



**HAL**  
open science

## A Simple Prognostic System in Myelofibrosis Patients Undergoing Allogeneic Stem Cell Transplant: A CIBMTR/EBMT analysis.

Roni Tamari, Donal P. McLornan, Kwang Woo Ahn, Noel Estrada-Merly, Juan Carlos Hernandez-Boluda, Sergio A. Giralt, Jeanne M. Palmer, Robert Peter Gale, Zachariah Defilipp, David I. Marks, et al.

### ► To cite this version:

Roni Tamari, Donal P. McLornan, Kwang Woo Ahn, Noel Estrada-Merly, Juan Carlos Hernandez-Boluda, et al.. A Simple Prognostic System in Myelofibrosis Patients Undergoing Allogeneic Stem Cell Transplant: A CIBMTR/EBMT analysis.. Blood Advances, 2023, Blood Advances, 10.1182/blood-advances.2023009886 . hal-04520162

**HAL Id: hal-04520162**

**<https://hal.univ-lille.fr/hal-04520162>**

Submitted on 25 Mar 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial - NoDerivatives 4.0 International License

# A simple prognostic system in patients with myelofibrosis undergoing allogeneic stem cell transplantation: a CIBMTR/EBMT analysis

Roni Tamari,<sup>1</sup> Donal P. McLornan,<sup>2</sup> Kwang Woo Ahn,<sup>3,4</sup> Noel Estrada-Merly,<sup>4</sup> Juan Carlos Hernández-Boluda,<sup>5</sup> Sergio Giral,<sup>6</sup> Jeanne Palmer,<sup>7</sup> Robert Peter Gale,<sup>8</sup> Zachariah DeFilipp,<sup>9</sup> David I. Marks,<sup>10</sup> Marjolein van der Poel,<sup>11</sup> Leo F. Verdonck,<sup>12</sup> Minoo Battiwalla,<sup>13</sup> Miguel Angel Diaz,<sup>14</sup> Vikas Gupta,<sup>15</sup> Haris Ali,<sup>16</sup> Mark Robert Litzow,<sup>17</sup> Hillard M. Lazarus,<sup>18</sup> Usama Gergis,<sup>19</sup> Asad Bashey,<sup>20</sup> Jane Liesveld,<sup>21</sup> Shahrukh Hashmi,<sup>22,23</sup> Jeffrey J. Pu,<sup>24</sup> Amer Beitinjaneh,<sup>25</sup> Christopher Bredeson,<sup>26</sup> David Rizzieri,<sup>27</sup> Bipin N. Savani,<sup>28</sup> Muhammad Bilal Abid,<sup>29</sup> Siddhartha Ganguly,<sup>30</sup> Vaibhav Agrawal,<sup>31</sup> Vera Ulrike Bacher,<sup>32</sup> Baldeep Wirk,<sup>33</sup> Tania Jain,<sup>34</sup> Corey Cutler,<sup>35</sup> Mahmoud Aljurf,<sup>36</sup> Tamila Kindwall-Keller,<sup>37</sup> Mohamed A. Kharfan-Dabaja,<sup>38</sup> Gerhard C. Hildebrandt,<sup>39</sup> Attaphol Pawarode,<sup>40</sup> Melhem M. Solh,<sup>41</sup> Jean A. Yared,<sup>42</sup> Michael R. Grunwald,<sup>43</sup> Sunita Nathan,<sup>44</sup> Taiga Nishihori,<sup>45</sup> Sachiko Seo,<sup>46</sup> Bart L. Scott,<sup>47</sup> Ryotaro Nakamura,<sup>48</sup> Betul Oran,<sup>49</sup> Tomasz Czerw,<sup>50</sup> Ibrahim Yakoub-Agha,<sup>51</sup> and Wael Saber<sup>4</sup>

<sup>1</sup>Department of Medicine, Memorial Sloan Kettering Cancer Center, New York, NY; <sup>2</sup>Department of Medicine, University College Hospital, London, United Kingdom; <sup>3</sup>Division of Biostatistics, Institute for Health and Equity and <sup>4</sup>Department of Medicine, Center for International Blood and Marrow Transplant Research, Medical College of Wisconsin, Milwaukee, WI; <sup>5</sup>Department of Hematology, Hospital Clínico Universitario, Valencia, Spain; <sup>6</sup>Department of Internal Medicine, Memorial Sloan Kettering Cancer Center, New York, NY; <sup>7</sup>Department of Medicine, Mayo Clinic Arizona and Phoenix Children's Hospital, Phoenix, AZ; <sup>8</sup>Department of Immunology and Inflammation, Haematology Centre, Imperial College London, London, United Kingdom; <sup>9</sup>Department of Medicine, Hematopoietic Cell Transplant and Cellular Therapy Program, Massachusetts General Hospital, Boston, MA; <sup>10</sup>Adult Bone Marrow Transplant, University Hospitals Bristol NHS Trust, Bristol, United Kingdom; <sup>11</sup>Division of Hematology, Department of Internal Medicine, GROW School for Oncology and Developmental Biology, Maastricht University Medical Center, Maastricht, The Netherlands; <sup>12</sup>Department of Hematology/Oncology, Isala Clinic, Zwolle, The Netherlands; <sup>13</sup>Outcomes Research, Sarah Cannon Blood Cancer Network, Nashville, TN; <sup>14</sup>Department of Hematology/Oncology, Hospital Infantil Universitario Niño Jesus, Madrid, Spain; <sup>15</sup>Department of Internal Medicine, Princess Margaret Cancer Centre, University of Toronto, Toronto, Canada; <sup>16</sup>Department of Hematology & Hematopoietic Cell Transplantation, City of Hope, Duarte, CA; <sup>17</sup>Division of Hematology and Transplant Center, Mayo Clinic, Rochester, MN; <sup>18</sup>Department of Hematology and Internal Medicine, University Hospitals Cleveland Medical Center, Case Western Reserve University, Cleveland, OH; <sup>19</sup>Division of Hematological Malignancies, Department of Medicine Oncology, Thomas Jefferson University, Philadelphia, PA; <sup>20</sup>Department of Medicine, Blood and Marrow Transplant Program at Northside Hospital, Atlanta, GA; <sup>21</sup>Department of Medicine, University of Rochester Medical Center, Rochester, NY; <sup>22</sup>Department of Internal Medicine, Mayo Clinic, Rochester, MN; <sup>23</sup>Department of Medicine, Sheikh Shakhboub Medical City, Abu Dhabi, United Arab Emirates; <sup>24</sup>Department of Medicine, Banner University Medical Center Tucson, Syracuse, NY; <sup>25</sup>Division of Transplantation and Cellular Therapy, University of Miami Hospital and Clinics, Sylvester Comprehensive Cancer Center, Miami, FL; <sup>26</sup>Department of Medicine, The Ottawa Hospital Transplant & Cellular Therapy Program, Ottawa, ON, Canada; <sup>27</sup>Novant Health Cancer Institute, Winston Salem, NC; <sup>28</sup>Division of Hematology/Oncology, Department of Medicine, Vanderbilt University Medical Center, Nashville, TN; <sup>29</sup>Divisions of Hematology/Oncology & Infectious Diseases, Department of Medicine, Medical College of Wisconsin, Milwaukee, WI; <sup>30</sup>Department of Medicine, Houston Methodist Hospital and Cancer Center, Houston, TX; <sup>31</sup>Division of Leukemia, Department of Hematology & Hematopoietic Cell Transplantation, City of Hope National Medical Center, Duarte, CA; <sup>32</sup>Department of Hematology, Inselspital, Bern University Hospital, University of Bern, Bern, Switzerland; <sup>33</sup>Department of Medicine, Bone Marrow Transplant Program, Penn State Cancer Institute, Hershey, PA; <sup>34</sup>Division of Hematological Malignancies and Bone Marrow Transplantation, Sidney Kimmel Comprehensive Cancer Center, Johns Hopkins University, Baltimore, MD; <sup>35</sup>Stem Cell Transplantation and Cellular Therapy, Dana-Farber Cancer Institute, Boston, MA; <sup>36</sup>Department of Oncology, King Faisal Specialist Hospital Center & Research, Riyadh, Saudi Arabia; <sup>37</sup>Division of Hematology/Oncology, University of Virginia Health System, Charlottesville, VA; <sup>38</sup>Division of Hematology-Oncology, Blood and Marrow Transplantation Program, Mayo Clinic, Jacksonville, FL; <sup>39</sup>Division of Hematology, Markey Cancer Center, University of Kentucky, Lexington, KY; <sup>40</sup>Division of Hematology/Oncology, Department of Internal Medicine, Blood and Marrow Transplantation Program, University of Michigan Medical School, Ann Arbor, MI; <sup>41</sup>The Blood and Marrow Transplant Program, Northside Hospital Cancer Institute, Atlanta, GA; <sup>42</sup>Division of Hematology/Oncology, Department of Medicine, Transplantation & Cellular Therapy Program, Greenebaum Comprehensive Cancer Center, University of Maryland, Baltimore, MD; <sup>43</sup>Department of Hematologic Oncology and Blood Disorders, Levine Cancer Institute, Atrium Health, Charlotte, NC; <sup>44</sup>Department of Internal Medicine, Section of Bone Marrow Transplant and Cell Therapy, Rush University Medical Center, Chicago, IL; <sup>45</sup>Department of Blood & Marrow Transplant and Cellular Immunotherapy, Moffitt Cancer Center, Tampa, FL; <sup>46</sup>Department of Hematology and Oncology, Dokkyo Medical University, Tochigi, Japan; <sup>47</sup>Clinical Research Division, Fred Hutchinson Cancer Research Center, Seattle, WA; <sup>48</sup>Department of Hematology & Hematopoietic Cell Transplantation, City of Hope, Duarte, CA; <sup>49</sup>Division of Cancer Medicine, Department of Stem Cell Transplantation, The University of Texas MD Anderson Cancer Center, Houston, TX; <sup>50</sup>Department of Haematology and BMT, Maria Skłodowska-Curie National Research Institute of Oncology, Gliwice, Poland; and <sup>51</sup>Department of Hematology, CHRU Hôpital Claude Huriez, Lille, France

Submitted 3 February 2023; accepted 17 April 2023; prepublished online on *Blood Advances* First Edition 3 May 2023; final version published online 28 July 2023. <https://doi.org/10.1182/bloodadvances.2023009886>.

CIBMTR supports accessibility of research in accord with the National Institutes of Health data sharing policy and the National Cancer Institute Cancer Moonshot Public Access and Data Sharing Policy. The CIBMTR only releases de-identified datasets that comply with all relevant global regulations regarding privacy and confidentiality.

Data are available on request from the corresponding author, Roni Tamari ([tamarir@mskcc.org](mailto:tamarir@mskcc.org)).

The full-text version of this article contains a data supplement.

© 2023 by The American Society of Hematology. Licensed under [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International \(CC BY-NC-ND 4.0\)](https://creativecommons.org/licenses/by-nc-nd/4.0/), permitting only noncommercial, nonderivative use with attribution. All other rights reserved.

## Key Points

- This analysis used US and European stem cell transplant registries and provides a simple and easily applicable predictive system.

To develop a prognostic model for patients undergoing allogeneic hematopoietic cell transplantation (allo-HCT) for myelofibrosis (MF), we examined the data of 623 patients undergoing allo-HCT between 2000 and 2016 in the United States (the Center for International Blood and Marrow Transplant Research [CIBMTR] cohort). A Cox multivariable model was used to identify factors prognostic of mortality. A weighted score using these factors was assigned to patients who received transplantation in Europe (the European Bone Marrow Transplant [EBMT] cohort;  $n = 623$ ). Patient age  $>50$  years (hazard ratio [HR], 1.39; 95% confidence interval [CI], 0.98-1.96), and HLA-matched unrelated donor (HR, 1.29; 95% CI, 0.98-1.7) were associated with an increased hazard of death and were assigned 1 point. Hemoglobin levels  $<100$  g/L at time of transplantation (HR, 1.63; 95% CI, 1.2-2.19) and a mismatched unrelated donor (HR, 1.78; 95% CI, 1.25-2.52) were assigned 2 points. The 3-year overall survival (OS) in patients with a low (1-2 points), intermediate (3-4 points), and high score (5 points) were 69% (95% CI, 61-76), 51% (95% CI, 46-56.4), and 34% (95% CI, 21-49), respectively ( $P < .001$ ). Increasing score was predictive of increased transplant-related mortality (TRM;  $P = .0017$ ) but not of relapse ( $P = .12$ ). The derived score was predictive of OS ( $P < .001$ ) and TRM ( $P = .002$ ) but not of relapse ( $P = .17$ ) in the EBMT cohort as well. The proposed system was prognostic of survival in 2 large cohorts, CIBMTR and EBMT, and can easily be applied by clinicians consulting patients with MF about the transplantation outcomes.

## Introduction

Myelofibrosis (MF) is a myeloproliferative neoplasm characterized by clonal myeloid proliferation, extramedullary hematopoiesis, peripheral cytopenias, bone marrow fibrosis, splenomegaly, and a heterogeneous symptom burden.<sup>1,2</sup> The discovery of the JAK2V617F driver mutation and subsequent introduction of JAK inhibitors into the therapeutic arena has had a significant impact on clinical care of MF.<sup>3-7</sup> Despite this, allogeneic hematopoietic cell transplantation (allo-HCT) remains the only curative therapy to date.<sup>8,9</sup>

The dynamic international prognostic scoring system (DIPSS)<sup>10</sup> and DIPSS-plus<sup>11</sup> are commonly used for disease risk stratification, and the data regarding the mutations landscape<sup>12,13</sup> add further prognostic information. However, these tools were reported mostly in cohorts without transplantations, yet, the transplantation outcomes are dependent not only on disease characteristics, and both patient- and transplantation-related factors affect outcomes. The MPD-101 study reported superior outcomes after a reduced intensity conditioning in patients who received a transplantation from a matched related donor compared with those who received a transplantation from an unrelated donor,<sup>14</sup> whereas a prospective study by the European group of blood and marrow transplantation reported that being of younger age ( $<55$  years) and having a matched donor were associated with improved outcomes.<sup>15</sup> In addition, transplantation-related mortality (TRM) in patients with MF undergoing allo-HCT is noted to be higher than that in patients with other myeloid malignancies, reportedly as high as 35% at 5 years after transplantation compared with 20% in patients with acute myelogenous leukemia.<sup>16</sup> These observations highlight some unique characteristics of the disease, such as an increased inflammatory milieu and extramedullary hematopoiesis as well as

the interaction between the host and donor among patients with MF, emphasizing the need for a prognostic score among patients with MF undergoing allo-HCT.

In this study, we used data from the Center for International Blood and Marrow Transplant Research (CIBMTR) registry to identify disease-, patient-, and transplantation-specific variables that are associated with the outcome in patients undergoing allo-HCT for MF; this cohort served as the training set. The primary objective of this study was to develop a simple clinical risk stratification tool prognostic of survival after transplantation. We then sought to determine whether this tool was also prognostic of relapse, TRM, and disease-free survival (DFS). The model was validated in an external cohort of patients reported in the European Bone Marrow Transplant (EBMT) registry.

## Patients and methods

### Data source

The study was performed through a collaboration between the CIBMTR Chronic Leukemia Working Committee and the EBMT Chronic Malignancies Working Party. The CIBMTR is a nonprofit research collaboration of the National Marrow Donor Program/Be The Match and the Medical College of Wisconsin. More than 330 medical centers worldwide submit to the CIBMTR clinical data about HCT and other cellular therapies. Participating centers are required to report all transplantations consecutively. The CIBMTR ensures data quality through computerized checks for discrepancies, physicians' review of submitted data, and on-site audits of participating centers. The CIBMTR complies with federal regulations that protect human research participants. The institutional review boards of the Medical College of Wisconsin and the National Marrow Donor

Program approved this study. EBMT is a voluntary organization comprising >600 transplantation centers, mainly from Europe. Accreditation as a member-center requires submission of a minimal essential data form from all consecutive patients to a central registry. Since 1996, accredited EBMT centers are subject to on-site audits. Since January 2003, all transplantation centers have been required to obtain written informed consent in accordance with the Declaration of Helsinki 1975 before data registration.

## Patients

Patients with MF aged  $\geq 40$  years who underwent allo-HCT from a related HLA-matched donor (as previously described<sup>17</sup>), unrelated HLA-matched donor (MURD), or unrelated HLA-mismatched donor (MMURD) reported to the CIBMTR from 2000 to 2016 were included. Patients undergoing syngeneic umbilical cord blood- (n = 18) or mismatched related-donor transplantation (haplo-identical) (n = 73) and patients with missing donor data (n = 2) were excluded from this analysis, in addition to 15 patients whose graft-versus-host disease (GVHD) prophylaxis was by ex vivo T-cell depletion (n = 5) or CD34<sup>+</sup>-selection procedure (n = 10). An additional 35 patients were removed because of missing date of diagnosis, unknown GVHD prophylaxis, or missing complete 100-day follow-up data. A total of 623 patients met all inclusion criteria and composed the CIBMTR cohort and, hence, were included in the multivariate analysis. The same inclusion criteria were applied for the EBMT cohort and resulted in 2672 candidates, of whom 623 patients had complete data for the required ultimately scoring system components and were included in the multivariate analysis.

## Statistical analysis

A total of 623 patients in the CIBMTR cohort were included to develop a Cox regression model, and 623 patients in the EBMT cohort were included for external validation. The scoring model and prognostic variables were constructed and selected based on the CIBMTR data, and the scoring system was then validated on the EBMT data. Ruxolitinib, a JAK1/JAK2 inhibitor, which was the first drug to gain approval by both the US Food and Drug Administration and the European Medicines Agency for treatment of MF,<sup>18</sup> was introduced into clinical practice in 2012. We, therefore, tested whether the introduction of ruxolitinib significantly affected the potential findings from this study; 3 cohorts with respect to commercial availability of ruxolitinib were tested: patients who received transplantation before 2012, patients undergoing allo-HCT after 2012 who were treated with ruxolitinib, and patients who received transplantation after 2012 and were not treated with ruxolitinib. Because this variable and interactions with this variable were not significant, the full cohort was used for subsequent analysis. A Cox proportional hazards model with stepwise selection procedure was used to select significant covariates for overall survival (OS), at a significance level of 0.1. A raw score (0, 1, or 2) for each risk factor was assigned based on the magnitude of log hazard ratios (HRs) in the Cox model. Next, scores were calculated for each patient, and a univariable Cox model with those scores were fitted. Finally, the risk score (low, intermediate, or high) was determined based on the HRs of the univariable Cox model with the scores. The new scoring system was evaluated with the validation data set (EBMT). In the CIBMTR cohort, The new scoring system was compared with DIPSS. After construction of the risk

score for the main outcome, OS, the analysis was performed for the secondary outcomes including DFS, relapse, and TRM. Probabilities of OS and DFS for each risk group were calculated using the Kaplan Meier estimator, with the variance estimated using Greenwood formula. Probabilities of TRM and relapse were generated using cumulative incidence estimates to accommodate the competing risk event. SAS version 9.4 (SAS Institute, Cary, NC) and R version 4.0 (R Foundation for Statistical Computing, Vienna, Austria) were used for the analysis.

## The development of a prognostic scoring system

Variables related to patients, donors, disease, treatment before transplantation, and transplantation itself were included in the analysis (supplemental Table 1). We then constructed a Cox proportional hazards model using the CIBMTR training set that included the following nonmodifiable variables: age, hemoglobin, white blood cell count, platelet count, constitutional symptoms, circulating blasts at diagnosis and at the time of allo-HCT, sex, Karnofsky performance status before allo-HCT, time from the date of diagnosis to allo-HCT, cytogenetics abnormalities, JAK 2 mutation (present or absent), spleen status at allo-HCT, and donor type. A multivariable model identified 3 independent predictors of survival: age, hemoglobin at time of allo-HCT, and donor type.

## Results

### Patients

Patient data for the CIBMTR and EBMT cohorts are listed in Table 1 and supplemental Table 1. The 2 cohorts were noted with differences in patient-, disease- and transplantation-related variables. The percentage of patients for whom follow-up data were reported was 98%, 94%, 91%, and 89% at 1, 2, 3, and 4 years, respectively, in the CIBMTR cohorts. Although the cohorts included patients with primary MF and secondary MF, the majority in both cohorts were patients with primary MF: 87% in the CIBMTR and 80% in the EBMT cohorts.

The OS rates at 1, 3, and 5 years were 65.7% (95% confidence interval [CI], 61.9-69.4), 54.6% (95% CI, 50.4-58.7), and 49.9% (95% CI, 45.5-54.3), respectively (Figure 1). DFS at 1, 3, and 5 years were 39.7% (95% CI, 35.7-43.7), 31.1% (95% CI, 27.3-34.9), and 26.5% (95% CI, 22.7-30.5), respectively. Rates of TRM at 1, 3, and 5 years were 20.6% (95% CI, 17.4-23.9), 24.7% (95% CI, 21.3-28.3), and 27.1% (95% CI, 23.5-31), respectively. The relapse rates at 1, 3, and 5 years were 39.7% (95% CI, 35.8-43.6), 44.2% (95% CI, 40.2-48.3), and 46.3% (95% CI, 42.2-50.5), respectively, in the CIBMTR cohort.

### Prognostic scoring system

Table 2 summarizes the variables relevant to OS identified in the multivariable analysis of data of the patients in the CIBMTR cohort. Based on an HR  $\geq 1.5$ , a weighted score of 2 was assigned to a hemoglobin level  $< 100$  g/L at the time of allo-HCT and having a MMURD, whereas other factors were assigned a score of 0 or 1, based on an HR  $\leq 1$  and from 1 to 1.5. The overall score ranged from 0 to 5, with increasing scores indicating greater risk (supplemental Table 2). Based on HR of the univariate Cox model with raw scores, we created a 3-category system: low, score from

**Table 1. Baseline characteristics for patients undergoing allo-HCT for MF from 2000 to 2016, who were included in the CIBMTR and EBMT cohorts**

Variable	CIBMTR	EBMT	P value
No. of patients	623	623	
Median follow-up of survivors (range), mo	42 (3-193)	83 (3-219)	
<b>Patient related</b>			
<i>Age at diagnosis, median (range), y</i>	54 (40-75)	52 (40-74)	<.01†
<i>Age at HCT, median (range), y</i>	58 (40-76)	57 (41-74)	<.01†
Sex			.12‡
Male	395 (63%)	421 (68%)	
<b>Karnofsky performance status score before HCT</b>			
90-100	373 (60%)	313 (50%)	<.01‡
<b>HCT-CI</b>			
0	113 (18%)	201 (32%)	<.01‡
1	62 (10%)	53 (9%)	
2	67 (11%)	41 (7%)	
3+	174 (28%)	84 (13%)	
<b>Disease related</b>			
<i>Disease at diagnosis</i>			<.01‡
MF	542 (87%)	499 (80%)	
Polycythemia vera	32 (5%)	52 (8%)	
Essential thrombocythemia	49 (8%)	52 (8%)	
Polycythemia vera/essential thrombocythemia		20 (3%)	
<i>Blast in peripheral blood of &gt;1% at diagnosis</i>	89 (14%)	104 (17%)	.09‡
<i>Hemoglobin level &lt;100 g/L at diagnosis</i>	216 (35%)	214 (34%)	<.01‡
<i>WBC count &gt;25 × 10<sup>9</sup>/L at diagnosis</i>	59 (9%)	49 (8%)	<.01‡
<i>Platelet count at diagnosis, 50 × 10<sup>9</sup>/L–100 × 10<sup>9</sup>/L</i>	80 (13%)	87 (14%)	<.01‡
<i>Constitutional symptoms at diagnosis</i>	183 (29%)	176 (28%)	<.01‡
<i>Blast in peripheral blood &gt;1% before HCT</i>	188 (30%)	200 (32%)	<.01‡
<i>Hemoglobin level &lt;100 g/L before HCT</i>	442 (71%)	411 (66%)	.06‡
<i>WBC count &gt;25 × 10<sup>9</sup>/L before HCT</i>	82 (13%)	95 (15%)	<.01‡
<i>Platelet count 50 × 10<sup>9</sup>/L–100 × 10<sup>9</sup>/L before HCT</i>	133 (21%)	107 (17%)	<.01‡
<i>Constitutional symptoms before HCT</i>	104 (17%)	181 (29%)	<.01‡
<b>DIPSS before HCT</b>			
Low	76 (12%)		
Intermediate-1	283 (45%)		
Intermediate-2	236 (38%)		
High	11 (2%)		
<b>Cytogenetics</b>			
Favorable (normal)	251 (40%)		
Favorable (other)	113 (18%)		
Unfavorable	113 (18%)		
Not tested	34 (5%)		
<b>JAK2 mutation</b>			
Yes	202 (32%)	213 (34%)	<.01‡
<b>Spleen status</b>			
Normal	132 (21%)	81 (13%)	<.01‡

Nonmodifiable variables are indicated in italic.

CSA, cyclosporine; Cy, cyclophosphamide; HCT-CI, HCT-specific comorbidity index; MAC, myeloablative conditioning; MMF, mycophenolate mofetil; MTX, methotrexate; NMA, nonmyeloablative; RIC, reduced intensity conditioning; TAC, tacrolimus; TBI, total body irradiation; WBC, white blood cell.

\*Missing data are presented in the supplemental Tables.

†Kruskal Wallis test.

‡Pearson  $\chi^2$  test.

Table 1 (continued)

Variable	CIBMTR	EBMT	P value
Splenomegaly	451 (72%)	304 (49%)	
Splenectomy	23 (4%)	90 (14%)	
<b>Treatment-related, nontransplantation</b>			
Prior therapy			<.01#
Yes	468 (75%)	340 (55%)	
Missing	4 (1%)	55 (9%)	
Number of lines of pretreatments			<.01#
0	151 (24%)	228 (37%)	
1	255 (41%)	126 (20%)	
2	108 (17%)	18 (3%)	
≥3	101 (16%)	16 (3%)	
Received Jakafi as prior therapy			<.01#
Yes	175 (28%)	85 (14%)	
<b>Treatment-related, transplantation</b>			
<i>Time from diagnosis to HCT (mo)</i>	18 (2-294)	26 (2-268)	<.01†
<i>Donor type</i>			<.01#
HLA-identical sibling	221 (35%)	469 (75%)	
Well-matched unrelated	322 (52%)	107 (17%)	
Partially matched unrelated	80 (13%)	47 (8%)	
Sex match of donor and recipient			.04#
M-M	257 (41%)	253 (41%)	
M-F	134 (22%)	168 (27%)	
F-M	136 (22%)	107 (17%)	
F-F	93 (15%)	95 (15%)	
Graft source			.58#
Peripheral blood	554 (89%)	560 (90%)	
Use of TBI			.59#
No	526 (84%)	530 (85%)	
Conditioning regimen intensity			<.01#
MAC	285 (46%)	181 (29%)	
RIC	292 (47%)	440 (71%)	
NMA	37 (6%)		
GVHD prophylaxis			<.01#
Post-CY + other(s)	9 (1%)	14 (2%)	
TAC + MMF ± other(s) (except post-CY)	88 (14%)	3 (0)	
TAC + MTX ± other(s) (except MMF, post-CY)	281 (45%)	6 (1%)	
TAC + other(s) (except MMF, MTX, post-CY)	32 (5%)	7 (1%)	
TAC alone	11 (2%)	3 (0)	
CSA + MMF ± other(s) (except post-CY)	63 (10%)	193 (31%)	
CSA + MTX ± other(s) (except MMF, post-CY)	111 (18%)	258 (41%)	
CSA + other(s) (except MMF, MTX, post-CY)	6 (1%)	15 (2%)	
CSA alone	12 (2%)	82 (13%)	
Other(s)	8 (1%)	15 (2%)	
Median follow-up of survivors (range), mo	42 (3-193)	83 (3-219)	

Nonmodifiable variables are indicated in *italic*.

CSA, cyclosporine; Cy, cyclophosphamide; HCT-CI, HCT-specific comorbidity index; MAC, myeloablative conditioning; MMF, mycophenolate mofetil; MTX, methotrexate; NMA, nonmyeloablative; RIC, reduced intensity conditioning; TAC, tacrolimus; TBI, total body irradiation; WBC, white blood cell.

\*Missing data are presented in the supplemental Tables.

†Kruskal Wallis test.

#Pearson  $\chi^2$  test.

**Table 2. Multivariable analysis**

OS (CIBMTR)		95% CI		95% CI		Overall	
Age (y) at HCT	HR	Lower limit	Upper limit	P value	P value	n	
≥50	1				.0694	86	0
>50	1.382	0.975	1.96	.0694		537	1
<b>Hemoglobin before HCT</b>							
≥100 g/L	1				.0011	148	0
<100 g/L	1.631	1.217	2.185	.0011		475	2
<b>Donor type</b>							
HLA-identical siblings	1				.0048	221	0
Well-matched URD	1.284	0.997	1.654	.0527		322	1
Partially matched URD	1.776	1.251	2.521	.0013		80	2
<b>Contrast</b>							
Well-matched URD vs partially matched URD	0.7232	0.5197	1.0065	.0546			

1 to 2 (HR, 1-1.5); intermediate, score from 3 to 4 (HR, 1.5-2.5); and high, score from 5 (HR, >2.5).

The HR for death (using the low-risk group as reference) was 1.64 (95% CI, 1.23-2.18) for the intermediate-risk group, and 2.65 (95% CI, 1.70-4.14) for the high-risk group (overall  $P = .0002$ ; Table 3). This 3-category system was predictive for DFS with a HR for death of 1.44 (95% CI, 1.14-1.81) for the intermediate-risk group, and 1.83 (95% CI, 1.24-2.71) for the high-risk group (overall  $P = .0015$ ; supplemental Table 3) and for TRM as well; the HR for death was 1.63 (95% CI, 1.10-2.44) for the intermediate-risk group, and 3.09 (95% CI, 1.75-5.48) for the high-risk group (overall  $P = .0017$ ; supplemental Table 3). This prognostic system was not predictive of relapse.

Considering that spleen size and splenectomy are a matter of debate regarding transplantation outcomes in patients with MF, we examined size of a healthy spleen vs one with splenomegaly vs with splenectomy. Patients who underwent a splenectomy had worse OS, but there was no difference in survival when comparing patients with splenomegaly vs those with normal spleen size. Yet, a very small

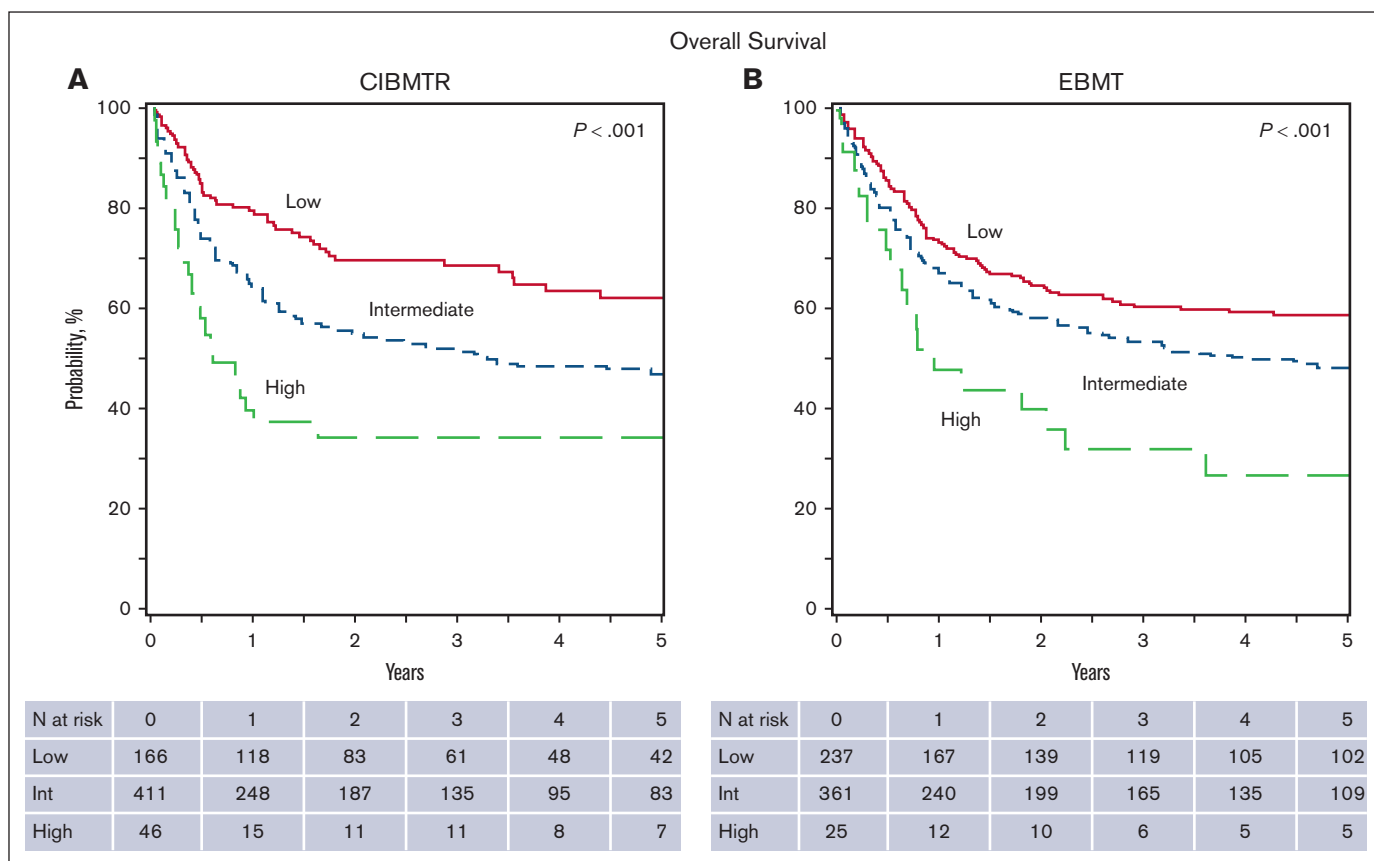
proportion of patients in this analysis had a splenectomy (4% in the CIBMTR cohort and 14% of the EBMT cohort), which limits the ability to draw conclusions on the role of splenectomy in outcomes in these cohorts.

### Application of CIBMTR scoring system to the EBMT cohort for validation

The prognostic score was applied to an external EBMT cohort for validation. Analyzed data within this data set comprised data from 623 patients for whom complete data were available. When compared with the CIBMTR cohort, the 1-, 3-, and 5-year OS were 68.6% (95% CI, 64.9-72.2), 55.0% (95% CI, 51.0-58.9), and 51.2% (95% CI, 47.1-55.2) in the EBMT cohort, respectively (Figure 1). In the EBMT cohort, the 3-year incidence of TRM was 27.9% (95% CI, 24.4-31.6), and relapse was 24.3% (95% CI, 20.9-27.8). The 3-category system was prognostic of OS ( $P = .0011$ ), DFS ( $P = .0007$ ), and TRM ( $P = .0021$ ), and, again, was not predictive of relapse ( $P = .1673$ ; supplemental Table 4). Because of a significantly

**Table 3. OS by prognostic score in CIBMTR and EBMT cohorts**

CIBMTR						
Rank	HR	95% CI Lower limit	95% CI Upper limit	P value	Overall P value	n
Low	1	Reference			.0002	166
Intermediate	1.636	1.226	2.183	.0008		411
High	2.649	1.695	4.138	<.0001		46
<b>Contrast</b>						
Intermediate vs high	0.6177	0.4186	0.9114	.0152		
EBMT						
Rank	HR	95% CI Lower limit	95% CI Upper limit	P value	Overall P value	n
Low	1	Reference			.0011	237
Intermediate	1.336	1.054	1.694	.0167		361
High	2.353	1.442	3.839	.0006		25
<b>Contrast</b>						
Intermediate vs high	0.5678	0.3544	0.9097	.0186		



**Figure 1. Overall survival in the CIBMTR and EBMT cohorts by the prognostic scoring system classification.** (A) CIBMTR; (B) EBMT.

longer follow-up time in the EBMT cohort (83 vs 42 months in the CIBMTR cohort), a sensitivity analysis was performed on the EBMT cohort, censoring the data at 42 months (supplemental Table 5), showing similar results.

### Comparison of proposed scoring system with DIPSS score

To study how the proposed new scoring system compares with the DIPSS prognostic tool, we generated a cross table with the proposed score (rows) and the DIPSS classification (Figure 2). This included a total of 606 patients from the CIBMTR cohort for whom a DIPSS score was available. This comparison showed agreement on patients classified as being at low and high risk based on the DIPSS score; however, patients characterized as being at intermediate-1 and intermediate-2 risk were distributed among all risk categories in the new proposed model. The performance of the new scoring system is favorable compared with the DIPSS as indicated by the differences seen in the HRs and the c-statistics when the follow-up was censored at 36 months after transplantation. (supplemental Table 6).

### Study limitations

This study included patients treated over a long time period, from 2000 to 2016, a period during which there had been changes and advances in the field of stem cell transplantation, whether related to HLA typing, prevention, and management of GVHD; improvement in detection and treatment of infectious diseases; and the introduction

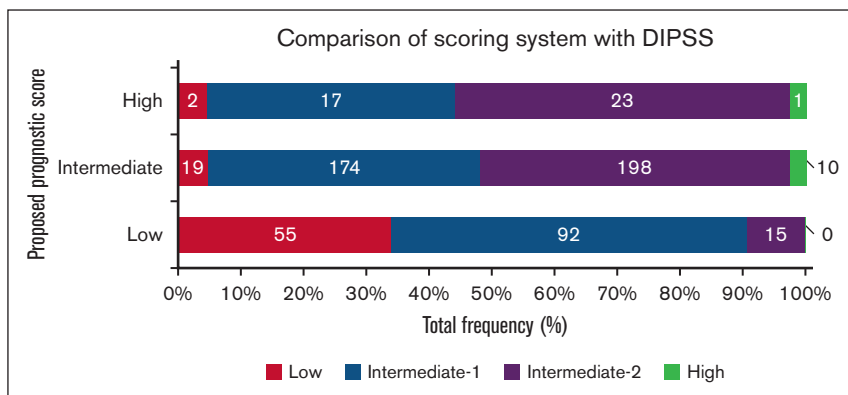
of new conditioning regimens that are not captured in this analysis. Of particular importance are the increased use of haplo-identical donors, which were excluded from this analysis because of very small numbers and will require follow-up studies, and the role of molecular mutations in prognostic scores of transplantations for MF. Furthermore, differences in practices between the United States and Europe as well insurance policies that affects transplantation referrals are beyond the scope of this analysis but need to be acknowledged. Lastly, the CIBMTR cohort did not have any patients with a score of 0, so no conclusions could be drawn from that group of patients.

### Discussion

In this large analysis, using CIBMTR and EBMT cohorts, we present a new scoring tool prognostic of outcomes in patients with MF undergoing allo-HCT. Using a multivariate analysis, we identified 3 simple and clinically relevant variables: patient age, pre-transplantation hemoglobin level, and donor type, to be prognostic for OS, DFS, and TRM.

The current available prognostic scoring systems such as the DIPSS, the DIPSS-plus, and, more recently, the mutation-enhanced international prognostic score system for transplantation-age patients with primary myelofibrosis (MIPSS-70),<sup>19</sup> were validated for patients with primary MF and can discriminate clearly between patients with MF who are at low or intermediate risk or with poor prognosis. Several studies using these prognostic systems in transplantation cohorts





**Figure 2. Categorization of patients based on the proposed scoring system vs the DIPSS.** Colored bars represent the DIPSS on the x-axis, within the stratification based on the new scoring system in rows.

have found them to be predictive, as expected, based on disease biology; however, the effects of transplantation-related factors, such as conditioning intensity and type of donors varied in the different studies.<sup>20,21</sup> Here, we note that the proposed system correlated well with the low- and high-risk DIPSS groups; however, for patients in the intermediate-1 and intermediate-2 risk groups, the new proposed system reassigned them to a different group in 45% of intermediate-1 risk group cases and in 16% of intermediate-2 risk group cases. This is particularly important in cases for which the DIPSS score is predictive of worse transplantation outcomes; for the same patients, if an HLA-matched related donor is available, the new score would be intermediate and associated with better outcomes than what the DIPSS scoring system would predict. In contrast, for patients with favorable outcomes based on the DIPSS score, if an unrelated donor is the only option, transplantation outcomes are predicted to be worse. This comparison between the DIPSS and the new transplantation-specific scoring system highlights that specific transplantation-related factors and, particularly, the graft source (ie, donor) are important and can overcome high-risk disease features. The findings, however, do not suggest that one scoring system is superior to the other but, rather, that they complement each other.

In this analysis, having a HLA-matched related donor was associated with better outcomes compared with both HLA-MURD and mismatched donors, which has been reported previously in patients with MF undergoing allo-HCT.<sup>22-24</sup> At the same time, the use of alternative donors such as haplo-identical donors and mismatched unrelated donors is growing rapidly.<sup>25</sup> A recent study by Kunte et al<sup>26</sup> reported overall good outcomes among 69 patients with MF who received transplantation using haplo-identical donors; at 3 years, the OS, relapse free survival, and GVHD-free relapse free survival were 72% (95% CI, 59-81), 44% (95% CI, 29-59), and 30% (95% CI, 17-43), respectively. Of note, the majority of patients included within this study (80%) received transplantation after 2016, with data being suggestive of changes in trends in recent years.

The impact of the spleen size, and intervention to reduce the spleen size via splenic radiation or splenectomy, on transplantation outcomes remain a matter of debate. A large analysis from EBMT<sup>27</sup> that included 1195 patients with MF who received transplantation between 2000 and 2017 reported that splenectomy was associated with lower rates of TRM but increased risk of relapse, without a significant effect on OS. This analysis highlights that, although

pretransplant splenectomy was done more commonly in the early years (in 28.3% before 2009 compared with in 14.1% after 2009), the numbers decreased further after the introduction of ruxolitinib in 2012. A very small proportion of patients in this analysis had a splenectomy, which limits the ability to draw conclusions on the role of splenectomy in outcomes in this cohort. Data suggest that the treatment with ruxolitinib before transplantation improves outcomes, whether by its effects on inflammatory milieu or by effect on spleen size,<sup>28</sup> and further studies to address this question in the era of JAK-STAT inhibitors are needed.

More recently, next-generation sequencing has been incorporated into contemporary predictive models for MF and several studies have evaluated its prognostic power in the context of allo-HCT as well. In a study by Gagelmann et al<sup>29</sup> incorporating clinical, molecular, and transplant-related factors to create the MF transplantation scoring system, patient age and unrelated donor (matched or mismatched) were predictive for outcomes similar to the findings from our analysis. The MF transplantation scoring system included data on molecular mutations; the presence of mutations in ASXL1 gene and non-CALR/MPL driver mutations genotype were predictive of outcomes, whereas a high number of mutations (>3) and the presence of so-called high-risk mutations were not. Several other retrospective analyses evaluated the role of molecular mutations and outcomes of patients with MF undergoing allo-HCT with conflicting conclusions.<sup>30-33</sup> Our proposed system lacks any data on molecular mutations because such data were not available in the earlier years of this study period nor collected systematically until very recently. However, the conflicting findings regarding the prognostic power of molecular mutations in the context of allo-HCT, and the lack of standardization of next-generation sequencing assays with relation to the genes involved in different panels, the levels of detection that are considered pathogenic, validation, and clinical interpretation<sup>34</sup> suggest that further and larger studies are needed to determine the prognostic role of mutational analysis in predicting transplant outcomes in patients with MF.

In summary, despite significant differences between the 2 cohorts, the proposed model was predictive of outcomes in these 2 large data sets. As highlighted earlier, we acknowledge several limitations, which are inherent to the nature of the study being a retrospective analysis of registry-reported data. Despite this, this analysis resulted in a simple, clinically relevant, and easily applicable score that, in addition to the current available risk scores, may help in the counseling of patients with MF undergoing allo-HCT.

## Acknowledgments

The CIBMTR is supported primarily by Public Health Service U24CA076518 from the National Cancer Institute, the National Heart, Lung, and Blood Institute, and the National Institute of Allergy and Infectious Diseases; HHS250201700006C from the Health Resources and Services Administration; and N00014-20-1-2832 and N00014-21-1-2954 from the Office of Naval Research. Support is also provided by Be the Mazak Foundation, the Medical College of Wisconsin, the National Marrow Donor Program, and from the following commercial entities: AbbVie, Actinium Pharmaceuticals Inc, Adaptive Biotechnologies Corporation, ADC Therapeutics, Adienne SA, Allogene, Allovir Inc, Amgen Inc, Anthem, Astellas Pharma US, AstraZeneca, Atara Biotherapeutics, BeiGene, bluebird bio Inc, Bristol Myers Squibb Co, CareDx Inc, CRISPR, CSL Behring, CytoSen Therapeutics Inc, Eurofins Viracor, DBA Eurofins Transplant Diagnostics, Fate Therapeutics, Gamida-Cell Ltd, Gilead, GlaxoSmithKline, HistoGenetics, Incyte Corporation, Iovance, Janssen Research & Development LLC, Janssen/Johnson & Johnson, Jasper Therapeutics, Jazz Pharmaceuticals Inc, Kadmon, Karius, Kiadis Pharma, Kite, Kyowa Kirin, Legend Biotech, Magenta Therapeutics, Mallinckrodt Pharmaceuticals, Medac GmbH, Medexus Pharma, Merck & Co, Millennium, the Takeda Oncology Co, Miltenyi Biotec Inc, MorphoSys, Novartis Pharmaceuticals Corporation, Omeros Corporation, OptumHealth, Orca Biosystems Inc, Ossium Health Inc, Pfizer Inc, Pharmacyclics LLC, Priothera, Sanofi, Sanofi-Aventis US Inc, Sobi Inc, Stemcyte, Takeda Pharmaceuticals, Talaris Therapeutics, Terumo Blood and Cell Technologies, TG Therapeutics, Vertex Pharmaceuticals, and Xenikos BV.

## Authorship

Contribution: R.T., K.W.A., N.E.-M., and W.S. conceptualized and designed the study; CIBMTR provided financial support and performed data collection and assembly; R.T., K.W.A., N.E.-M., and W.S. performed data analysis; R.T. and W.S. wrote the manuscript; and all authors interpreted the data and approved the final version of the manuscript.

Conflict-of-interest disclosure: M.B. reports research support to the institute from Novartis. V.G. reports consultancy work for Novartis, Incyte, Bristol Myers Squibb (BMS)-Celgene, Sierra Oncology, MorphoSys, Pfizer, and Takeda, and received a research

grant through the institution from Novartis and Incyte. H.A. reports serving on the advisory board and speaker bureau for Incyte and BMS. U.G. reports payment or honoraria for lectures, presentations, speaker bureaus, manuscript writing, or education events. C.B. reports serving as an advisory board member of Kite and Novartis (unrelated to this study). D.R. reports serving as a speaker bureau and advisory boards for Incyte. S.G. reports as speaker for Seattle Genetics and Kite Pharma, and advisory board for Sanofi, BMS, Daiichi Sankyo, Astellas, Janssen, and AstraZeneca. T.J. reports institutional research support from CTI Biopharma, Syneos Health, Incyte, and advisory board participation with BMS, Incyte, and CTI. M.A.K.-D. reports consultancy for Daiichi Sankyo. J.A.Y. reports honorarium for 1-time advisory board meeting with Kite and Omeros, and research funding from Gilead. M.R.G. reports consulting fees from and advisory board involvement at AbbVie, Agios, Amgen, Astellas, Blueprint Medicines, BMS, Cardinal Health, CTI Biopharma, Daiichi Sankyo, Gamida-Cell, Gilead, Incyte, Invitae, Karius, Ono Pharmaceutical, Pfizer, Pharmacosmos, Premier, Sierra Oncology, Stemline, and Trovogene; stock ownership with Medtronic; medical writing for Incyte, Amgen, Jazz, Janssen, and Genentech/Roche; and research support from Incyte, Genentech/Roche, and Janssen. T.N. reports clinical trial research support from Novartis to the institution and clinical trial support (drug only supply) from Karyopharm to the institution. The remaining authors declare no competing financial interests.

ORCID profiles: R.T., [0000-0002-2386-0850](https://orcid.org/0000-0002-2386-0850); K.W.A., [0000-0003-4567-8037](https://orcid.org/0000-0003-4567-8037); J.C.H.-B., [0000-0002-4289-3113](https://orcid.org/0000-0002-4289-3113); S.G., [0000-0003-1944-5053](https://orcid.org/0000-0003-1944-5053); Z.D., [0000-0002-7994-8974](https://orcid.org/0000-0002-7994-8974); V.G., [0000-0002-1419-8607](https://orcid.org/0000-0002-1419-8607); H.A., [0000-0002-9728-7292](https://orcid.org/0000-0002-9728-7292); M.R.L., [0000-0002-9816-6302](https://orcid.org/0000-0002-9816-6302); S.H., [0000-0002-8914-7927](https://orcid.org/0000-0002-8914-7927); J.J.P., [0000-0001-7498-3159](https://orcid.org/0000-0001-7498-3159); V.U.B., [0000-0001-8771-947X](https://orcid.org/0000-0001-8771-947X); T.J., [0000-0001-6854-773X](https://orcid.org/0000-0001-6854-773X); C.C., [0000-0001-8728-4314](https://orcid.org/0000-0001-8728-4314); M.A.K.-D., [0000-0001-7394-5185](https://orcid.org/0000-0001-7394-5185); J.A.Y., [0000-0002-5346-6299](https://orcid.org/0000-0002-5346-6299); T.N., [0000-0002-2621-7924](https://orcid.org/0000-0002-2621-7924); S.S., [0000-0002-4308-2152](https://orcid.org/0000-0002-4308-2152); B.L.S., [0000-0001-9620-7839](https://orcid.org/0000-0001-9620-7839); R.N., [0000-0002-9082-0680](https://orcid.org/0000-0002-9082-0680); T.C., [0000-0003-3108-4035](https://orcid.org/0000-0003-3108-4035); I.Y.-A., [0000-0003-4524-8782](https://orcid.org/0000-0003-4524-8782).

Correspondence: Roni Tamari, Department of Medicine, Bone Marrow Transplant Service, Memorial Sloan Kettering Cancer Center Department of Medicine, Weill Cornell Medical College, David H. Koch Center, 530 E 74th St, New York, NY 10021; email: [tamarir@mskcc.org](mailto:tamarir@mskcc.org).

## References

1. Arber DA, Orazi A, Hasserjian R, et al. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood*. 2016;127(20):2391-2405.
2. Tefferi A, Guglielmelli P, Pardanani A, Vannucchi AM. Myelofibrosis treatment algorithm 2018. *Blood Cancer J*. 2018;8(8):72.
3. Harrison C, Kiladjian JJ, Al-Ali HK, et al. JAK inhibition with ruxolitinib versus best available therapy for myelofibrosis. *N Engl J Med*. 2012;366(9):787-798.
4. James C, Ugo V, Le Couedic JP, et al. A unique clonal JAK2 mutation leading to constitutive signalling causes polycythaemia vera. *Nature*. 2005;434(7037):1144-1148.
5. Kralovics R, Passamonti F, Buser AS, et al. A gain-of-function mutation of JAK2 in myeloproliferative disorders. *N Engl J Med*. 2005;352(17):1779-1790.
6. Levine RL, Wadleigh M, Cools J, et al. Activating mutation in the tyrosine kinase JAK2 in polycythemia vera, essential thrombocythemia, and myeloid metaplasia with myelofibrosis. *Cancer Cell*. 2005;7(4):387-397.

7. Verstovsek S, Gotlib J, Mesa RA, et al. Long-term survival in patients treated with ruxolitinib for myelofibrosis: COMFORT-I and -II pooled analyses. *J Hematol Oncol*. 2017;10(1):156.
8. Jain T, Mesa RA, Palmer JM. Allogeneic stem cell transplantation in myelofibrosis. *Biol Blood Marrow Transplant*. 2017;23(9):1429-1436.
9. Gowin K, Ballen K, Ahn KW, et al. Survival following allogeneic transplant in patients with myelofibrosis. *Blood Adv*. 2020;4(9):1965-1973.
10. Passamonti F, Cervantes F, Vannucchi AM, et al. A dynamic prognostic model to predict survival in primary myelofibrosis: a study by the IWG-MRT (International Working Group for Myeloproliferative Neoplasms Research and Treatment). *Blood*. 2010;115(9):1703-1708.
11. Gangat N, Caramazza D, Vaidya R, et al. DIPSS plus: a refined dynamic international prognostic scoring system for primary myelofibrosis that incorporates prognostic information from karyotype, platelet count, and transfusion status. *J Clin Oncol*. 2011;29(4):392-397.
12. Vannucchi AM, Lasho TL, Guglielmelli P, et al. Mutations and prognosis in primary myelofibrosis. *Leukemia*. 2013;27(9):1861-1869.
13. Tefferi A, Lasho TL, Finke CM, et al. CALR vs JAK2 vs MPL-mutated or triple-negative myelofibrosis: clinical, cytogenetic and molecular comparisons. *Leukemia*. 2014;28(7):1472-1477.
14. Rondelli D, Goldberg JD, Isola L, et al. MPD-RC 101 prospective study of reduced-intensity allogeneic hematopoietic stem cell transplantation in patients with myelofibrosis. *Blood*. 2014;124(7):1183-1191.
15. Kroger N, Holler E, Kobbe G, et al. Allogeneic stem cell transplantation after reduced-intensity conditioning in patients with myelofibrosis: a prospective, multicenter study of the Chronic Leukemia Working Party of the European Group for Blood and Marrow Transplantation. *Blood*. 2009;114(26):5264-5270.
16. Perram J, Ross DM, McLornan D, et al. Innovative strategies to improve hematopoietic stem cell transplant outcomes in myelofibrosis. *Am J Hematol*. 2022;97(11):1464-1477.
17. Weisdorf D, Spellman S, Haagenson M, et al. Classification of HLA-matching for retrospective analysis of unrelated donor transplantation: revised definitions to predict survival. *Biol Blood Marrow Transplant*. 2008;14(7):748-758.
18. Mascarenhas J, Hoffman R. Ruxolitinib: the first FDA approved therapy for the treatment of myelofibrosis. *Clin Cancer Res*. 2012;18(11):3008-3014.
19. Guglielmelli P, Lasho TL, Rotunno G, et al. MIPSS70: mutation-enhanced international prognostic score system for transplantation-age patients with primary myelofibrosis. *J Clin Oncol*. 2018;36(4):310-318.
20. Scott BL, Gooley TA, Sorror ML, et al. The dynamic international prognostic scoring system for myelofibrosis predicts outcomes after hematopoietic cell transplantation. *Blood*. 2012;119(11):2657-2664.
21. Samuelson Bannow BT, Salit RB, Storer BE, et al. Hematopoietic cell transplantation for myelofibrosis: the dynamic international prognostic scoring system plus risk predicts post-transplant outcomes. *Biol Blood Marrow Transplant*. 2018;24(2):386-392.
22. Gupta V, Malone AK, Hari PN, et al. Reduced-intensity hematopoietic cell transplantation for patients with primary myelofibrosis: a cohort analysis from the Center for International Blood and Marrow Transplant Research. *Biol Blood Marrow Transplant*. 2014;20(1):89-97.
23. Keyzner A, Han S, Shapiro S, et al. Outcome of allogeneic hematopoietic stem cell transplantation for patients with chronic and advanced phase myelofibrosis. *Biol Blood Marrow Transplant*. 2016;22(12):2180-2186.
24. Robin M, Tabrizi R, Mohty M, et al. Allogeneic haematopoietic stem cell transplantation for myelofibrosis: a report of the Societe Francaise de Greffe de Moelle et de Therapie Cellulaire (SFGM-TC). *Br J Haematol*. 2011;152(3):331-339.
25. CIBMTR. The US summary slides - HCT trends and survival data. <https://www.cibmtr.org/ReferenceCenter/SlidesReports/SummarySlides/pages/index.aspx>
26. Kunte S, Rybicki L, Viswabandya A, et al. Allogeneic blood or marrow transplantation with haploidentical donor and post-transplantation cyclophosphamide in patients with myelofibrosis: a multicenter study. *Leukemia*. 2022;36(3):856-864.
27. Polverelli N, Mauff K, Kroger N, et al. Impact of spleen size and splenectomy on outcomes of allogeneic hematopoietic cell transplantation for myelofibrosis: a retrospective analysis by the chronic malignancies working party on behalf of European society for blood and marrow transplantation (EBMT). *Am J Hematol*. 2021;96(1):69-79.
28. Chhabra S, Narra RK, Wu R, et al. Fludarabine/busulfan conditioning-based allogeneic hematopoietic cell transplantation for myelofibrosis: role of ruxolitinib in improving survival outcomes. *Biol Blood Marrow Transplant*. 2020;26(5):893-901.
29. Gagelmann N, Ditschkowski M, Bogdanov R, et al. Comprehensive clinical-molecular transplant scoring system for myelofibrosis undergoing stem cell transplantation. *Blood*. 2019;133(20):2233-2242.
30. Tamari R, Rapaport F, Zhang N, et al. Impact of high-molecular-risk mutations on transplantation outcomes in patients with myelofibrosis. *Biol Blood Marrow Transplant*. 2019;25(6):1142-1151.
31. Ali H, Aldoss I, Yang D, et al. MIPSS70+ v2.0 predicts long-term survival in myelofibrosis after allogeneic HCT with the Flu/Mel conditioning regimen. *Blood Adv*. 2019;3(1):83-95.
32. Kroger N, Panagiota V, Badbaran A, et al. Impact of molecular genetics on outcome in myelofibrosis patients after allogeneic stem cell transplantation. *Biol Blood Marrow Transplant*. 2017;23(7):1095-1101.
33. Jain T, Tsai HL, DeZern AE, et al. Post-transplantation cyclophosphamide-based graft-versus-host disease prophylaxis with nonmyeloablative conditioning for blood or marrow transplantation for myelofibrosis. *Transplant Cell Ther*. 2022;28(5):259.e1-259.e11.
34. Conway JR, Warner JL, Rubinstein WS, Miller RS. Next-generation sequencing and the clinical oncology workflow: data challenges, proposed solutions, and a call to action. *JCO Precis Oncol*. 2019;3:1-10.