



**HAL**  
open science

## Effective anti-BCMA retreatment in multiple myeloma.

Nicolas Gazeau, David Beauvais, Ibrahim Yakoub-Agha, Suman Mitra,  
Timothy B. Campbell, Thierry Facon, Salomon Manier

► **To cite this version:**

Nicolas Gazeau, David Beauvais, Ibrahim Yakoub-Agha, Suman Mitra, Timothy B. Campbell, et al..  
Effective anti-BCMA retreatment in multiple myeloma.. Blood Advances, 2021, Blood Advances, 5  
(15), pp.3016-3020. 10.1182/bloodadvances.2021004176 . hal-04423912

**HAL Id: hal-04423912**

**<https://hal.univ-lille.fr/hal-04423912v1>**

Submitted on 7 Sep 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

## Effective anti-BCMA retreatment in multiple myeloma

Nicolas Gazeau,<sup>1</sup> David Beauvais,<sup>1</sup> Ibrahim Yakoub-Agha,<sup>1,2</sup> Suman Mitra,<sup>3</sup> Timothy B. Campbell,<sup>4</sup> Thierry Facon,<sup>1</sup> and Salomon Manier<sup>1,3\*</sup>

<sup>1</sup>Department of Hematology, Centre Hospitalo-Universitaire (CHU) Lille, <sup>2</sup>INSERM U1286, Infinite, <sup>3</sup>Canther, INSERM UMR-S1277 and CNRS UMR9020, Lille University, Lille, France; and <sup>4</sup>Bristol Myers Squibb, Princeton, NJ

### Key Points

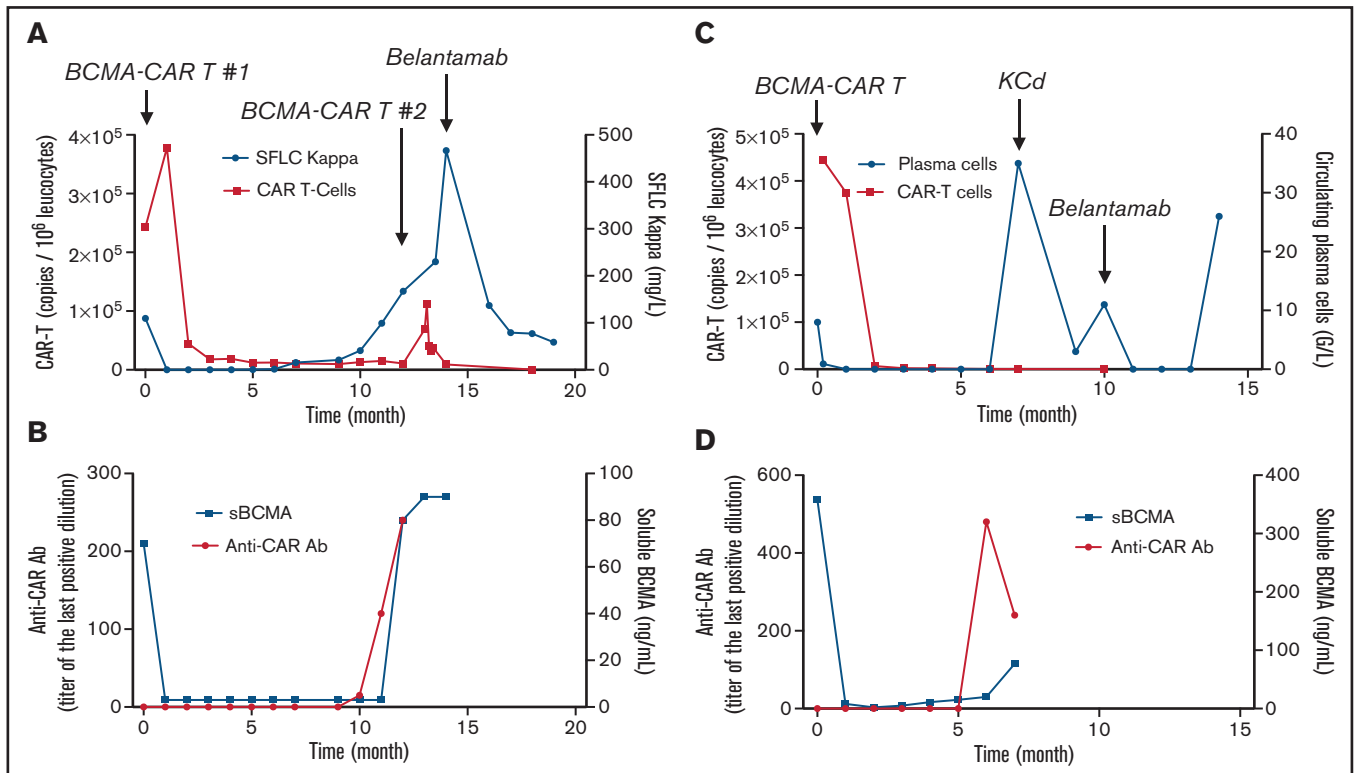
- We report clinical evidence of effectiveness using anti-BCMA therapies sequentially.
- Careful monitoring of the mechanism of relapse can help to guide clinical decisions about anti-BCMA retreatment.

The recent emergence of anti-B-cell maturation antigen (BCMA) therapies holds great promise in multiple myeloma (MM). These include chimeric antigen receptor (CAR) T cells, bispecific antibodies, and antibody-drug conjugates. Their development in clinical trials and further approval are changing the strategy for treating MM. Considering that a cure has not been reached, a central question in the coming years will be the possibility of using these therapies sequentially. Here, we report 2 cases of the serial use of anti-BCMA therapies with parallel monitoring of BCMA expression and anti-CAR antibodies. We further discuss recent data from clinical studies that have informed us about the different mechanisms of resistance to anti-BCMA therapies, including antigen escape, BCMA shedding, anti-drug antibodies, T-cell exhaustion, and the emergence of an immunosuppressive microenvironment. This knowledge will be essential to help guide the strategy of serial treatments with anti-BCMA therapies.

### Introduction

The B-cell maturation antigen (BCMA) has emerged as a central target in multiple myeloma (MM). BCMA, also designated as TNFRSF17, belongs to the tumor necrosis factor receptor superfamily, which is a family of cytokine receptors. BCMA's main ligands are the cytokines B-cell activating factor and a proliferation-inducing ligand.<sup>1</sup> The interaction between BCMA and its ligands activates the NF- $\kappa$ B signaling pathway that plays an important role in B-cell proliferation and maturation<sup>2</sup> and is essential for the survival of long-lived bone marrow plasma cells.<sup>3,4</sup> BCMA is expressed preferentially on mature B cells and has minimal expression on hematopoietic stem cells or other cell types.

In recent years, 3 BCMA-targeting therapies have emerged: bispecific antibodies (BsAbs; (AMG420,<sup>5</sup> CC93269,<sup>6</sup> and teclistamab<sup>7</sup>), an antibody-drug conjugate (ADC; belantamab mafodotin<sup>8</sup>), and chimeric antigen receptor (CAR) T cells (primarily bb2121,<sup>9</sup> bb21217,<sup>10</sup> JNJ4528<sup>11</sup> and orvacabtagene autoleucel<sup>12</sup>). These new anti-BCMA immunotherapies have demonstrated a high efficacy in the context of relapsed/refractory MM. BCMA-targeting CAR T cells are being evaluated in phase 3 trials in MM. The reported phase 1-2 data from the KarMMa,<sup>13</sup> CARTITUDE-1,<sup>11</sup> EVOLVE,<sup>12</sup> and BB21217<sup>10</sup> studies reveal overall response rates ranging from 73% to 100% in heavily pretreated patients who received a median of 5 to 6 prior lines of treatment. Of note, the complete remission (CR) rates range from 33% to 86%, with rates of negative minimal residual disease (at  $10^{-5}$ ) ranging from 28% to 50%. However, relapses are systematically observed after BCMA-targeting CAR T-cell therapy, even in patients who achieved minimal residual disease negativity, with a median progression-free survival (PFS) of 12.1 months at the target dose of 450 million CAR T cells in the KarMMa<sup>14</sup> trial and with a PFS rate of 86% at 9 months in the CARTITUDE-1 study.<sup>11</sup> Similar results are seen in dose-escalation studies of BsAbs targeting BCMA and CD3, with ORRs of 67% to 89% and CR rates of 38% to 50% at effective doses.<sup>5-7</sup> Response rates are lower with belantamab mafodotin, an anti-BCMA antibody (Ab) conjugated



**Figure 1. Treatment course of patients receiving serial BCMA-targeted therapies.** CAR T-cell expression and serum free light chain (SFLC) expression (A) and sBCMA and anti-CAR antibodies (B) in patient #1 receiving 2 infusions of BCMA-targeted CAR T cells and belantamab. CAR T-cell expression and circulating plasma cells (C) and sBCMA and anti-CAR antibodies (D) in patient #2 receiving 1 infusion of BCMA-targeted CAR T cells followed by belantamab.

to monomethyl auristatin F, with an ORR of 32% and a PFS of 2.8 months (with 2.5 mg/kg) in the DREAMM-2 phase 2 trial.<sup>8</sup> Despite the remarkable efficacy of anti-BCMA immunotherapies, relapses occur systematically, even in patients who achieved deep responses. This observation raises 2 essential questions: What are the mechanisms of resistance to anti-BCMA therapies? and Can we use BCMA-targeting therapies sequentially?

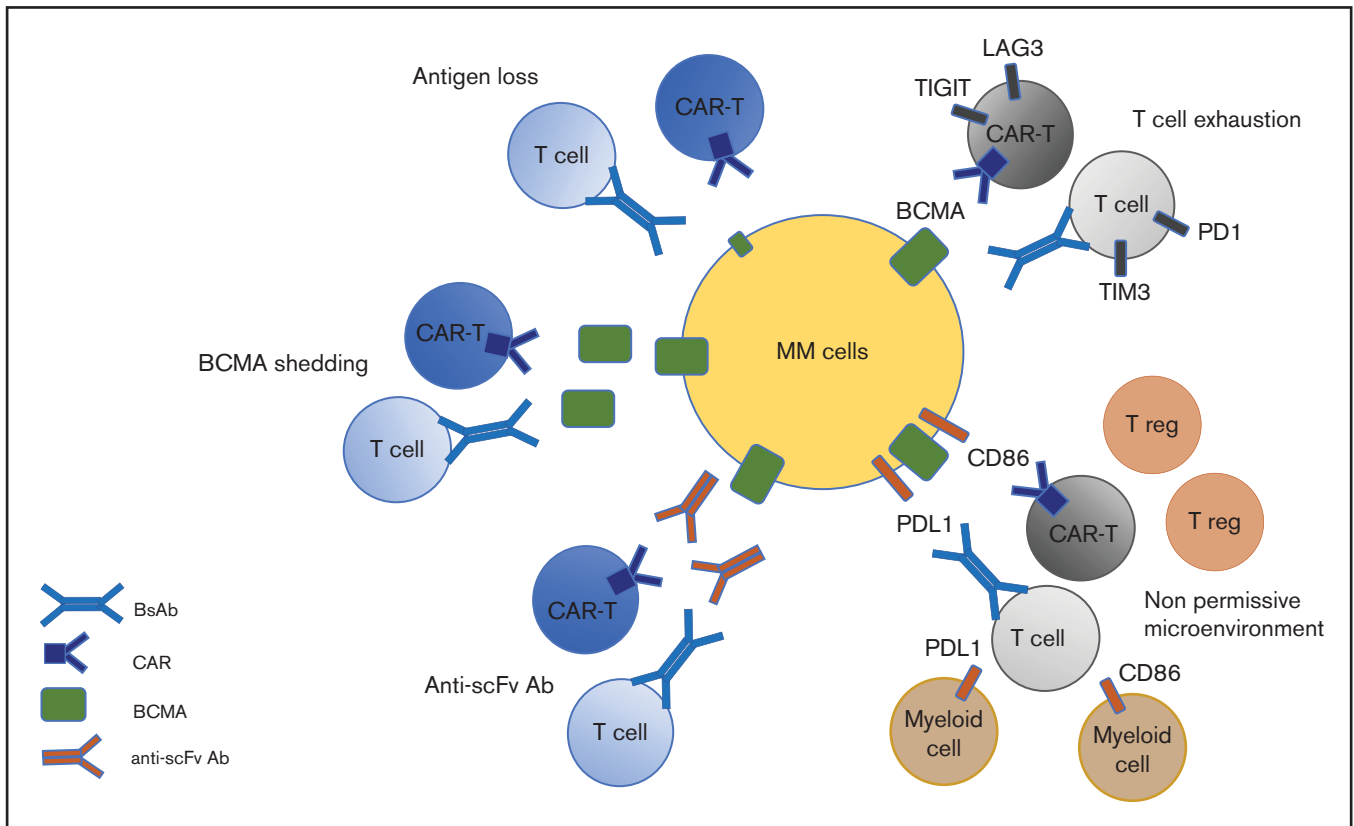
Here, we describe 2 clinical cases of patients receiving sequential anti-BCMA therapies, illustrating the feasibility of this approach. We also discuss the biological and clinical evidence highlighting the mechanism of resistance to anti-BCMA therapies.

## Case descriptions

The first case is a 74-year-old man with  $\kappa$  light chain MM, with unknown cytogenetic status, who was treated on a phase 2 trial of BCMA-directed CAR T cells (bb2121, KarMMA<sup>13</sup>). At enrollment, the patient was refractory to bortezomib, lenalidomide, pomalidomide, and daratumumab, with 4 prior lines of treatment including 2 autograft transplantations. He received lymphodepleting therapy (fludarabine and cyclophosphamide) and was infused with  $450 \times 10^6$  CAR+ T cells. He achieved a stringent CR at day 30 (Figure 1A) and remained disease free for 1 year before progressing with increased  $\kappa$  light chains, Bence Jones proteinuria, and new bone lesions. CAR T-cell peak expansion exceeded the median observed across treated patients after infusion and was detectable out to 9 months postinfusion. However, CAR T cells dropped below the level of detection in the blood at the time of relapse. The expression of

soluble BCMA (sBCMA) declined in parallel with the initial response, but then increased at relapse, suggesting the persistence of BCMA expression by tumor cells. Persistent BCMA expression in >50% of bone marrow plasma cells was confirmed at progression by immunohistochemistry. As allowed per protocol, the patient received a second infusion of  $450 \times 10^6$  CAR T cells. The monitoring of CAR T cells demonstrated an  $\sim 26$ -fold lower expansion in response to the second infusion relative to the first infusion (Figure 1A). The patient's MM did not respond, and the disease continued to progress. A retrospective analysis revealed the presence of anti-CAR antibodies at the time of relapse after the first infusion, potentially contributing to the lower expansion of CAR T cells after the second infusion (Figure 1B). He received subsequent treatment with belantamab mafodotin, a conjugated anti-BCMA Ab (GlaxoSmithKline; compassionate use). The patient achieved a very good partial response after 3 injections of belantamab mafodotin and is still in response 5 months later. In this clinical case, the patient received 3 sequential anti-BCMA therapies: 2 BCMA-targeting CAR T-cell infusions and 1 line of BCMA-targeted ADC. A potential mechanism of failure in response to the second infusion of CAR T cells was the presence of anti-CAR antibodies. The increase in sBCMA and persistent tumor BCMA expression at the time of CAR T-cell failure permitted the effective use of another anti-BCMA therapy with a different single chain variable fragment (scFv). This case illustrates that a systematic evaluation of the mechanism of resistance to anti-BCMA therapies can guide their sequential use in clinical practice.

The second case is a 68-year-old man with IgG  $\kappa$  MM, with t(14;16), deletion of 17p, and gain of 1q, who was treated on the



**Figure 2. Potential mechanism of resistance to anti-BCMA immunotherapies.** Three mechanisms are target dependent, including antigen loss, BCMA shedding by sBCMA, and the secretion of anti-drug antibodies. Two mechanisms of resistance are T-cell dependent, including T-cell exhaustion and the emergence of a nonpermissive microenvironment.

phase 2 KarMMA trial.<sup>13</sup> At enrollment, he had received 4 prior lines of treatment, including 1 autograft transplantation and was refractory to bortezomib, lenalidomide, pomalidomide, and daratumumab. He received bridging therapy with melphalan (50 mg/m<sup>2</sup>), lymphodepleting therapy (fludarabine and cyclophosphamide), and was infused with 450 × 10<sup>6</sup> CAR+ T cells. The patient achieved a stringent CR at day 30 and remained disease free for 10 months before progressing with a plasma cell leukemia (Figure 1C). Although bone marrow evaluation at relapse was nonevaluative for tumor BCMA expression, his sBCMA level was high at relapse, reflecting the expression of BCMA by the tumor cells. He also had detectable anti-CAR antibodies at the time of relapse (Figure 1D). He received a seventh line of treatment (carfilzomib, cyclophosphamide, and dexamethasone) with no response and was subsequently treated with belantamab mafodotin (GlaxoSmithKline; compassionate use). He had a rapid response with regard to the peripheral tumor, with a rapid clearance of circulating plasma cells together with a mild tumor lysis syndrome, but he had no response on the M-spike after 2 injections and progressed thereafter (Figure 1C).

These cases emphasize the possibility of using sequential anti-BCMA therapies in BCMA<sup>+</sup> relapses, which remain a functional target even after prior therapeutic pressure on this axis. A switch in the antigen binding fragment might be necessary in the case of anti-scFv antibodies. Therefore, careful monitoring of these 2 parameters will be essential to guide treatment decisions in clinical practice in the near future.

## Methods

The clinical trial referenced in this article is the KarMMA study,<sup>13</sup> which is registered at [www.clinicaltrials.gov](http://www.clinicaltrials.gov) as #NCT03361748. It was approved by the institutional review board and scientific committee of Centre Hospitalo-Universitaire (CHU) Lille and conducted in accordance with the Declaration of Helsinki.

### CAR T-cell expansion/pharmacokinetics

DNA was purified from peripheral blood CD3<sup>+</sup> cells. Vector transgene copies were measured per micrograms of genomic DNA by quantitative polymerase chain reaction (qPCR), as previously described.<sup>13</sup> CAR T-cell levels were also assessed by an in-house vector-based quantitative polymerase chain reaction (qPCR) assay, as previously described.<sup>15</sup>

### sBCMA evaluation

Baseline and post-CAR T-cell infusion levels of sBCMA were evaluated in patient serum using a Luminex immunoassay (cat. no. LXSAHM-01; R&D Systems).

### Anti-drug antibodies

The potential immune response to idecabtagene vicleucel was evaluated for humoral responses. Serum samples collected postinfusion were evaluated for the formation of anti-drug antibodies using an

immunoassay that was designed and validated to detect antibodies to the extracellular CAR domain, as previously described.<sup>13</sup>

## Results and discussion

The recent development of multiple anti-BCMA immunotherapies has raised the question of using them sequentially. Emerging clinical experience supports the effectiveness of this approach. In addition to the 2 cases reported here, 5 cases were described previously in the literature.<sup>16,17</sup> Among them, 3 received murine-scFv anti-BCMA CAR T cells, followed by fully human BCMA CAR T-cell therapy; 2 patients achieved a CR, and 1 achieved a very good partial response.<sup>16</sup>

Recent data from clinical studies have informed us about the different mechanisms of resistance to anti-BCMA therapies. Some are antigen dependent, such as antigen escape, BCMA shedding, and anti-scFv antibodies. Others are T-cell dependent, such as T-cell exhaustion and the emergence of an immunosuppressive microenvironment (Figure 2). Antigen escape has been reported but it does not appear to be frequent in the context of anti-BCMA therapies, probably because it is essential for the survival of plasma cells.<sup>3</sup> In the KarMMA study, of 16 patients with an evaluable bone marrow sample at relapse, only 1 patient had an antigen loss.<sup>18</sup> Biallelic deletion on chromosome 16, encompassing the BCMA locus, has been described as a mechanism of BCMA loss.<sup>19,20</sup> BCMA can also undergo  $\gamma$ -secretase-mediated shedding from plasma cells, leading to the circulation of sBCMA.<sup>21</sup> Theoretically, high levels of sBCMA could interfere with anti-BCMA therapies by coating a BCMA-directed binding fragment (scFv or Ab) and, thereby, function as an antigen-masking mechanism.<sup>22</sup> However, no clinical evidence exists to suggest that sBCMA levels can negatively impact BCMA-targeting therapies, but additional clinical experience, including testing  $\gamma$ -secretase inhibitors, will help to elucidate the effect of sBCMA levels on the function of BCMA-targeting therapies. Until recently, most of the BCMA-targeting CAR T cells evaluated in the clinic have derived their scFv from nonhuman species (mouse for bb2121<sup>9</sup> and camelid for JNJ4528<sup>11</sup>). The use of nonhuman scFv can induce immunogenicity resulting from an adaptive immune response after CAR T-cell infusion, which may play a role in limiting the persistence of the CARs. A recent study of 17 patients with relapsed/refractory MM treated with the biepitopic BCMA-targeting CAR T-cell therapy ciltacabtagene autoleucel revealed that 7 of them had developed high levels of anti-CAR antibodies.<sup>23</sup> The incidence of relapse was significantly higher than that in patients without detectable humoral immunogenicity. Therefore, anti-CAR antibodies are associated with a high risk for relapse after CAR T-cell therapy. The presence of anti-scFv antibodies in patients relapsing following BCMA-targeting CAR T-cell therapy also emphasizes the strategy of developing fully human or humanized scFv and antibodies.

## References

1. Bossen C, Schneider P. BAFF, APRIL and their receptors: structure, function and signaling. *Semin Immunol*. 2006;18(5):263-275.
2. Moreaux J, Legouffe E, Jourdan E, et al. BAFF and APRIL protect myeloma cells from apoptosis induced by interleukin 6 deprivation and dexamethasone. *Blood*. 2004;103(8):3148-3157.
3. O'Connor BP, Raman VS, Erickson LD, et al. BCMA is essential for the survival of long-lived bone marrow plasma cells. *J Exp Med*. 2004;199(1):91-98.

T-cell exhaustion remains a potential major mechanism of relapse during BCMA-targeting therapies. In the context of CAR T cells or BsAbs, responses have been associated with a higher CD4/CD8 ratio and an increased frequency of CD45RO<sup>-</sup>CD27<sup>+</sup>CD8<sup>+</sup> T cells, reflective of stem memory T cells.<sup>24,25</sup> In contrast, T cells with exhausted or senescent phenotypes are enriched in patients who are resistant to anti-BCMA BsAb or CAR T cells.<sup>25</sup> This indicates that the efficacy of BCMA-targeting therapies may be greater during early stages of the disease when patients are less immunosuppressed. Moreover, little is known regarding the response of the tumor microenvironment to immunotherapies and its capacity to impact the response to a subsequent immunotherapy. Further studies with sequential sampling will help to determine the role of the microenvironment in anti-BCMA failure.

Knowledge about the mechanisms of resistance to anti-BCMA therapies will help to define how we can use them sequentially. BCMA<sup>+</sup> relapses represent an opportunity for sequential treatment using anti-BCMA therapies. The switch in antigen binding fragment prevents the inhibition of subsequent anti-BCMA therapies by anti-drug antibodies developed during a previous exposure. Another aspect, but less well explored, is the impact of T-cell failure, as well as an immunosuppressive microenvironment, and how it can affect retreatment with anti-BCMA CAR T-cell or BsAb therapies. Monitoring of exploratory biomarkers, such as sBCMA, tumor BCMA expression, and the emergence of anti-drug antibodies, will help to further define the major mechanisms of resistance and guide clinicians in making therapeutic decisions.

## Authorship

Contribution: All authors contributed to the outline, design, writing, and editing of the manuscript.

Conflict-of-interest disclosure: I.Y.-A. has received honorarium from Celgene/Bristol Myers Squibb, Janssen, Kite/Gilead, and Novartis. T.B.C. works for Bristol Myers Squibb corporation. T.F. is a member of the speaker's bureau for Celgene/Bristol Myers Squibb and Janssen. S. Manier has received research funding from AbbVie, Amgen, Celgene/Bristol Myers Squibb, GlaxoSmithKline, Janssen, and Takeda. The remaining authors declare no competing financial interests.

ORCID profiles: D.B., 0000-0003-1866-828X; I.Y.-A., 0000-0003-4524-8782.

Correspondence: Salomon Manier, Service d'Hématologie, Hôpital Huriez, CHU Lille, rue Michel Polonovski, 59000 Lille, France; e-mail: salomon.manier@chru-lille.fr.



4. Novak AJ, Darce JR, Arendt BK, et al. Expression of BCMA, TACI, and BAFF-R in multiple myeloma: a mechanism for growth and survival. *Blood*. 2004;103(2):689-694.
5. Topp MS, Duell J, Zugmaier G, et al. Anti-B-cell maturation antigen BiTE molecule AMG 420 induces responses in multiple myeloma. *J Clin Oncol*. 2020;38(8):775-783.
6. Costa LJ, Wong SW, Bermúdez A, et al. First clinical study of the B-cell maturation antigen (BCMA) 2 + 1 T cell engager (TCE) CC-93269 in patients (Pts) with relapsed/refractory multiple myeloma (RRMM): interim results of a phase 1 multicenter trial. *Blood*. 2019;134(suppl 1):143.
7. Usmani SZ, Mateos M-V, Nahi H, et al. Phase I study of teclistamab, a humanized B-cell maturation antigen (BCMA) x CD3 bispecific antibody, in relapsed/refractory multiple myeloma (R/R MM). *J Clin Oncol*. 2020;38(15 suppl):100.
8. Lonial S, Lee HC, Badros A, et al. Belantamab mafodotin for relapsed or refractory multiple myeloma (DREAMM-2): a two-arm, randomised, open-label, phase 2 study. *Lancet Oncol*. 2020;21(2):207-221.
9. Raje N, Berdeja J, Lin Y, et al. Anti-BCMA CAR T-cell therapy bb2121 in relapsed or refractory multiple myeloma. *N Engl J Med*. 2019;380(18):1726-1737.
10. Berdeja JG, Alsina M, Shah ND, et al. Updated results from an ongoing phase 1 clinical study of bb21217 anti-Bcma CAR T cell therapy. *Blood*. 2019;134(suppl 1):927.
11. Madduri D, Usmani SZ, Jagannath S, et al. Results from CARTITUDE-1: a phase 1b/2 study of JNJ-4528, a CAR-T cell therapy directed against B-cell maturation antigen (BCMA), in patients with relapsed and/or refractory multiple myeloma (R/R MM). *Blood*. 2019;134(suppl 1):577.
12. Mailankody S, Jakubowiak AJ, Htut M, et al. Orvacabtagene autoleucl (orva-cel), a B-cell maturation antigen (BCMA)-directed CAR T cell therapy for patients (pts) with relapsed/refractory multiple myeloma (RRMM): update of the phase 1/2 EVOLVE study (NCT03430011). *J Clin Oncol*. 2020;38(15 suppl):8504.
13. Munshi NC, Anderson LD Jr, Shah N, et al. Idecabtagene vicleucl in relapsed and refractory multiple myeloma. *N Engl J Med*. 2021;384(8):705-716.
14. Munshi NC, Larry D, Anderson J, et al. Idecabtagene vicleucl (ide-cel; bb2121), a BCMA-targeted CAR T-cell therapy, in patients with relapsed and refractory multiple myeloma (RRMM): initial KarMMa results. *J Clin Oncol*. 2020;38(15 suppl):8503.
15. Baras A, Bocket L, Beauvais D, et al. A real-time quantitative PCR targeting the viral vector for the monitoring of patients treated with axicabtagene ciloleucl. *J Mol Diagn*. 2021;23(4):447-454.
16. Li C, Zhou J, Wang J, et al. Clinical responses and pharmacokinetics of fully human BCMA targeting CAR T-cell therapy in relapsed/refractory multiple myeloma. *J Clin Oncol*. 2019;37(15 suppl):8013.
17. Cohen AD, Garfall AL, Dogan A, et al. Serial treatment of relapsed/refractory multiple myeloma with different BCMA-targeting therapies. *Blood Adv*. 2019;3(16):2487-2490.
18. Martin N, Thompson E, Dell'Arima J, et al. Correlation of tumor BCMA expression with response and acquired resistance to idecabtagene vicleucl in the KarMMa study in relapsed and refractory multiple myeloma. Paper presented at the 25th Congress of the European Hematology Association (EHA25 Virtual), 11-14 June 2020.
19. Da Vià MC, Dietrich O, Truger M, et al. Homozygous BCMA gene deletion in response to anti-BCMA CAR T cells in a patient with multiple myeloma. *Nat Med*. 2021;27(4):616-619.
20. Samur MK, Fulciniti M, Aktas Samur A, et al. Biallelic loss of BCMA as a resistance mechanism to CAR T cell therapy in a patient with multiple myeloma. *Nat Commun*. 2021;12(1):868.
21. Laurent SA, Hoffmann FS, Kuhn PH, et al.  $\gamma$ -Secretase directly sheds the survival receptor BCMA from plasma cells. *Nat Commun*. 2015;6(1):7333.
22. Chen H, Li M, Xu N, et al. Serum B-cell maturation antigen (BCMA) reduces binding of anti-BCMA antibody to multiple myeloma cells. *Leuk Res*. 2019;81:62-66.
23. Xu J, Chen LJ, Yang SS, et al. Exploratory trial of a bi-epitopic CAR T-targeting B cell maturation antigen in relapsed/refractory multiple myeloma. *Proc Natl Acad Sci USA*. 2019;116(19):9543-9551.
24. Cohen AD, Garfall AL, Stadtmauer EA, et al. B cell maturation antigen-specific CAR T cells are clinically active in multiple myeloma. *J Clin Invest*. 2019;129(6):2210-2221.
25. Leblay N, Maity R, Barakat E, et al. Cite-seq profiling of T cells in multiple myeloma patients undergoing BCMA targeting CAR-T or Bites immunotherapy. *Blood*. 2020;136(suppl 1):11-12.