



When a *Neisseria meningitidis* PCR limitation contributes to an immunological disease diagnosis

[Correspondence]

Claire Duployez, Caroline Loiez, Frédéric Wallet, Laure Marceau, Myriam Simon, Ala-Eddine Deghmane, Muhamed-Kheir Taha, Anne Vachée

► To cite this version:

Claire Duployez, Caroline Loiez, Frédéric Wallet, Laure Marceau, Myriam Simon, et al.. When a *Neisseria meningitidis* PCR limitation contributes to an immunological disease diagnosis [Correspondence]. *Journal of Microbiology, Immunology and Infection*, In press, 10.1016/j.jmii.2023.10.009 . hal-04270102

HAL Id: hal-04270102

<https://hal.science/hal-04270102>

Submitted on 3 Nov 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial - NoDerivatives 4.0 International License

Available online at www.sciencedirect.com

ScienceDirect

journal homepage: www.e-jmii.com

Correspondence

When a *Neisseria meningitidis* PCR limitation contributes to an immunological disease diagnosis

KEYWORDS

Capsule;
Neisseria meningitidis;
 PCR;
 16S rRNA gene
 sequencing

dismutase, a generic conserved gene in *Neisseria meningitidis*) PCR was positive while the *ctrA* (capsule transport gene) PCR was negative, suggesting a capsule null locus (*cnl*) strain.³ The strain belonged to the clonal complex CC175 (Table 1). This finding led us to look for other factors contributing to IMD such as underlying disease: only hypocomplementemia was detected (complement hemolytic 50 level <14 U/mL).

The major virulence factor leading to IMD is the polysaccharide capsule. Non-capsulated *N. meningitidis* strains can colonize nasopharynx of healthy subjects and are low-pathogenic strains, because not resistant to complement mediated lysis and opsonophagocytosis.^{3,6} Few infection cases are reported in the literature, mostly in immuno-compromised patients. In 2018, Kurose summarized 13 cases, including nine meningitis: three patients with C6 deficiency, one patient with IgG4-related disease, and five with unknown risk factor.⁶

Patients with a deficiency in the terminal complex and the factor properdin of the complement system have an increased risk of recurrent capsulated IMD. Less frequently, IMD caused by non-capsulated strains have been reported.⁷ Rosain reported 61 IMD among subjects with terminal complement pathway deficiencies: 8 % were due to non-capsulated strains.² Ladhami described 20 IMD among patients with inherited and acquired complement deficiency: four among nine IMD in patients on eculizumab therapy were due to non-groupable or group E (less virulent) isolates.¹ Thus, it is necessary to investigate immune comorbidities after non-capsulated IMD.

The capsule being a virulence factor responsible for IMD, *ctrA* is frequently used to detect *N. meningitidis*, including in commercial PCR assays. Nevertheless, it has always been described that nucleotide substitution/rearrangement in *ctrA* gene may be responsible for false-negative results, including in invasive strains.⁴ Carriage isolates may even lack this gene.⁵ Then our last taking-home message is that

Dear editor,

Capsulated *Neisseria meningitidis* is responsible for invasive meningococcal disease (IMD). Only few non-capsulated IMD are described in the literature.^{1,2}

A 36-year-old patient, without previous medical history, presented for stiffness and headache. He had fever (39 °C), photophobia, vomiting, purpura on ankles and torso. Blood leukocyte count was $29,98 \times 10^9$ cells/L (95.5 % polymorphonuclear), C-reactive protein 33.77 mg/dL, procalcitonin 18.4 ng/mL. He was given 2 g cefotaxime and 20 mg dexamethasone few hours before lumbar puncture and blood sample. Evolution was quickly favorable.

The cerebrospinal fluid revealed a leukocyte count of $20,50 \times 10^9$ cells/L (92 % neutrophils), protein 729 mg/dL, glucose <4 mg/dL, lactic acid >16 mmol/L, and gram-negative diplococci were observed on the Gram stain. Blood and cerebrospinal fluid remained sterile. FilmArray® meningitis/encephalitis panel multiplex PCR (BioMérieux, Marcy l'Étoile, France) and *N. meningitidis* real-time PCR were negative. By 16S rRNA sequencing, a 677 bp DNA bacterial fragment was obtained and was 99.5 % identity with *Neisseria* sp. strains including *N. meningitidis*. The French National Reference Laboratory detected a non-capsulated *N. meningitidis* strain with two routinely used PCR: the *sodC* (Cu-Zn superoxide

<https://doi.org/10.1016/j.jmii.2023.10.009>

1684-1182/Copyright © 2023, Taiwan Society of Microbiology. Published by Elsevier Taiwan LLC. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Table 1 Molecular biology tests performed on the cerebrospinal fluid sample.

Tests performed	Targets	Result
BioFire® FilmArray meningitis/encephalitis panel (BioMérieux, Marcy l'Étoile, France)	« Encapsulated <i>N. meningitidis</i> (groups A, B, C, W, Y) and DNA from a strain with a variant <i>ctrA</i> gene »	Not detected
<i>N. meningitidis</i> real-time PCR (Diagenode Diagnostics, Liège, Belgium)	<i>ctrA</i> (capsule transport in encapsulated strains) <i>csaB</i> (group A) <i>csb</i> (group B) <i>csc</i> (group C) <i>csw</i> (group W) <i>csy</i> (group Y)	Not detected
16S rRNA sequencing (377 ABI Prism; PE Applied Biosystems, Foster City, CA., USA)	Universal 16S RNA PCR	<i>Neisseria</i> sp.
Analysis with BLAST system		
<i>sodC</i> and <i>ctrA</i> real-time PCR (Centre National de Référence des méningocoques, Institut Pasteur, Paris, France)	<i>sodC</i> (Cu–Zn superoxide dismutase gene) <i>ctrA</i> (capsule transport in encapsulated strains)	Detected Not detected
MLST (Centre National de Référence des méningocoques, Institut Pasteur, Paris, France)	Multilocus sequence typing	Clonal complex CC175
Final result		Non-capsulated <i>N. meningitidis</i>

biological tests cannot be 100 % sensitive. This warrants a systematic *sodC* PCR in addition to *ctrA* PCR since *sodC* is conserved in *N. meningitidis* strains regardless of capsule status but not in other *Neisseria* species.⁵ Presume meningococcal disease with negative PCR may be due to a cnl strain and clinical history remains the best indicator for diagnosis.

Declaration of competing interest

None.

References

1. Ladhani SN, Campbell H, Lucidarme J, Gray S, Parikh S, Willerton L, et al. Invasive meningococcal disease in patients with complement deficiencies: a case series (2008-2017). *BMC Infect Dis* 2019 Jun 14;19(1):522.
2. Rosain J, Hong E, Fieschi C, Martins PV, El Sissy C, Deghmane AE, et al. Strains responsible for invasive meningococcal disease in patients with terminal complement pathway deficiencies. *J Infect Dis* 2017 Apr 15;215(8):1331–8.
3. Claus H, Maiden MCJ, Maag R, Frosch M, Vogel U. Many carried meningococci lack the genes required for capsule synthesis and transport. *Microbiology (Read)* 2002 Jun;148(Pt 6):1813–9.
4. Jaton K, Ninet B, Bille J, Greub G. False-negative PCR result due to gene polymorphism: the example of *Neisseria meningitidis*. *J Clin Microbiol* 2010;48(12):4590–1.
5. Dolan Thomas J, Hatcher CP, Satterfield DA, Theodore MJ, Bach MC, Linscott KB, et al. *sodC*-based real-time PCR for detection of *Neisseria meningitidis*. *PLoS One* 2011;5(5):e19361.
6. Kurose S, Onozawa K, Yoshikawa H, Yaita K, Takahashi H, Shimono N, et al. Invasive meningococcal disease due to a non-capsulated *Neisseria meningitidis* strain in a patient with IgG4-related disease. *BMC Infect Dis* 2018;2;18(1):146.
7. Fijen CA, Kuijper EJ, Tjia HG, Daha MR, Dankert J. Complement deficiency predisposes for meningitis due to nongroupable meningococci and *Neisseria*-related bacteria. *Clin Infect Dis* 1994;18(5):780–4.

Claire Duployez*
CHU Lille, 59000 Lille, France
Univ. Lille, 59000 Lille, France

Caroline Loiez
CHU Lille, 59000 Lille, France

Frédéric Wallet
CHU Lille, 59000 Lille, France
Univ. Lille, 59000 Lille, France

Laure Marceau
CHU Lille, 59000 Lille, France

Myriam Simon
Hôpital Victor Provo, 59100 Roubaix, France

Ala-Eddine Deghmane
Muhamed-Kheir Taha
Institut Pasteur, Invasive Bacterial Infections Unit and
National Reference Centre for Meningococci and
Haemophilus Influenzae, 75724 Paris, France

Anne Vachée
Hôpital Victor Provo, 59100 Roubaix, France

*Corresponding author. Laboratoire de Bactériologie,
Institut de Microbiologie, Centre de Biologie Pathologie,
F-59037 Lille CEDEX, France.
E-mail address: claire.duployez@chu-lille.fr (C. Duployez)