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RESEARCH ARTICLE

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Establishment and characterization of canine mammary tumoroids for translational research

Antonella Raffo-Romero¹, Soulaïmane Aboulouard¹, Emmanuel Bouchaert^{1,2}, Agata Rybicka^{1,2}, Dominique Tierny^{1,2}, Nawale Hajjaji^{1,3}, Isabelle Fournier^{1,4}, Michel Salzet^{1,4*†} and Marie Duhamel^{1*†} 

Abstract

Background Cancer heterogeneity is a main obstacle for the development of effective therapies, as its replication in in vitro preclinical models is challenging. Around 96% of developed drugs are estimated to fail from discovery to the clinical trial phase probably because of the unsuitability and unreliability of current preclinical models (Front Pharmacol 9:6, 2018; Nat Rev Cancer 8: 147–56, 2008) in replicating the overall biology of tumors, for instance the tumor microenvironment. Breast cancer is the most frequent cancer among women causing the greatest number of cancer-related deaths. Breast cancer can typically be modeled in vitro through the use of tumoroids; however, current approaches using mouse tumoroids fail to reproduce crucial aspect of human breast cancer, while access to human cells is limited and the focus of ethical concerns. New models of breast cancer, such as companion dogs, have emerged given the resemblance of developed spontaneous mammary tumors to human breast cancer in many clinical and molecular aspects; however, they have so far failed to replicate the tumor microenvironment. The present work aimed at developing a robust canine mammary tumor model in the form of tumoroids which recapitulate the tumor diversity and heterogeneity.

Results We conducted a complete characterization of canine mammary tumoroids through histologic, molecular, and proteomic analysis, demonstrating their strong similarity to the primary tumor. We demonstrated that these tumoroids can be used as a drug screening model. In fact, we showed that paclitaxel, a human chemotherapeutic, could kill canine tumoroids with the same efficacy as human tumoroids with 0.1 to 1 μ M of drug needed to kill 50% of the cells. Due to easy tissue availability, canine tumoroids can be produced at larger scale and cryopreserved to constitute a biobank. We have demonstrated that cryopreserved tumoroids keep the same histologic and molecular features (ER, PR, and HER2 expression) as fresh tumoroids. Furthermore, two cryopreservation techniques were compared from a proteomic point of view which showed that tumoroids made from frozen material allowed to maintain the same molecular diversity as from freshly dissociated tumor.

[†]Michel Salzet and Marie Duhamel contributed equally to this work.

*Correspondence:

Michel Salzet
michel.salzet@univ-lille.fr

Marie Duhamel
marie.duhamel@univ-lille.fr

Full list of author information is available at the end of the article



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Conclusions These findings revealed that canine mammary tumoroids can be easily generated and may provide an adequate and more reliable preclinical model to investigate tumorigenesis mechanisms and develop new treatments for both veterinary and human medicine.

Keywords Breast cancer, Dog patients, Tumoroid, Biobank, Drug screening

Background

A major obstacle in preclinical drug development for cancer is the lack of appropriate cell culture model systems. Two-dimensional cancer cell lines are frequently used for the first screening of newly developed drugs and for the study of cancer development. However, cancer cell lines completely lack interaction with the tumor microenvironment, which is a crucial reason for drug resistance [1, 2]. Mouse models present also several drawbacks leading to difficulties in the translation to human diseases. Such models do not fully recapitulate the diversity and architecture of the primary disease, thus providing inaccurate analysis of tumor pathogenesis and sensitivity to therapy. Around 96% of the drugs fail from discovery to the clinical trial phase, probably because the preclinical models are not close enough to the tumor biology in patients [3].

Tumoroid cultures represent a robust three-dimensional (3D) *in vitro* system that faithfully recapitulate the genetic and phenotypic characteristics of the tumor from which they are derived [4]. The 3D tumoroid system has been utilized to study different types of cancers [5–8]. Tumoroids can serve to better understand the biology but also to test drug efficacy *in vitro* before clinical trials in human patients. Most of the tumoroid studies have been conducted on mouse and human tissue samples. Mouse tumor tissues do not fully recapitulate the human disease and therefore are not the best models for human translation. On the other hand, the use of human samples is the optimal solution but the difficulty to access to sufficient amounts of fresh tissues (in general only biopsies or small pieces of tissues from tumor resection are available) and ethical issues can slow down the large-scale screening of new drugs. That is why it is of prime importance that new models emerge than can fully and faithfully recapitulate the human disease. Moreover, as tumoroids are found to have more and more relevance and applications, large-scale production of tumoroids become inevitable but presents many challenges due to the difficulty of accessing large quantities of human fresh tissue.

In that regard, companion dogs with spontaneous tumors present a unique, ethical, non-experimental model for comparative research and drugs development [9–11]. Canine mammary tumor (CMT) is the third most common type of cancer in dogs and first in bitches with an incidence of around 230 cases per 100,000 dogs per year [12–14]. It possesses several advantages over highly

inbred and genetically modified laboratory animals, such as clinical profile (age at onset, predominance of carcinomas, and type of metastases), genetics (role of BRCA1/2, overlapping gene signature), and molecular similarities with its human counterpart [15, 16]. CMT, the same as human breast cancer (BC), can be characterized by expression of estrogen, progesterone, and HER2 receptors. The involvement of companion dogs with spontaneous CMT in translational oncology is already seen in numerous publications and several ongoing clinical trials [17]. Canine tumoroids developed from dog patients with spontaneous CMT could therefore provide a more representative and ethical translational model to test drug efficacy and toxicity in pre-human studies, as well as canine tumoroids could be an innovative screening tool in drug discovery, while reducing the number of experimental animals needed for *in vivo* studies. Treatment options of human breast cancer depend mainly on the subtype and stage of disease but is typically based on surgery followed by adjuvant radiotherapy, chemotherapy, and/or hormone therapy. As in human cancer patients, disease staging for CMT is mandatory before beginning treatment. Surgery is the most common treatment option for CMT. Chemotherapy is not commonly used in dogs; however, a case study has shown that the combination of 5-fluorouracil and cyclophosphamide improved overall survival in dogs with malignant mammary tumors [18].

Few canine tumoroids studies have been made so far [19–22]; one of them has developed tumoroids from canine normal and tumor breast stem cells [23]. None of them have developed tumoroids from CMT heterogeneous tissue recapitulating the tumor microenvironment.

The enormous potential of tumoroids as preclinical models has given rise to the development of tumoroid biobanks. Tumoroid biobanks have been obtained from various tumor tissues [24]. However, there is a lack of knowledge about cryopreservation procedures and whether the cryopreserved tumoroids maintain similar molecular and functional features as their fresh counterparts.

In this study, we have developed for the first time tumoroids from heterogeneous canine mammary tumor tissues. We have demonstrated morphologic, histologic, and molecular stability between fresh CMT tumoroids and cryopreserved tumoroids. However, tumoroids made from frozen CMT material show more similarities

with fresh tumoroids compared to cryopreserved tumoroids. Treatment with a chemotherapeutic drug also confirmed these results. Taken together, our study aimed at creating and characterizing a new biobank of canine mammary tumoroids with similar features as human tissues which can be used for large-scale drug screening in preclinical studies.

Results

Feasibility of tumoroid culture from freshly resected canine mammary tumors

Canine mammary tumor was collected in the operating room at the time of tumor resection. For the characterization of the tumoroids' cultures, 6 on 31 patients of the established biobank were included (Additional File 1: Table 1). For all of them, the resection was a primary mammary tumor. Patient's age ranged between 5 and 14 years old and were all female. Based on the 2010 histologic classification for canine mammary tumors, the 6 tumors were annotated (Additional File 1: Table 1) [25]. The 6 tumors were characterized with the most important and frequent biomarkers of breast cancer: estrogen receptor (ER), progesterone receptor (PR), and HER2.

Among the 6 tumors, 4 have a triple-negative signature, signifying the absence of HER2, ER, and PR proteins expression (TM-02, TM-03, TM-05, TM-06), while 2 tumors are of luminal subtype with PR expression (TM-01) or PR/ER expression (TM-04) (Additional File 2: Fig. S1 and Additional File 1: Table 1). In addition, the positive Ki67 labeling of each tumor was evaluated. We found that all 6 tumors showed Ki67 positive cells, with variable levels (Additional File 3: Fig. S2).

After tumor resection, the tumor fragment was divided into three pieces: the first piece was kept fresh for tumoroid culture generation, the second one was frozen without prior fixation, and the last piece was fixed in PFA and cryopreserved (Fig. 1A). Frozen tissue was used for proteomics while fixed tissue was used for histology.

Fresh tissue pieces were mechanically and enzymatically dissociated to obtain single cell suspensions which were plated in Matrigel drops and overlaid with optimized mammary tumoroids culture medium. Cultures were followed by microscopy for evidence of tumoroids formation. We successfully generated tumoroid cultures from 31 of 33 tumor samples, an establishment success rate of 94%, with long-term expansion. Indeed, all

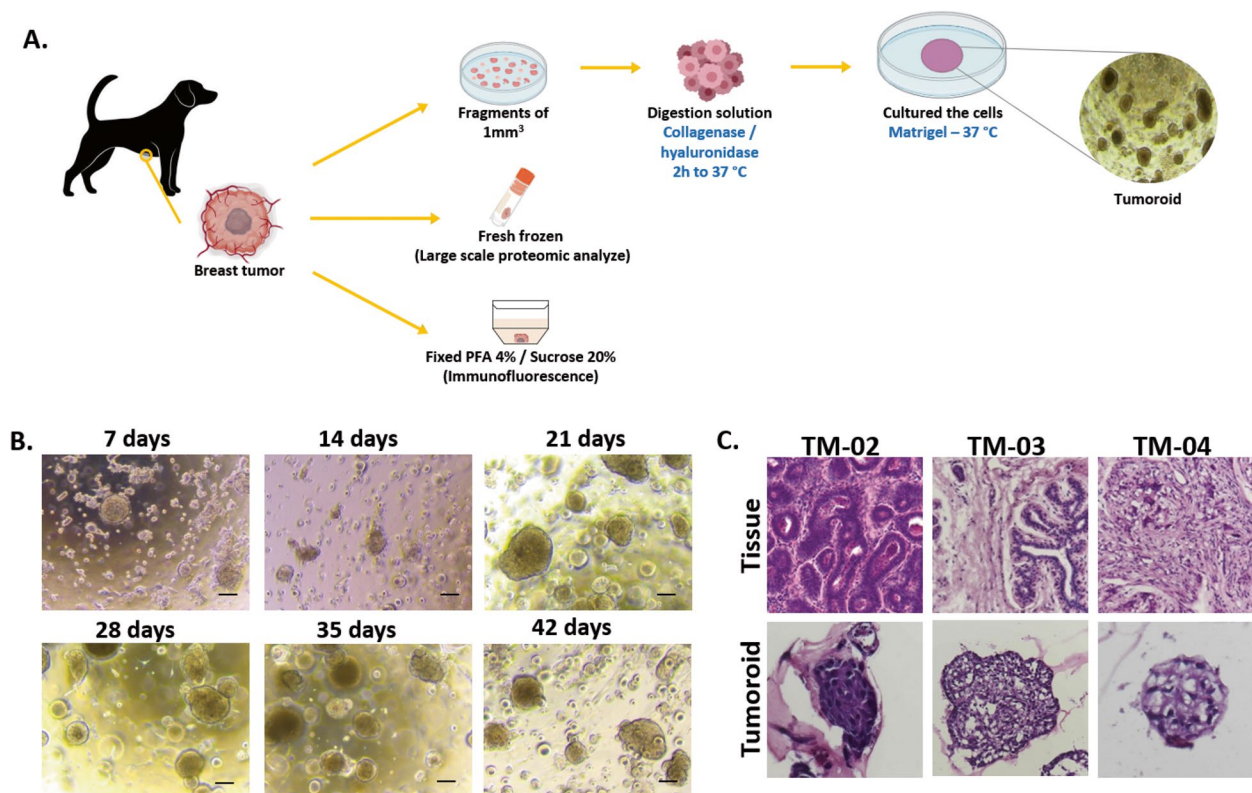


Fig. 1 Breast cancer tumoroids culture established from canine patient. **A** Diagram presenting the strategy used to culture tumoroids from a canine mammary tumor. **B** Representative images of canine tumoroids at different time points of culture. Scale bar (200 μm) is indicated. **C** H&E staining comparison between tumoroids and the tumor of origin for three different dog patients ($n = 3$)

tumoroids were grown for at least 42 days (6 passages) (Fig. 1B). Majority of tumoroid lines were cryopreserved. The tumoroids morphologically reflected the original tumor they were derived from (Fig. 1C). Tumoroids presented patient-specific heterogeneous morphologies, ranging from compact structures (TM-02) to more irregular structures (TM-03 and TM-04).

Canine mammary tumoroids can be generated from both fresh and frozen cells and can be cryopreserved with similar histological and molecular features

Next, we wanted to evaluate whether tumoroids could be generated from frozen cells while keeping the same characteristics as fresh tumoroids. From the primary tumor sample, we divided the cell suspension into two parts: one part kept fresh for direct tumoroid formation, named “fresh tumoroids” thereafter, and the second part was frozen for indirect tumoroid formation, named “tumoroids from frozen cells” thereafter. In addition, in order to characterize our biobank, we wanted to make sure that cryopreservation did not affect the tumoroids features. We therefore compared these two types of tumoroid cultures to thawed tumoroids, named “frozen tumoroids” thereafter (Fig. 2A). Tumoroids from these different culture conditions were left in culture during 4–5 weeks (date 1) or 6–7 weeks (date 2) and compared to study tumoroids drift overtime.

Histologic and molecular drifts of tumoroids after cryopreservation and after long-term culture were studied. First, the efficiency of tumoroids formation was evaluated in all 6 tumors in triplicate. All tumoroids show an exponential growth curve. Almost all of them increase their growth between 230 and 335% compared to the initial time (T0) after 7 days of culture. TM-01 showed a more modest growth, reaching values of about 85% augmentation of growth after 7 days of culture (Fig. 2B and Additional File 4: Table 2). In addition, the culture of tumoroids was successful for each culture condition and after serial passages as well (Fig. 2C). Tumoroid formation was not found visually different between cryopreserved cells and fresh cells. Tumoroid cultures from fresh and frozen cells could be similarly long-term cultured and passaged (Fig. 2C). At the histological level, tumoroids derived from fresh cells, frozen cells, or after cryopreservation retained the same architecture. Figure 2D presents representative images of H&E staining of tumoroids derived from three different tumors. Tumoroids derived from TM-02 were compact while tumoroids derived from TM-03 and TM-04 were more irregular whatever the culture condition and time in culture. The freezing procedure did not affect tumoroids morphology.

The ER, PR, and HER2 expression profiles of breast cancer tumoroids were compared with their original breast cancer tissues. For this, 3 tumors were used: TM-03 (triple-negative subtype), TM-04 (luminal subtype) (Fig. 3), and TM-05 (triple-negative subtype) (Additional File 5: Fig. S3). The results showed that the tumoroids maintain the same expression profile of the tumor of origin. In the case of TM-03 and TM-05, tumoroids present a triple-negative subtype as the tumor they are derived from. In the case of TM-04 tumoroids, we can observe ER and PR positive cells similar to the tumor of origin. This result supports those previously demonstrated in humans and dogs, in which the majority of tumoroids reflect the same histological characteristics as the tumor from which they originate [6].

Finally, we verified if the proliferation of the tumoroids in all three conditions was similar. We used tumors of the two different subtypes, a luminal TM-04 and a triple-negative TM-05, to answer this question. The proliferative activity of the tumoroids was determined by the percentage of Ki67+ cells with respect to the total number of cells. Proliferation activity of tumoroids do not show a significant difference between Fresh, Frozen and FrozenCell tumoroids. In TM-04, the percentage of Ki67+ cells remained around 7% for all conditions (Fresh—7.64%, Frozen—7.79% and FrozenCell—7.41%), as well as in the TM-05, around 7% were Ki67+ (Fresh—8.61%, Frozen—6.96% and FrozenCell—6.29) (Fig. 4 and Additional File 4: Table 2).

Similar proteomic profiles are observed between tumoroids generated from fresh and frozen cells over time while cryopreservation seems to trigger a more pronounced molecular drift

We have shown that the freezing procedure as well as the passages did not impact the morphology of tumoroids neither their histological features. In order to understand, whether the frozen tumoroids or tumoroids made from frozen cells kept similar molecular profiles as the fresh tumoroids or the original tumor, we have performed a large-scale unbiased proteomic analysis. The study was carried out on 3 tumors: TM-01, TM-02, and TM-03, of which the three types of tumoroids were made and compared with each other and with the original tumor. For this, the extracted proteins were quantified and the same amount of proteins was used.

To understand if there was any molecular drift over time, we have analyzed the proteome of tumoroids at two dates of tumoroid passage (D1 and D2), 1796 proteins were identified in the six conditions: D1-Fresh, D1-FrozenCell, D1-Frozen, D2-Fresh, D2-FrozenCell, and D2-Frozen (62% of all the proteins identified) (Fig. 5A) (Additional File 6: Table 3). The D1-Frozen tumoroids

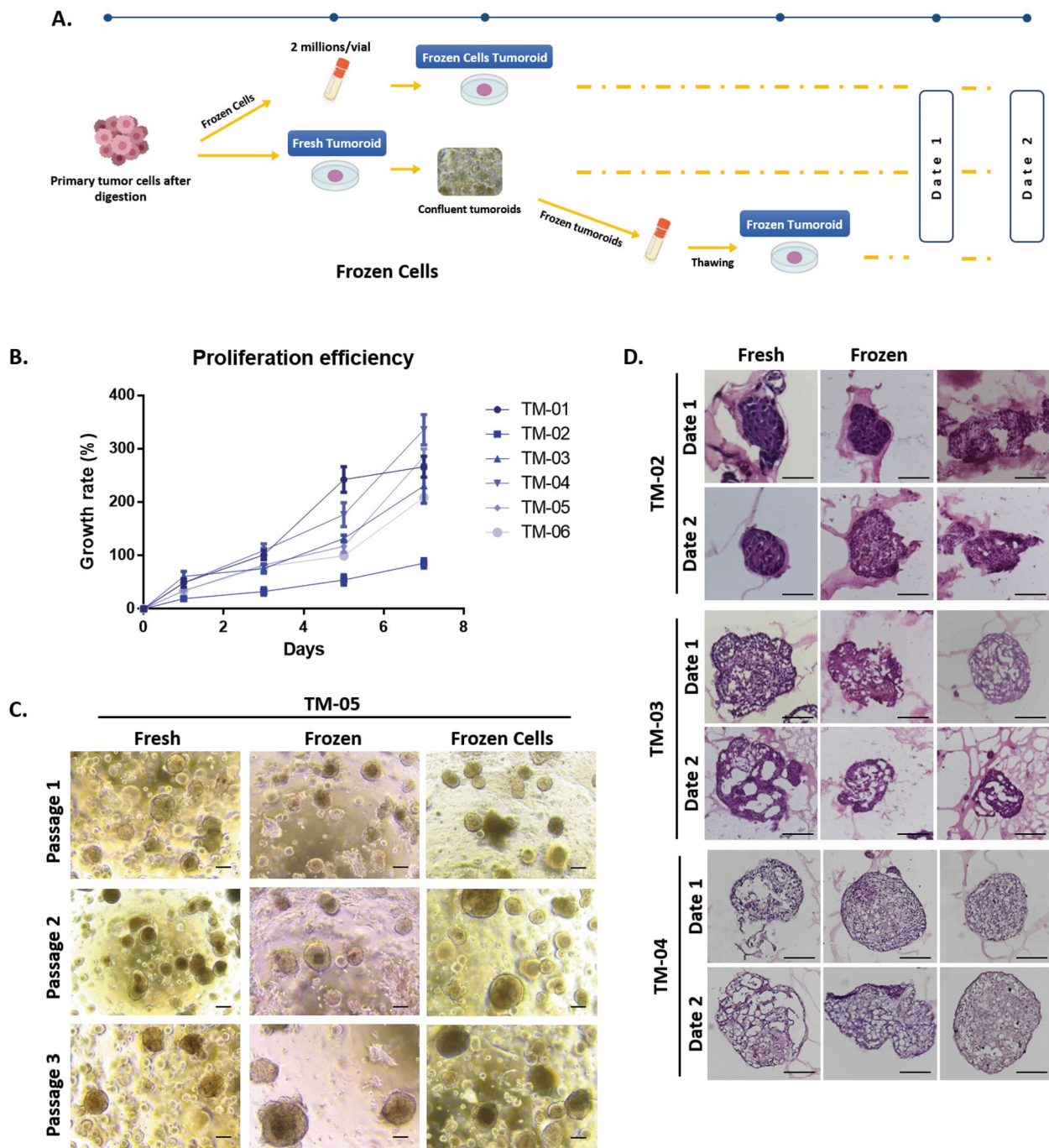


Fig. 2 Study of canine mammary tumoroids drift. **A** Diagram showing the strategy used to generate the different types of tumoroids. After tumor digestion, a part of the cells were frozen and then thawed to generate the “Frozen Cell Tumoroids.” The remaining cells were used to generate the “Fresh tumoroids.” A part of these tumoroids were cryopreserved and then thawed, corresponding to the “Frozen Tumoroids.” The three types of tumoroids were compared at the same time point post-culture at date 1 (4–5 weeks) or date 2 (6–7 weeks). **B** Evaluation of the efficiency of tumoroids formation for the 6 cases in triplicates ($n = 6$). The average of the triplicates is shown, and error bars mean SD. **C** Representative images of the three types of canine tumoroids at passage 1, 2, and 3 after date 1. Scale bar (200 μm) is indicated. **D** H&E staining comparison of fresh, frozen, and frozen cells tumoroids. Scale bar = 100 μm ($n = 3$)

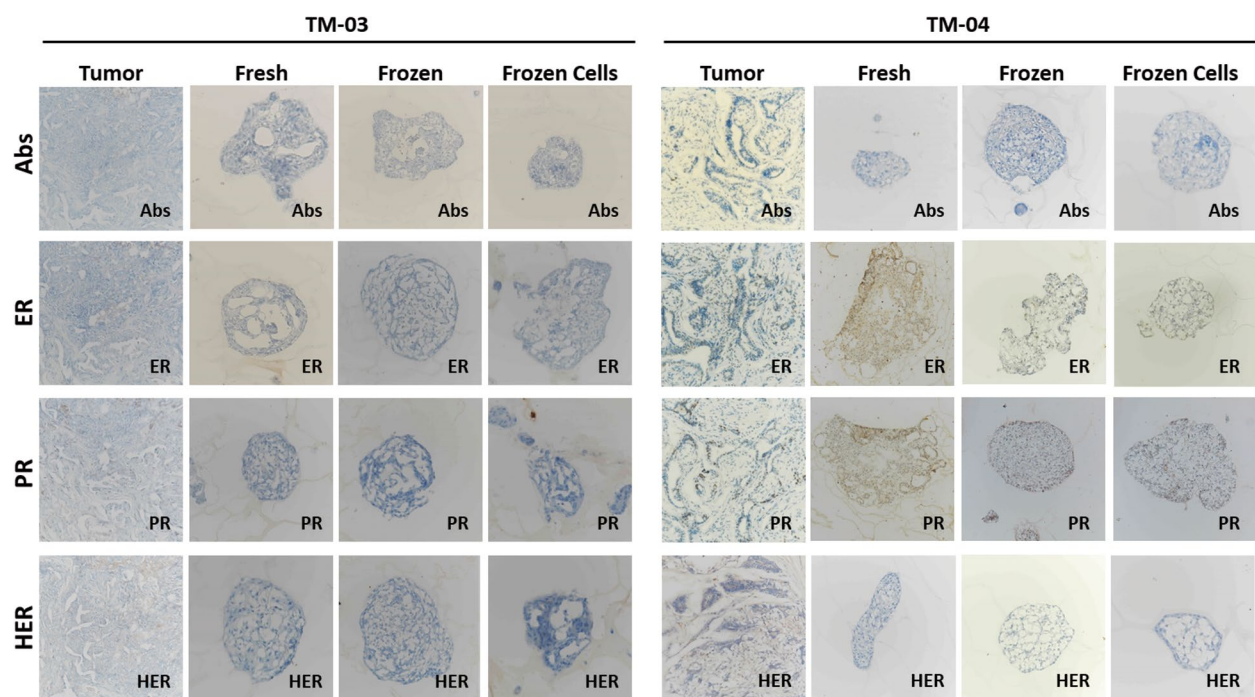


Fig. 3 Histology and receptor status (ER, PR, HER2) of breast cancer tumoroids. Comparative histological and immunohistochemical images of breast cancer tumoroids and their original breast cancer tissues

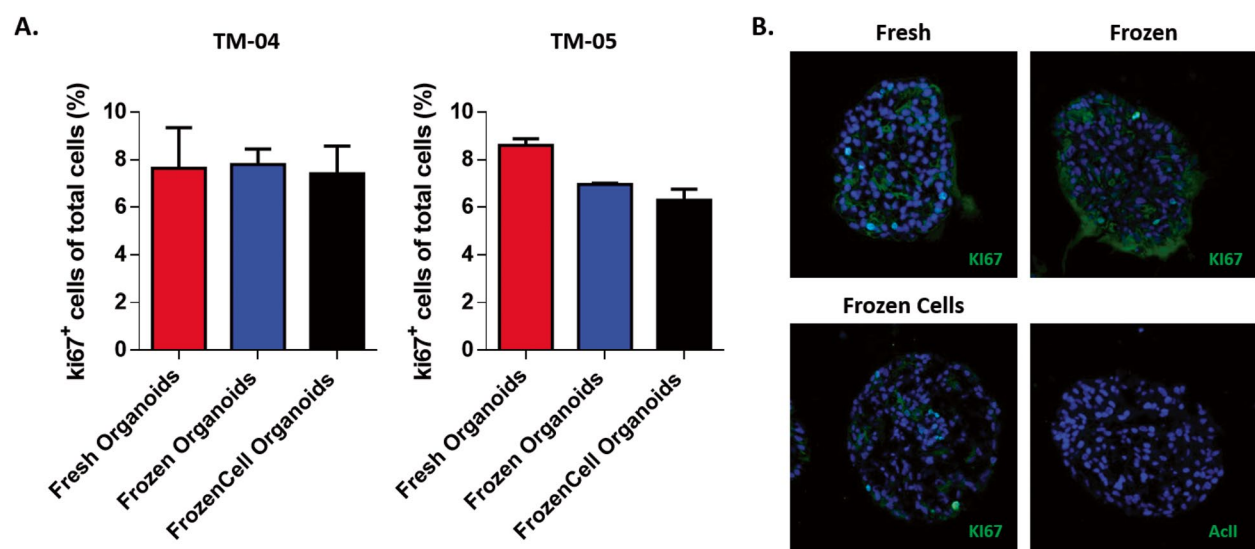


Fig. 4 Proliferative activity of the tumoroids. **A** Comparative quantification of the percentage of Ki67 + cells in tumoroids Fresh, Frozen, and FrozenCell of the tumor TM-04 and TM-05. The average of the triplicates is shown and error bars mean SEM ($n = 3$, P value ≤ 0.05). **B** Representative images of canine Ki67 staining (green) of fresh, frozen and frozen cells tumoroids of the tumor TM-05

seem to be the most different compared to all the other conditions as shown on Fig. 5A. In fact, 343 proteins were identified in all conditions except in D1-Frozen. This may be due to a lack of protein diversity in this condition, as all samples were quantified to have the same amount of proteins. However, if we observe more closely the proteins lacking in the condition of D1-Frozen (Fig. 5B), we found several proteins involved in cellular metabolic process and energy pathways such as the metabolic process of cellular aromatic compound (with HMOX2,

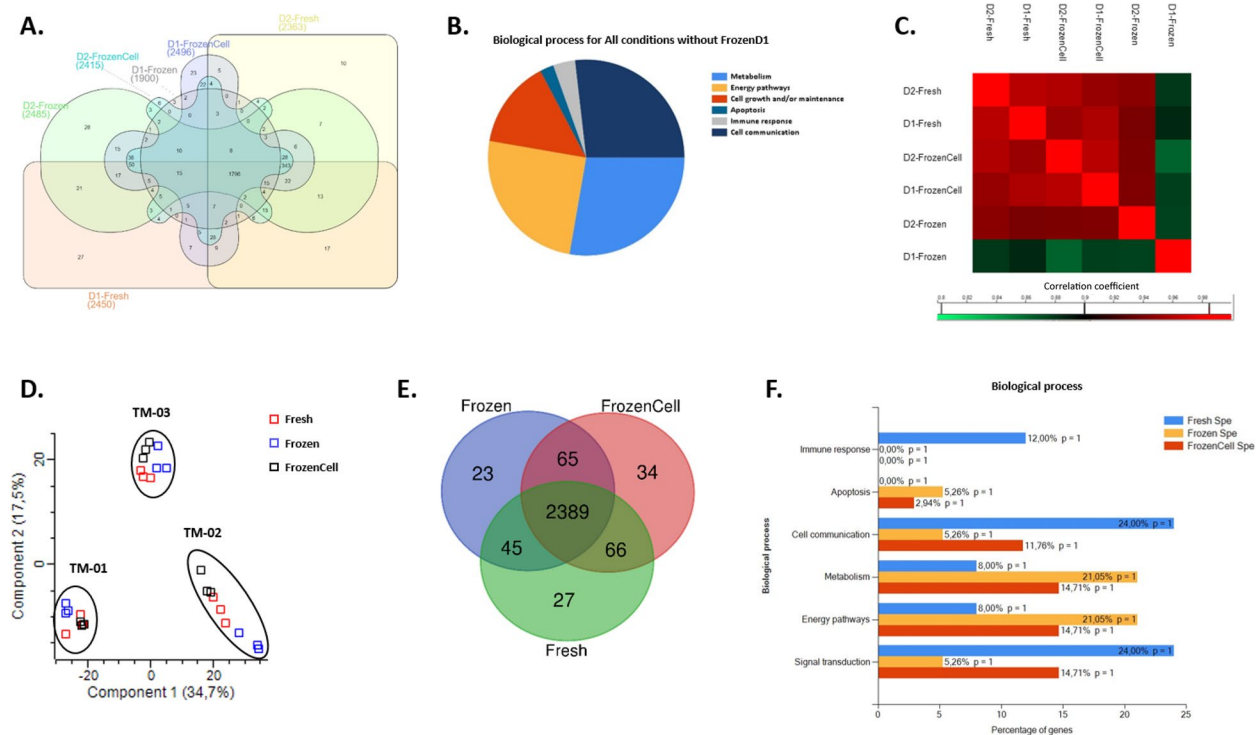


Fig. 5 Proteomics analysis of CMT tumoroids. **A** Venn diagram representing specific proteins identified in the Fresh, Frozen, and FrozenCell tumoroids at date 1 and date 2 ($n = 3$ for each condition). **B** Biological processes of the proteins identified in common except in Frozen D1 tumoroids ($n = 3$). **C** Matrix correlation studies between the two dates in the tree different conditions ($n = 3$ for each condition). **D** PCA analysis of the proteomics data from the tree different tumoroid conditions ($n = 3$ for each condition). **E** Venn diagram representing specific proteins in the Fresh, Frozen, and FrozenCell tumoroids ($n = 3$ for each condition). **F** FunRich biological process distribution of the specific proteins identified in Fresh, Frozen, and FrozenCell tumoroids ($n = 3$ for each condition)

FAH, ACSL4, PMVK, AARS2, GGH, FECH, PRPSAP1, HMGCS1, and ADA) and the cellular catabolic process (with HMOX2, PLA2G4A, MANBA, FAH, HEXB, ARG2, BLMH, SARDH, ECI1, BCKDHB, and ADA). In addition, we found proteins involved in the inhibition of apoptosis (API5, RIPK3, FIS1, FAF1, and CTNBL1), in cell growth (GOLGA3, SOD2, COL5A2, SYNE2, ARVCF, GOLGA2, PARVA, CGN, ADD3, TIMP3, DSG2, VCAN, PAK4, ELMO3, FKBP15, WASL, SYNPO, TRIOBP, ACTR10, GAN, ACTN4, LGALS7, ANK2, ITM2B, RRS1, ARPC1A, TNXB, SPON1, FGL2, CAV1), and cell communication (ATP6AP2, ITGAV, LDLR, PPFIBP2, SBF1, SORBS2, ABR, NCKIPSD, NAE1, PRPF6, EPB41L2, BMP2K, APP, RAB4A, MTOR, RAP1B). Very interestingly, proteins involved in immune response processes were enriched (NRP1, PROCR, ALCAM, CD109, LBP, ST6GAL1, LRRC8A, CFB), these proteins which were not found in the D1-Frozen condition have a proliferative, immune, and anti-apoptosis profile, demonstrating a lack of these biological processes in the D1-Frozen condition.

Likewise by a Pearson correlation analysis, hierarchical clustering of all the samples based on the correlation coefficients between them revealed higher similarity

between Fresh and FrozenCell tumoroids at dates 1 and 2. Frozen tumoroids were more different, specifically at date 1 (Fig. 5C). The similarity of D1-Frozen with the other conditions was less than 87%, while all the other conditions showed more than 95% similarity. The duration of the tumoroids culture did not seem to have a big impact on their proteomic profiles. The fact that D1-Frozen tumoroids were more distinct suggests that the tumoroids should be preferentially left in culture long enough to recover after freezing, which was not observed from D1-FrozenCells.

Knowing that the time in culture did not impact too much their molecular profile, we wanted to verify whether if the culture condition impacted or not their proteome. For that, we have compared the proteomic profiles of tumoroids from three culture conditions: fresh, frozen, and tumoroids made from frozen cells. First of all, the principal component analysis (PCA) based on the LFQ values of the protein identification showed that the samples were grouped by tumor and not according to the type of culture condition (Fig. 5D). This sample grouping by PCA means that there was a high level of similarity between the biological replicates of each condition

but also between the tumoroids without influence of their culture condition. Furthermore, a Venn diagram showing the number of common and unique proteins in all conditions showed that a majority of proteins were identified in all three conditions of culture (2389 proteins, representing 90% of all proteins). However, some proteins were found specifically expressed in each condition: 27 identified specifically in fresh tumoroids, 23 in frozen tumoroids, and 34 in tumoroids made from frozen Cells (Fig. 5E) (Additional File 7: Table 4). Based on the GO term enrichment analysis of the biological processes using FunRich software, we observed that these proteins, specifically expressed in each condition, were linked to different biological processes (Fig. 5F). An effect on the cellular metabolic and energy pathways was found in Frozen and FrozenCell tumoroid conditions compared to Fresh tumoroids (with: *AMY1A*, *SDR9C7*, *CDA*, *ARG1*, *FKBP*, *NDUFB10*, *DDO*, *ADH5*, and *SIAE*). In addition, in the Frozen and FrozenCell condition, we have identified proteins involved in apoptosis like the apoptosis facilitator *BCL2L13*, *ATG5*, and *TXNRD2*. In the Fresh tumoroids, a higher number of proteins linked to cell communication and to signal transduction were identified. Interestingly, some of the specific proteins identified in fresh tumoroids were involved in the immune response, such as *GZMB* (Granzyme B) expressed by cytotoxic T and NK cells, the cell adhesion molecule *Siglec1* (Sialoadhesin) expressed by macrophages, as well as *CD163* (Cluster Differentiation 163), a marker of anti-inflammatory macrophages, *AMBP* (alpha-1 microglobulin/bikunin) precursor of a glycoprotein synthesized by lymphocytes and *CD177* (*CD177* molecule), a marker of neutrophil activation.

In order to better understand the differences linked to the culture conditions, an analysis of the variation of abundance of common proteins to all conditions (2389 proteins) was carried out, using a multiple sample test ANOVA with an FDR of 0.05. A total of 489 proteins showed significantly different expression between the three groups. These specific variations were analyzed by hierarchical clustering and then illustrated by a Heatmap (Fig. 6A). Six clusters of proteins were identified: one cluster representing the specific underexpressed proteins and one representing the specific overexpressed proteins for each condition (Additional File 8: Table 5). Based on over- and under-expressed proteins, fresh tumoroids and tumoroids made from frozen cells showed more similarities compared to frozen tumoroids, as observed before. In order to characterize the impact of these proteins, the GO terms analysis of each cluster was carried out with Cytoscape and ClueGO software, allowing to generate the networks connecting the proteins overexpressed (in red) and underexpressed (in green) to their biological process.

The results showed that in the fresh condition (clusters 3–4) (Fig. 6A, B), the cellular respiration and the amino acid metabolism were underexpressed compared to frozen tumoroids and FrozenCell tumoroids, which could be explained by the cryopreservation. In another hand, the assembly of the cell-substrate junction and the RNA translation by RNA polymerase appeared to be overexpressed in the Fresh condition compared to the other conditions, and the biological process of cell adhesion which can be explained by a conservation or a faster reformation of the extracellular matrix and the cell compaction. For the FrozenCell condition (clusters 1–2) (Fig. 6C), there is a higher abundance of proteins linked to chromatin remodeling and cellular metabolism as we observed before (*UBA52*, *PSMC1*, *PSMD5*, *RAB7A*, *HSPA9*, *HSPA5*, *HSPA8*, *DDB1*, *MDH2*, *SLC25A12*). Again, a high metabolic activity can be a consequence of freezing. In addition, the chromatin remodeling involved in the cell division cycle can be linked to a process of multiplication and recovery from freezing that seems important in this condition.

Finally, in frozen tumoroids (clusters 5–6) (Fig. 6D), many proteins related to protein translation, extracellular matrix assembly, vesicle-mediated transport, and RNA nuclear export were found overexpressed. The biological process of vesicle-mediated transport can be subdivided on transport of extracellular vesicles, vesicle budding from membrane, vesicle targeting, vesicle coating, and COPPII-coated vesicles cargo with the presence of distinct protein such as *ARCN1*, *AP2A1*, *DYNC1H1*, *AP2B1*, *ANXA7*, and *SEC13*. In addition, the biological process of protein translation, protein-RNA nuclear export, and telomerase RNA localization are represented by proteins like eukaryotic translation initiation factor (4A-III, 3 subunit A, 3 subunit L, 3 subunit B, 3 subunit E, among others) and 40S, 60S ribosomal protein (*RPL10*, *RPL13A*, *RPL14*, *RPL15*, *RPS11*, *RPS13*, *RPS18*, *RPS28*, *RPS3*, *RPS8*, *RPSA*). These analysis highlight a dysfunction in the translation pathways that we know contribute to cancer progression, for example, in the deregulation of ncRNAs that leads to aberrant protein translation in cancers [26], and which seems more active in frozen tumoroids.

On the contrary, the underexpressed proteins are related to the metabolism of amino acids or nucleotides, negative regulation of cytokines, immune effector process, and the organization of the cytoskeleton, cell adhesion, and death. Different metabolic pathways were touched, such as dicarboxylic acid metabolic process, purine ribonucleotide biosynthetic process, pyruvate metabolic process, and generation of precursor metabolites and energy. Regarding the organization of the cytoskeleton and cell adhesion, different isoforms of laminin, collagen, catenin and Coronin-1B were found to be underexpressed in this

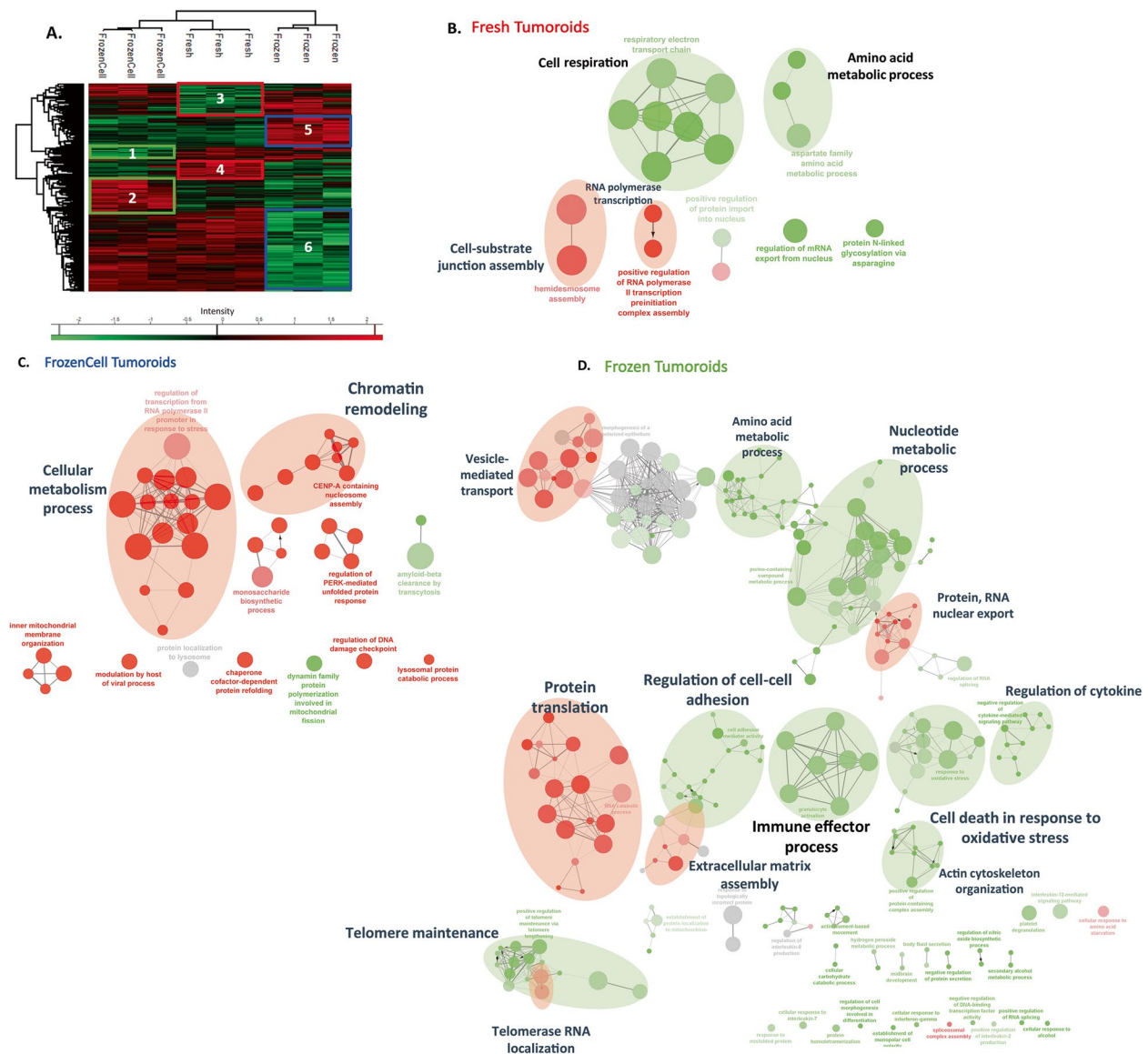


Fig. 6 Proteomics analysis of Fresh, Frozen, and FrozenCells tumoroids. **A** Hierarchical clustering of the most variable proteins between the 3 conditions ($n = 3$ for each condition, ANOVA with permutation-based FDR < 0.05). Network of proteins overexpressed (red) or underexpressed (green) in Fresh (**B**), FrozenCell (**C**), and Frozen (**D**) tumoroids and their associated GO terms. The networks were enriched through addition of STRING network to the identified proteins using ClueGO application on Cytoscape

condition. Apoptosis and cell death proteins (CYP1B1, HSPA1, ARL6IP5, TRAP1, among others) were also found underexpressed. In addition, we have identified underexpressed proteins linked to a regulation of cytokines and to the immune effector process, in which we find proteins: CD44, thrombospondin-1, SAMHD1, TINAGL1, GAA, LGALS9 (Galectin), among others.

The Cytoscape and ClueGO analysis shows that Fresh and FrozenCell conditions have limited amount of underexpressed and overexpressed proteins, while frozen condition shows three time more proteins with significant variation.

On the other side, even if the FrozenCell is closer to Fresh Tumoroids than Frozen condition, the degree of similarity stays high.

The proteome of canine mammary tumoroids is very similar to the tumor they originate and therefore represent a faithful breast cancer model

We next wanted to determine whether the three different types of tumoroids were similar to the tumor of origin, since the tumoroids will be used as a model of breast cancer.

For this, a Venn diagram (Fig. 7A) was made and showed the number of common and unique proteins in all conditions (Additional File 9: Table 6). It can be observed that a majority of proteins were identified in all three conditions of culture (2138 proteins, representing 74% of all proteins). However, there were specific proteins for each condition: 4 identified specifically in fresh tumoroids, 15 in frozen tumoroids, 15 in tumoroids made from frozen cells, and 154 specific proteins that were found specifically in the tumor. These 154 proteins are involved in different biological processes such as cell growth and/or maintenance, cell communication, signal transduction, and immune response (Fig. 7B). Interestingly, an immunological profile can be observed in the tumor compared to the tumoroids. We found several proteins involved in the complement signaling pathway (complement factor I, complement component 4 binding protein, complement component 5, complement component 7, and complement component 8) involved in immunological response and phagocytosis and found overexpressed in different types of cancer, such as breast cancer [27]. In addition, proteins such as CD93, CD34, CLEC4G, haptoglobin, and joining chain of multimeric IgA and IgM have been identified and are all implicated in immune response. The AOC3 protein was also identified and has been recently described to play a role in the reduction of immune cell recruitment and impacting the promotion and progression of lung cancer [28].

To better understand the differences between tumor and tumoroids, an analysis of the variation of abundance of common proteins was carried out. A total of 512 proteins showed significantly different expression between the four groups. The HeatMap of the specific variations of abundance (Fig. 7C) shows few variations between the three types of tumoroids. Frozen tumoroids were more different compared to the two other culture conditions confirming the previous results. Interestingly, a small cluster of overexpressed proteins was observed in Tumor, Fresh, and FrozenCell tumoroids, while underexpressed in Frozen tumoroids. This result shows again that the conditions more similar to the tumor of origin are the Fresh and FrozenCell tumoroids.

We next focused on the two clusters that showed the significant differences between the tumor of origin and the tumoroids (Additional File 10: Table 7). The HeatMap shows two clusters of proteins over- or underexpressed in tumor compared to the different tumoroids. Functional annotation and characterization of these two clusters of proteins were performed using the FunRich software. The results showed that the biological processes underexpressed in the tumor compared to tumoroids are different processes involved in cellular metabolic and energy process such as cellular respiration (with NDUFV1, GPD2, CS, NDUF51, SDHA) and carbohydrate derivative metabolic process (GPD2, ENO2, CAD, UGGT1, HK1, PFKP, FASN, AMPD2, MOGS), transport with two principal pathway detected: the protein import into nucleus (with

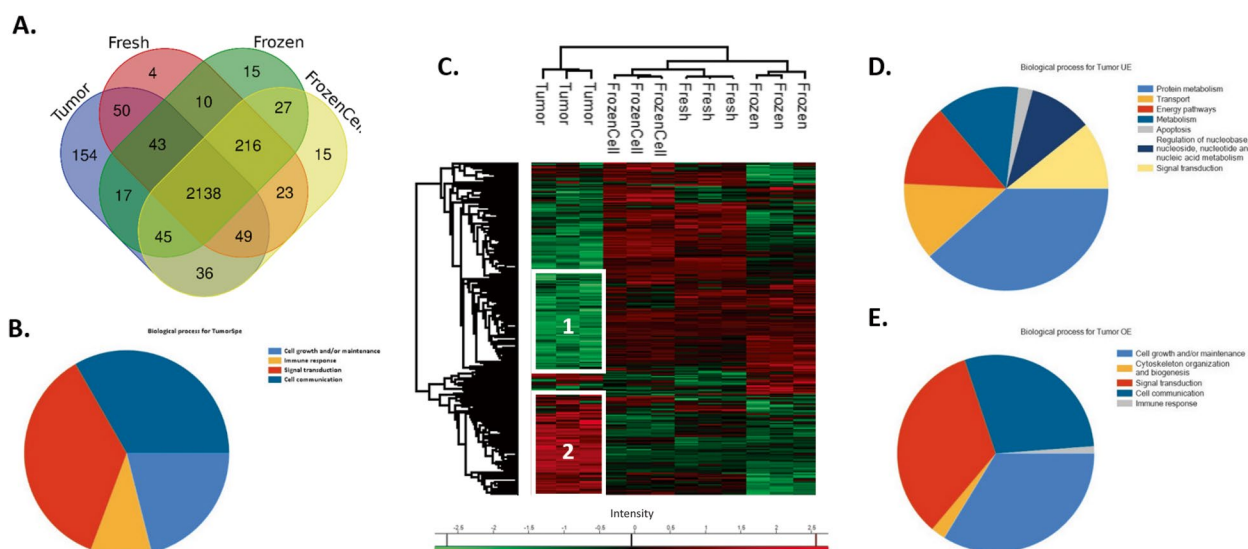


Fig. 7 Proteomics analysis comparing the primary tumor to their derived tumoroids. **A** Venn diagram representing specific proteins in tumor of origin, Fresh, Frozen, and FrozenCell tumoroids ($n = 3$ for each condition). **B** Biological processes of the specific proteins identified in primary tumors using FunRich and ClueGO ($n = 3$ for each condition). **C** Hierarchical clustering of the most variable proteins between the tumor of origin and the 3 tumoroid conditions ($n = 3$ for each condition, ANOVA with permutation-based FDR < 0.05). Biological processes distribution of underexpressed (**D**) and overexpressed (**E**) proteins in tumors compared to tumoroids using FunRich and ClueGO ($n = 3$ for each condition)

presence of interacting protein CSE1L, IPO7, KPNB1, NUP93, TNPO1), and endoplasmic reticulum to Golgi vesicle-mediated transport (with the complex of protein like COPA, COPB1, COPB2, COPG2, SEC13, SEC23A, USO1), apoptosis, and signal transduction (Fig. 7D).

On the contrary, we can observe proteins overexpressed in the tumor compared to tumoroids involved in cell growth and maintenance, actin filament-based process (through ACTG1, ACTN1, ARPC2, EZR, FLNA, GSN, MYH11, MYH14, MYH9, MYL6, MYO1C, TLN1, TPM3, VIM), cell communication, signal transduction, and immune response (Fig. 7E).

These results confirm that in tumoroids we find a higher metabolic activity, especially in frozen tumoroids as demonstrated above. In addition, we can observe apoptotic proteins such as BCL2-associated athanogene 6, heat shock 60 kDa protein 1, PH domain and leucine rich repeat protein phosphatase 2, cell cycle and apoptosis regulator 2, underexpressed in the tumor and consequently overexpressed in the tumoroids. Also, in the tumor, we can find an important proliferative profile demonstrated by the overexpression of proteins involved in cell communication and linked to the organization of the cytoskeleton and cell growth (actin beta-like 2, collagen type VI, tubulin, beta 4B class IVb, lamin A/C, actin alpha 2, actin-related protein, among others). Finally, we can find an immune profile more present in the original tumor.

Canine mammary tumoroids can be used to test human drugs and cryopreservation of tumoroids does not impact drug response

In order to evaluate canine mammary tumoroids as useful tools for translational in vitro drug screening studies, we performed cell viability assays in presence of a chemotherapeutic agent used in human medicine, paclitaxel. Tumoroids were treated with paclitaxel for 7 days before cell viability was measured. Representative images of tumoroids derived from TM-05 tumor are shown in Fig. 8A demonstrating drug sensitivity depending on the drug concentration. Using 6 concentrations of paclitaxel, we generated dose–response curves (Fig. 8B and Additional File 4: Table 2). First, we could demonstrate that tumoroids derived from fresh canine mammary tumors responded well to paclitaxel with an IC₅₀ ranging from 0.1 to 1 μ M. 0.1 μ M paclitaxel was needed to kill 50% of tumor cells for TM-04 and TM-06, while around 1 μ M was needed for TM-05 demonstrating higher resistance to paclitaxel.

We next compared the paclitaxel response between fresh tumoroids, frozen tumoroids, and tumoroids made from frozen cells in order to verify whether cryopreserved tumoroids could represent faithful models for

drug testing. Killed curves from these three culture conditions were similar for the three tumors tested (Fig. 8B). As we have observed before with the proteomic analysis, fresh tumoroids and tumoroids derived from frozen cells were the most similar in term of paclitaxel responses. Nonetheless, tumoroids derived from frozen cells appear to become slightly more resistant at higher concentrations of paclitaxel (Fig. 8A, B) compared to fresh tumoroids. Indeed, an increase of viability of tumoroids derived from frozen cells can be observed for each tumor at a concentration of 100 μ M. A 50% viability of tumoroids was measured in this condition, while only 35% of cells were viable in fresh tumoroids (Fig. 8B). Finally, the response to paclitaxel of frozen tumoroids appears to be slightly different compared to the two other conditions, even if not significant. For TM-04 and TM-06, fresh tumoroids and tumoroids made from frozen cells seem to be more sensitive whatever the concentration of paclitaxel used compared to frozen tumoroids. For TM-05, the three curves are more similar. A 50% viability decrease of fresh tumoroids and tumoroids made from frozen cells is observed between 0.1 and 1 μ M of drug, while between 1 and 10 μ M of paclitaxel are needed to kill 50% of tumor cells in the frozen tumoroids condition (Fig. 8B).

In the end, we have shown that paclitaxel response between luminal subtype CMT tumoroids and human breast tumoroids was similar with 0.1 μ M of paclitaxel needed to kill 50% of tumor cells. In contrast between triple-negative subtype CMT tumors and human breast tumors, although they show a similar response trend, they respond differently to the range of paclitaxel concentrations. CMT tumors (TM-05 and TM-06) show an amount between 0.1 and 1 μ M paclitaxel needed to kill 50% of tumor cells, whereas the human tumor shows around 10 μ M paclitaxel (Fig. 8C and Additional File 4: Table 2). Further study on triple-negative tumors will be necessary to make more detailed conclusions. Unfortunately, triple-negative tumors are the most complicated in terms of tumoroids growth and the collection of a cohort of only human triple-negative tumors would be challenging in terms of time [29].

In conclusion, canine tumoroids respond well to a chemotherapeutic agent used in human medicine. The way the tumoroids are preserved has little impact on drug response. It seems, however, that tumoroids made from frozen cells best mimic the drug response of fresh tumoroids.

Discussion

Tumoroids provide an alternative to pre-clinical animal experiments and can help predict tumor response to therapy and screen new drugs. Until now, breast cancer tumoroids have been derived mainly from murine

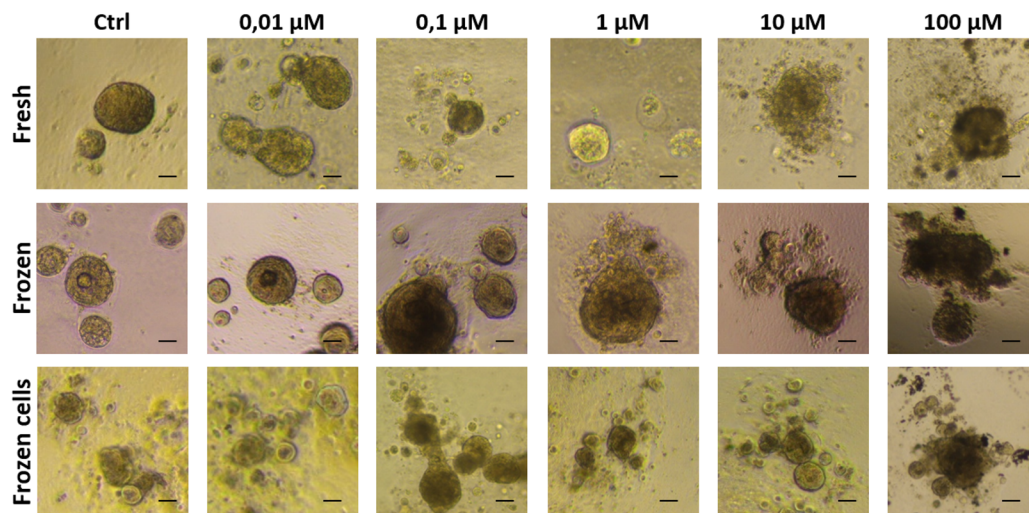
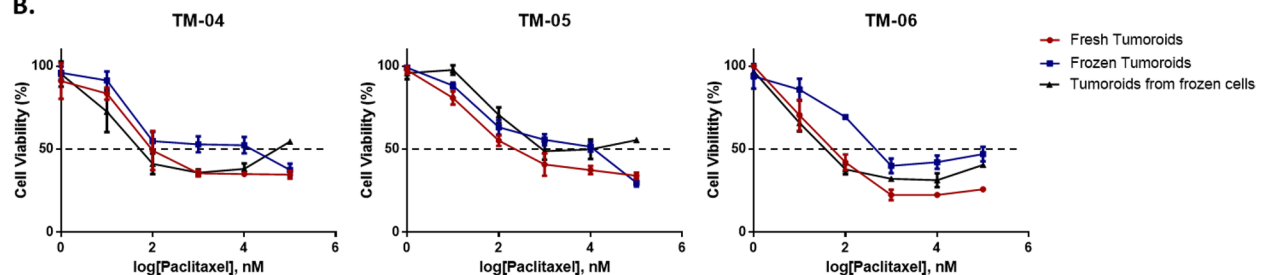
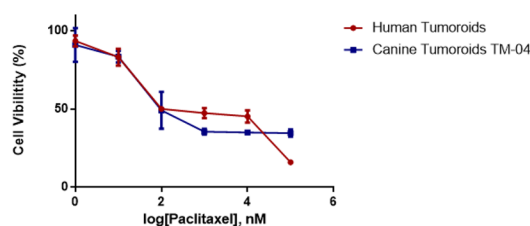
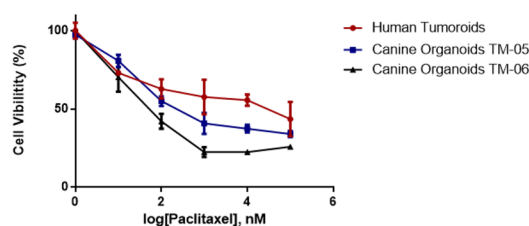
A. TM-05**B.****C. Luminal Breast cancer Tumoroids****Triple-negative Breast cancer Tumoroids**

Fig. 8 Drug response of canine and human tumoroids to paclitaxel. **A** Representative bright field images showing the morphology of the three types of tumoroids after 7 days treatment with paclitaxel at different concentrations. Scale bar (100 μ m) is indicated. **B** Quantification of the tumoroids viability following paclitaxel treatment. Tumoroids were generated from three different canine mammary tumors (TM-04, TM-05, and TM-06), and drug response was compared between fresh, frozen, and frozen cell tumoroids. Data are the means \pm SD ($n=3$ for each condition). **C** Tumoroids were generated from human and canine luminal and triple-negative mammary tumors, and drug response was compared between human and canine tumoroids. Different concentrations of drug were used and compared to non-treated tumoroids. Data are the means \pm SD ($n=3$ for each condition)

and human tissues [6, 30]. However, murine tumors do not reliably reflect the human pathology, and the use of human tumors faces several challenges such as ethical issues and the difficulty to access sufficient amount of fresh tissues to culture tumoroids, thus limiting high-throughput screening. In the present study, we have established the culture of canine mammary tumoroids in

order to develop a biobank which could be used to provide a better comprehension of breast cancer pathogenesis and for large-scale drug screening and therapeutic development for both veterinary and human medicine. In fact, dogs develop naturally numerous tumors in the presence of a functioning immune system that have similar features to human cancers [10, 18, 31–33]. CMT are

the most commonly diagnosed cancer in female dogs (50% of all cancers), which is a significant advantage since a large cohort of dogs could be recruited for preclinical studies. Moreover, pet dogs are more and more included in the development path of new human cancer drugs, for evaluating the safety and activity of a novel drug or understanding the pharmacokinetic of a novel drug in the context of a naturally occurring cancer model [34].

Subtype classification of CMT has been investigated in a number of studies using IHC expression of PR, ER and HER2. Several distinct subtypes were identified including luminal A (14.3%), luminal B (9.4%), and triple negative (76.3%), while no HER2-overexpressing CMT were observed [11]. Of the 6 dog patients included in the study, 4 tumors were of triple-negative subtype, while 2 tumors were of luminal subtype, representing 67% of triple-negative tumors and 33% of luminal tumors, which is consistent with previous findings. In human, as in dogs, the triple-negative subtype is more aggressive leading to shorter survival rates compared to other tumors [18]. Since therapeutic options for this subtype are limited, developing a reliable model to discover new effective treatments is highly needed.

We successfully generated tumoroids from CMT with a success rate of more than 94%. For comparison, in a recent study, human tumoroid establishment efficiency was around 40% for triple-negative subtype [29]. This difference can be explained by a higher amount of tissue which can be obtained from canine tumors, leading to high success rates. These tumoroids keep similar histological features as the original tumors as well as the same molecular subtype. Moreover, by a global proteomic analysis, we have shown that tumoroids were highly similar to the original tumor; 74% of proteins were identified in common between tumoroids and tumor. The tumor-specific signature is of course due to a higher cellular diversity in the primary tumor compared to tumoroids, as demonstrated by the over-expression of immune related proteins such as proteins of the complement pathway. On the contrary, an enriched metabolic signature is noticed in tumoroids compared to the primary tumor, which can be explained by the stress of the culture triggering a higher cellular activity. Nevertheless, a high degree of similarity is kept between tumor and tumor-derived tumoroids. Interestingly, a recent study found that the main pathways that were enriched in breast cancer were linked to cell communication, cell growth, and maintenance and signal transduction, which correlate well with our findings and is an additional proof that CMT are highly similar to human breast cancer [35]. In the same perspective, a recent study has performed a genetic assessment of canine mammary tumoroids in comparison to human tumoroids [36]. They have shown

that canine mammary tumoroids may offer a valuable model for specific human breast cancer types such as PI3KCA mutated tumors since these mutations were also found in canine mammary tumors.

Recently many tumoroid biobanks have emerged from different cancer types [37]. Many of these studies have demonstrated that tumoroids preserve the genetic composition of the original tumor. However, the extent of molecular drift at later passage and after cryopreservation has been relatively low studied so far. In the presented study, we have compared histologic and molecular features (marker-based subtype and global proteome) as well as therapeutic response of tumoroids maintained in culture without cryopreservation (fresh tumoroids), put in culture after cryopreservation (frozen tumoroids), or developed from frozen cells (issued from the initial tumor, frozen cell tumoroids). We found that from a morphological point of view, the three types of tumoroids were similar and kept the same architecture and growth rates. The CMT subtype was also maintained after cryopreservation. We have also found that the type of culture or the number of passages did not impact too much the proteome of tumoroids. Indeed, the main variations were observed between tumoroids derived from different tumors rather than between different culture conditions. However, with this global proteomic analysis, we still found that fresh tumoroids and tumoroids made from frozen cells were more similar with a higher proteome diversity compared to frozen tumoroids. A previous study showed that tumor heterogeneity and cell diversity was conserved between fresh tumor tissue and cryopreserved tissue fragments or from cryopreserved cell suspensions [38]. In the same study, the authors found that cryopreserved cell suspensions displayed higher correlations to fresh cells compared to tissue fragments. This can therefore explain our observations. Moreover, maintenance of stromal cell populations in tumoroids system is really challenging. At this time, the tumoroids culture system promote the expansion of the tumor cells but do not support the maintenance of immune cells and stromal cells [39]. Stromal cells and immune cells are maintained during the first passages and tend to decrease overtime. By using an air-liquid interface to reconstitute the tumor microenvironment, tumoroids integrating immune components were successfully generated but immune cells tend to decline over time [40]. Our proteomics results tend to demonstrate this fact, when tumoroids are kept fresh or are made from cells frozen after tumor dissociation, many proteins involved in metabolism, cell communication, and immune response were identified. This immune signature was even much more pronounced for fresh tumoroids as demonstrated by the expression of T cell and macrophage markers (Granzyme B, Siglec1, and

CD163). Previous studies have shown that CMT showed inflammatory infiltrate consisting predominantly of lymphocytes, macrophages, plasma cells, and mast cells supporting our proteomics findings [41]. These results suggest that the cellular diversity may be higher in fresh tumoroids and tumoroids made from frozen cells compared to cryopreserved tumoroids. Metabolic and stress signatures were enriched in frozen tumoroids, which can be explained by cryopreservation [38].

To finish demonstrating that the culture conditions do not impact too much the tumoroids behavior, we have performed a drug response of tumoroids with a known chemotherapy used in human medicine. Paclitaxel response was similar between tumoroids, whatever the condition (fresh or cryopreserved). We however observed that cryopreserved tumoroids were slightly more resistant to paclitaxel, reflected by a higher concentration of drug needed to kill 50% of cancer cells. These results corroborate our previous observations. Nevertheless, CMT tumoroids are sensitive to a human chemotherapy in a dose dependent manner with a similar response as human breast tumoroids.

Conclusions

In conclusion, for the first time, dog mammary tumoroids were produced from heterogeneous tumors. The tumoroids recapitulated the tumor histologic and molecular heterogeneities. Cryopreservation, which is often used for bio banking, did not seem to affect the molecular features and drug response of tumoroids. Nevertheless, we showed that cryopreservation of tumor cells after dissociation seem to best mimic the fresh tumoroids. Canine tumoroids can be used to screen human drugs without limitations about tissue availability allowing large-scale production. However, to make tumoroids even closer to the primary tumor, it is necessary to develop tumoroid models including stromal components such as immune cells which are lost during traditional tumoroids culture.

Methods

Human and dog patients' tissue collection

This study was carried out with canine mammary tumors ($n=6$), 4 triple-negative tumors and 2 luminal ones to be representative of the subtypes the more represented in dogs (luminal and triple-negative tumors represent 24% and 76% respectively of all canine mammary tumors) [11]. The tumors were collected at different veterinary clinics from dogs undergoing scheduled surgery. The samples were delivered with the written consent of the owners. The dogs included in the study were treated surgically by their veterinarian, and none of them received any additional treatment before the mastectomy. A veterinary pathologist reviewed the tissue blocks to confirm

the diagnosis and define the lesions for dissection. For this study, we received a piece of fresh tumor of approximately 1 cm³.

Human breast tumor tissue was obtained from a patient undergoing surgery for early breast cancer. Fresh tumor tissue was provided by the pathologist for organoid culture. The sample was anonymized prior to its transfer to the lab. The study was approved by the local research committee of Oscar Lambret Cancer center and a French Ethical Committee (study IdRCB 2021-A00670-41). The written informed consent for the study was obtained from the patient before any procedure.

Tissue processing

Each tumor sample was divided into three pieces: one piece was snap frozen in liquid nitrogen before being stored at -80°C for proteomics large-scale study, the second piece was fixed in paraformaldehyde 4% for 24 h followed by dehydration in 20% sucrose for 24 h, embedding in gelatin and storage at -80°C for histopathological analysis and hematoxylin and eosin staining. The last tumor fragment was used for tumoroids culture. For this, the tumor fragment was minced into 1 mm³ pieces before its enzymatic digestion as described below.

Tumoroid culture

The minced tumor tissue was digested in 2 mL Hank's Balanced Salt Solution (HBSS, Gibco) with antibiotics and anti-fungal (1X Penicillin/Streptomycin, 1X Amphotericin) containing 1 mg/mL collagenase type IV (Sigma) and 5 U/mL hyaluronidase (Sigma) at 37°C for 2 h. During this time, the medium containing the tumor tissue was mixed every 15 min to help digestion. After 2 h, 10 mL of HBSS with antibiotics was added, and the cell suspension was strained over a 100- μm filter (Dutcher) which retained remaining tissue pieces. The suspension was centrifuged at 300 g for 5 min. In case of a visible red pellet, erythrocytes were lysed in 1 mL red blood cell lysis buffer (RBC, Invitrogen) for 5 min at room temperature. Then, the suspension was completed with 10 mL HBSS with antibiotics and centrifuged at 300 g for 5 min. The viable cell suspension was counted and 150,000 cells were used for the generation of tumoroids. The cells were resuspended in a reduced growth factor solubilized basement membrane matrix for Organoid Culture (Matrigel®, Corning) and plated as a drop in 24-well plates. The Matrigel was allowed to solidify for 30 min in the incubator, and then 500 μL of complete culture medium was added. The culture medium was composed of Advanced DMEM (Gibco) supplemented with 1X Glutamax, 10 mM Hepes, 1X Penicillin/Streptomycin, 1X Amphotericin, 50 $\mu\text{g/mL}$ Primocin, 1X B27 supplement, 5 mM Nicotinamide, 1.25 mM N-Acetylcystein,

250 ng/mL R-spondin 1, 5 nM Heregulin β -1, 100 ng/mL Noggin, 20 ng/mL FGF-10, 5 ng/mL FGF-7, 5 ng/mL EGF, 500 nM A83-01, 500 nM SB202190, and 5 μ M Y-27632.

Tumoroids were split when tumoroids are confluent, i.e., when they occupy approximately 70–80% of the Matrigel drop. Ice cold PBS was used to harvest tumoroids from the Matrigel. They were collected in a 15-mL falcon that was pre-coated with PBS containing 1% BSA solution to prevent the tumoroids from adhering to the tube. The tumoroids were centrifuged at 300 g for 5 min and then digested with TrypLE solution (Gibco) for 5 min at 37° C. After enzymatic neutralization and washing, the tumoroid fragments were resuspended in Matrigel and reseeded as explained above to allow formation of new tumoroids.

Furthermore, after the initial digestion of the tumor tissue, 2 million cells were cryopreserved for the subsequent development of tumoroids. To create the tumoroids from the frozen cells, the vial was thawed slowly and the cells were centrifuged in 10 mL of HBSS with antibiotics at 300 g for 5 min. Then, the cells were counted and seeded in the same way as with fresh cells.

Freezing and thawing of tumoroids

Once the tumoroids were confluent, they were collected as described before, centrifuged at 300 g for 5 min, separated with a syringe mounted with a 21 G needle before being centrifuged again and frozen in 90% fetal bovine serum (FBS) and 10% DMSO.

Cryopreserved tumoroids were thawed slowly and 1 mL of the thawing solution was added to the vial (Advanced DMEM (Gibco), 15 mM Hepes (Gibco), 1% BSA (Sigma)). Then, the solution was transferred to a tube containing 2 mL of thawing solution. The tumoroids were centrifuged at 300 g for 5 min, and the pellet of tumoroids was resuspended with 30 μ L of Matrigel and cultured as explained before.

HE, immunohistochemistry, and immunofluorescence staining

The tumoroids were fixed in 2% paraformaldehyde with 0.1% glutaraldehyde for 24 h followed by dehydration in 20% sucrose for 24 h, embedding in gelatin and freezing at – 80 °C.

Standard H&E staining was carried out on 5 μ m thick tumor and tumoroid sections to appreciate the cellular and tissue structure details, using Tissue-Tek Prisma® Automated Slide Stainer. Images were acquired on a Nikon Eclipse NI-U with the Nikon Elements BR 4.50.00 software.

The immunohistochemical staining was carried out on 5 μ m thick tumor and tumoroid sections using an

automated protocol developed for the Discovery XT automated slide staining system (Ventana Medical Systems, Inc.). Tumor and tumoroid sections were incubated for 40 min with the appropriate antibody before incubation with Discovery UltraMap anti-Rabbit (760–4315, Roche) or anti-mouse horseradish peroxidase (HRP) (760–4313, Roche) secondary antibodies and the Discovery ChromoMap DAB kit reagents (760–159, Roche). Counterstaining and post-counterstaining were performed using hematoxylin and bluing reagent (Ventana, Roche Diagnostics). The following commercially available antibodies were used for the characterization: estrogen receptor (ER)– α (SC-8005, Santa Cruz), progesterone receptor (PR) (790–4296, Roche), and human epidermal growth factor 2 (HER-2) (790–4493, Roche).

The immunofluorescence staining was carried out on 12 μ m thick tumor and tumoroid sections. Tumor and tumoroid sections were washed 3 times in PBS, pre-incubated in blocking buffer in 0.3% Triton, 5% Normal Donkey Serum (NDS), and 2% ovalbumin in PBS for 1 h at room temperature. Then, the samples were incubated overnight at 4 °C with proliferation marker Ki67 (790–4286, Roche). After 3 washes with PBS, samples were incubated 1 h at 37 °C with secondary donkey anti-rabbit antibody conjugated to Alexa Fluor 488 (1:200, Invitrogen, Carlsbad CA, USA) in blocking buffer. They were rinsed with PBS and the cell nuclei were counterstained with Hoechst 33,342 fluorescent dye (1/10000, Invitrogen, Carlsbad CA, USA) for 20 min at 4 °C. Finally, the tumor and tumoroid sections were mounted on the slide with Dako Fluorescent Mounting Medium (Agilent, Santa Clara CA, USA). Samples without the addition of primary antibody were used as negative control. The presented pictures are representative of independent triplicates.

Proliferation of tumoroids

To measure the efficiency of tumoroids formation, the same amount of tumoroids was dissociated with cold PBS. The pellet was then digested with TrypLE solution (Gibco) for 5 min at 37 °C. The tumoroids were then diluted in HBSS and then passed through a 100- μ m filter (Dutcher) to remove large tumoroids. Subsequently, the tumoroids were centrifuged at 300 g for 5 min and then suspended in 2% Matrigel/tumoroid culture medium (3–5000 tumoroids/mL). Proliferation was measured at 1, 3, 5, and 7 days using CellTiter-Glo 3D (Promega) according to the manufacturer's instructions. A measurement was made at the beginning of the experiment which was considered as 100% for all the tumoroids and then the results are expressed as the percentage of growth compared to the T0 measurement. For each time of measurement, 100 μ L of CellTiter-Glo3D reagent (Promega, Madison, WI,

USA) was added to each well, and the plate was shaken at room temperature for 25 min. Luminescence was read on a TriStar2 S LB 942 Multimode Microplate Reader, and the data were analyzed using GraphPad Prism 6.

Total protein extraction

Sections of fresh frozen tumor and corresponding tumoroids were collected in triplicate for each condition. The tumor sections and the tumoroids pellet were lysed with RIPA buffer (150 mM NaCl, 50 mM Tris, 5 mM EGTA, 2 mM EDTA, 100 mM NaF, 10 mM sodium pyrophosphate, 1% NP40, 1 mM PMSF, and 1X protease inhibitors) for total protein extraction. Three steps of 30 s sonication at amplitude 50% on ice was applied, cell debris were removed by centrifugation ($16,000 \times g$, 10 min, 4 °C), the supernatants were collected, and protein concentrations were measured using a Bio-Rad Protein Assay Kit, according to the manufacturer's instructions. To normalize the tumoroids and tumor protein quantities, 100 µg of each sample was used for protein digestion and subsequent shotgun proteomics analysis.

Shotgun proteomics

Protein digestion was performed using the FASP method [42]. Briefly, reduction solution was added to the sample (100 mM DTT in 8 M urea in 0.1 M Tris/HCl, pH 8.5 (UA buffer)) and incubated for 15 min at 95 °C. The protein solution was then loaded onto 10 kDa Amicon filters, supplemented with 200 µL of UA buffer and centrifuged for 30 min at 14,000 g. Next, 200 µL of UA buffer were loaded onto the filter and centrifuged for 30 min at 14,000 g. Then, 100 µL of alkylation solution (0.05 M iodoacetamide in UA buffer) was added and incubated for 20 min in the dark before centrifugation for 30 min at 14,000 g. Finally, a 50-mM ammonium bicarbonate solution (AB) was added and centrifuged again for 30 min at 14,000 g. This last step was repeated three times. For the digestion, 50 µL LysC/Trypsin at 20 µg/mL in AB buffer was added and incubated at 37 °C overnight. The digested peptides were recovered after centrifugation for 30 min at 14,000 g. Then, two washes with 100 µL of AB buffer were performed by centrifugation for 30 min at 14,000 g. Finally, the eluted peptides were acidified with 10 µL of 0.1% trifluoroacetic acid (TFA) and dried under vacuum.

LC-MS/MS analysis

The samples once dried were reconstituted in 20 µL of a 0.1% TFA solution and desalted using a C18 ZipTip (Millipore, Saint-Quentin-en-Yvelines, France). After elution with 20 µL of 80% acetonitrile (ACN)/0.1% TFA, the peptides were vacuum dried. Samples were then

reconstituted in 0.1% formic acid/ACN (98:2, v/v) and separated by reverse phase liquid chromatography by an Easy-nLC 1000 nano-UPLC (Thermo Scientific) in the reverse phase using a preconcentration column (75 µm ID \times 2 cm, 3 µm, Thermo Scientific) and an analytical column (Acclaim PepMap C18, 75 µm ID \times 50 cm, 2 µm, Thermo Scientific) interfaced with a nanoelectrospray ion source on an Q-Exactive Orbitrap mass spectrometer (Thermo Scientific). Separation was performed using a linear gradient starting at 95% solvent A (0.1% FA in water) and 5% solvent B (0.1% FA in ACN) up to 70% solvent A and 30% solvent B for 120 min at 300 nL/min. The LC system was coupled onto a Thermo Scientific Q-Exactive mass spectrometer set to Top10 most intense precursors in data-dependent acquisition mode, with a voltage of 2.8 kV. The survey scans were set to a resolving power of 70,000 at FWHM (m/z 400), in positive mode and using a target AGC of $3E+6$. For the shotgun proteomics, the instrument was set to perform MS/MS between +2 and +8 charge state.

Data analyses

All the MS data were processed with MaxQuant (version 1.5.6.5) software [43] using the Andromeda search engine [44]. Proteins were identified by searching MS and MS/MS data against a database of *Canis lupus familiaris* obtain from Uniprot database and containing 45401 sequences. For identification, the FDR at the peptide spectrum matches (PSMs) and protein level was set to 1%. Label-free quantification of proteins was performed using the MaxLFQ algorithm with the default parameters. Analysis of the proteins identified were performed using Perseus (version 1.5.6.0) software [45, 46]. Multiple-sample tests were performed using ANOVA test with a p -value of 5% and preserving grouping in randomization. Visual heatmap representations of significant proteins variation were obtained using hierarchical clustering analysis. Functional annotation and characterization of identified proteins were obtained using PANTHER (version 13.0) software [47] and STRING (version 9.1) [48]. The analysis of gene ontology, cellular components, and biological processes was performed with FunRich 3.0 analysis tool [49].

Tumoroid response to paclitaxel

For tumoroid culture and drug response analysis, the same amount of tumoroids was dissociated with cold PBS. The pellet was then digested with TrypLE solution (Gibco) for 5 min at 37° C. The tumoroids were then diluted in HBSS and then passed through a 100-µm filter (Dutcher) to remove large tumoroids. Subsequently, the tumoroids were centrifuged at 300 g for 5 min and then suspended in 2% Matrigel/tumoroid culture medium

(3–5000 tumoroids/mL). For the drug response, 100 μ L of tumoroid solution was placed in wells of 96-well plates coated with 1.5% agarose. The tumoroids were allowed to form during 72 h and then treated with paclitaxel for 7 days before performing the viability test. Cell viability was performed using CellTiter-Glo 3D (Promega) according to the manufacturer's instructions, and results were normalized to controls. Paclitaxel concentrations ranged from 0.01 μ mol to 100 μ mol (5 concentrations), and DMSO controls were added. After 7 days, 100 μ L of CellTiter-Glo3D reagent (Promega, Madison, WI, USA) was added to each well, and the plate was shaken at room temperature for 25 min. Luminescence was read on a TriStar2 S LB 942 Multimode Microplate Reader and the data were analyzed using GraphPad Prism 6.

Abbreviations

ER	Estrogen receptor
PR	Progesterone receptor
HER2	Human epidermal growth factor receptor-2
3D	Three-dimensional
CMT	Canine mammary tumor
TM-01, 02, 03, 04, 05 and 06	Tumor 1, 2, 3, 4, 5, and 6, respectively
Ki67	Marker of proliferation Ki-67
PFA	Paraformaldehyde
D1 and D2	Date1 and date 2 of tumoroids passage
PCA	Principal component analysis
LFQ	Label-free quantification
GO	Gene Ontology
IC50	Concentration of drug required for 50% inhibition
IHC	Immunohistochemistry
HBSS	Hank's Balanced Salt Solution
BSA	Bovine serum albumin
DMSO	Dimethyl sulfoxide
TFA	Trifluoroacetic acid
ACN	Acetonitrile
PBS	Phosphate-buffered saline

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12915-023-01516-2>.

Additional file 1: Fig. S1. Histology and receptor status (ER, PR, HER2) of the 6 canine mammary tumors used in the study.

Additional file 2: Table 1. Summary table of canine tumors used in the study.

Additional file 3: Fig. S2. Immunofluorescence images of Ki67 stained canine mammary tumors.

Additional file 4: Table 2. Data generated that support the findings of this study: (I) summary table of growth efficiency of canine tumoroids used in the study showed in Figure 2B, (II) summary table of the percentage of Ki67+ cells showed in Figure 4A, (III) summary table of viability quantification of tumoroids showed in Figure 8B, (IV) summary table of viability quantification of tumoroids showed in Figure 8C and (V) availability of MS data.

Additional file 5: Fig. S3. Histology and receptor status (ER, PR, HER2) of breast cancer tumoroids. Comparative histological and immunohistochemical images of breast cancer tumoroids and their original breast cancer tissues.

Additional file 6: Table 3. List of proteins in the Venn diagram representing proteins identified in Fresh, Frozen, and FrozenCell tumoroids at date 1 and date 2.

Additional file 7: Table 4. List of proteins in the Venn diagram representing proteins in the Fresh, Frozen and FrozenCell tumoroids.

Additional file 8: Table 5. List of proteins in the 6 clusters identified in the Hierarchical clustering of the most variable proteins between the Fresh, Frozen and FrozenCell conditions.

Additional file 9: Table 6. List of proteins in the Venn diagram representing proteins in the Tumor, Fresh, Frozen and FrozenCell tumoroids conditions.

Additional file 10: Table 7. List of proteins in the 2 clusters identified in the Hierarchical clustering of the most variable proteins between the Tumor, Fresh, Frozen and FrozenCell tumoroids conditions.

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Authors' contributions

ARR, MD, and MS wrote the manuscript. MD and MS were responsible for the concept and design of the overall study and interpretation of the data. ARR and MD were involved in the experimental design, acquisition of the data, and analysis and interpretation of the data. SA and EB provided technical assistance and guidance. NH, AR, and DT provided the samples. MD, MS, and IF supervised the project and provided critical revision of manuscript. MD, MS, IF, and DT have obtained funding. All authors read and approved the final manuscript.

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Availability of data and materials

All data generated that support the findings of this study are included in the additional table 2 of this article. In addition, the mass spectrometry proteomics datasets generated and analyzed during the current study have been deposited to the ProteomeXchange Consortium via the PRIDE partner repository and are available with dataset identifier PXD031440.

Declarations

Ethics approval and consent to participate

All animal and human studies were reviewed and approved by the local ethics committees, as detailed in the Materials and Methods section.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Université Lille, Inserm, CHU Lille, U1192, Laboratoire Protéomique, Réponse Inflammatoire Et Spectrométrie de Masse (PRISM), Lille, France. ²OCR (Oncovet Clinical Research), Parc Eurasanté Lille Métropole, 80 Rue du Dr Yersin, 59120 Loos, France. ³Breast Cancer Unit, Oscar Lambret Center, Lille, France. ⁴Institut Universitaire de France, Paris, France.

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