
Time between last treatment and order or infusion	Continuous (days)	At order	At infusion
Performance status (ECOG scale)	0-1 vs >2	At decision of CAR T treatment	At lymphodepletion or at infusion if value at lymphodepletion is missing
Ann Arbor Stage	1 vs 2 vs 3 vs 4	At decision of CAR T treatment	At decision of CAR T treatment
Number of prior treatment lines	2-4 vs >4	X	X
Response after bridging therapy	Responder (PR, CR) vs Not responder (SD, PD) vs Not Evaluated vs No bridging	Not applicable	X
Prior transplant (autologous or allogenic)	Yes vs No	X	X
Bulk (largest diameter cut-off 5 cm)	Yes vs No	Not available	X
Center (all centers with <20 patients were grouped into a single category)	Center 1 vs center 2 vs ...	X	X
Time between 1 st CAR-T order of the center and CAR-T order for the patient (days)	Continuous	X	X
Diagnosis (at CAR-T treatment order if available, or at initial diagnosis otherwise)	DLBCL NOS or HGBL vs transformed from indolent lymphoma	X	X

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Data from the DESCAR-T registry are subject to controlled access by the LYSARC due to privacy and legal requirement and to proprietary reasons. Anonymized individual patient data (IPD) request will be promptly reviewed by the corresponding author (EB) and the scientific committee of the DESCAR-T registry. Individual de-identified participant data will be made available for replication and validation purpose of results from the present study only. For any other reason, agreement for data sharing will depend on the nature of the request, the intended use of the data and their availability, as well as the merit of the research project. Agreement will be made following the DESCAR-T scientific committee decision and a data sharing agreement will have to be signed before any data transfer. All requests should be addressed to descar-t@lysarc.org. Reply will be provided within one month following data request.

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Life sciences study design

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Sample size	No sample size calculation was required for this retrospective analysis with no statistical assumption a priori. All patients included in the French DESCAR-T registry were analyzed in the study.
Data exclusions	No data from patients with DLBCL treated in ≥ 3 rd line of treatment and part of the DESCAR-T registry were excluded (see patients flow).
Replication	Does not apply to clinical patient data included in the study. Many sensitivity analyses were performed to ensure robustness of the results with missing data indicator category, complete case analysis, analyses from CAR-T order instead of infusion, multiple imputation approaches and unmeasured confounder evaluation.
Randomization	No randomization was considered in this retrospective study. All potential measured confounders were taken into consideration using propensity score matching and inverse probability of treatment weighting. An extensive list of fourteen covariates were used for matching.
Blinding	No blinding was performed in the study since commercial CAR-T product order and infusion in routine practice was analyzed (real-world evidence data).

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Population characteristics	All patient characteristics are detailed in Table 1 of the manuscript
Recruitment	All patients treated in France with axi-cel or tisa-cel from December 2019 to October 2021 and retrospectively included in the DESCAR-T registry sponsored by LYSARC were considered. Data export from the registry was set on the 18TH of October 2021. All patients with DLBCL for whom a CAR-T therapy with tisa-cel or axi-cel was ordered in the setting of the European Medical Agency (EMA) approval label (i.e., after at least 2 prior lines of treatment) were considered.
Ethics oversight	The protocol was approved by national ethics committees and the data protection agency from France, and the study was undertaken in accordance with the Declaration of Helsinki.

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Clinical data

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All manuscripts should comply with the ICMJE [guidelines for publication of clinical research](#) and a completed [CONSORT checklist](#) must be included with all submissions.

Clinical trial registration	DESCAR-T is registered under the ClinicalTrials.gov identifier NCT04328298.
Study protocol	No specific protocol for planned ancillary study based on the retrospective DESCAR-T registry is available.
Data collection	All patients treated in France with axi-cel or tisa-cel from December 2019 to October 2021 and retrospectively included in the DESCAR-T registry sponsored by LYSARC were considered. Data export from the registry was set on the 18TH of October 2021. A full list of participating centers can be found on the ClinicalTrial.gov website: https://clinicaltrials.gov/ct2/show/NCT04328298
Outcomes	Primary outcome was PFS according to local investigator. Secondary outcomes were overall survival OS, best ORR and CRR (according to Lugano 2014 criteria), DOR and safety. All information can be retrieved from the Patients & Methods section of the manuscript.