



High-throughput strain identification and production of fungal enzymatic cocktails for the valorisation of lignocellulosic biomass

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The **perfect** enzyme cocktail
for **biomass degradation**

Virtual lab tour:

<https://www.vip-studio360.fr/galerie360/visites/vv-centrale-lille/vv-realcat-c.html>

The REALCAT platform

REALCAT

- Advanced High-Throughput Technologies Platform for all types of experiments in Chemistry and Biology, dedicated to biomass valorization.

3 main areas of expertise

➤ Catalysis

- **Chemical catalysis:** homogeneous and heterogeneous
- **Biocatalysis:** enzyme and fermentation

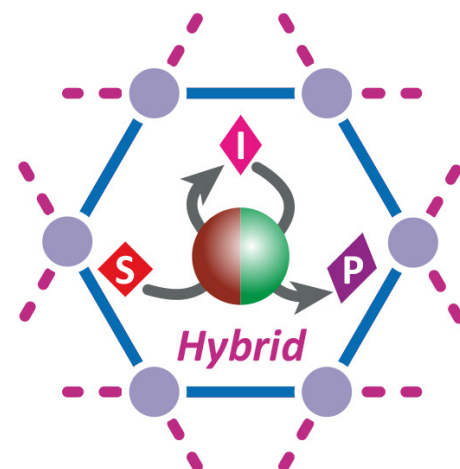
- **Structural and compositional characterisation** of chemical and biological materials used/generated (catalysts, metabolites, polymers, enzymes, etc.)

➤ Side activities in biology

- Proteomics, NRPomics
- Search for new antimicrobial molecules
- Production of secondary metabolites



*Combination of
chemo & Bio*



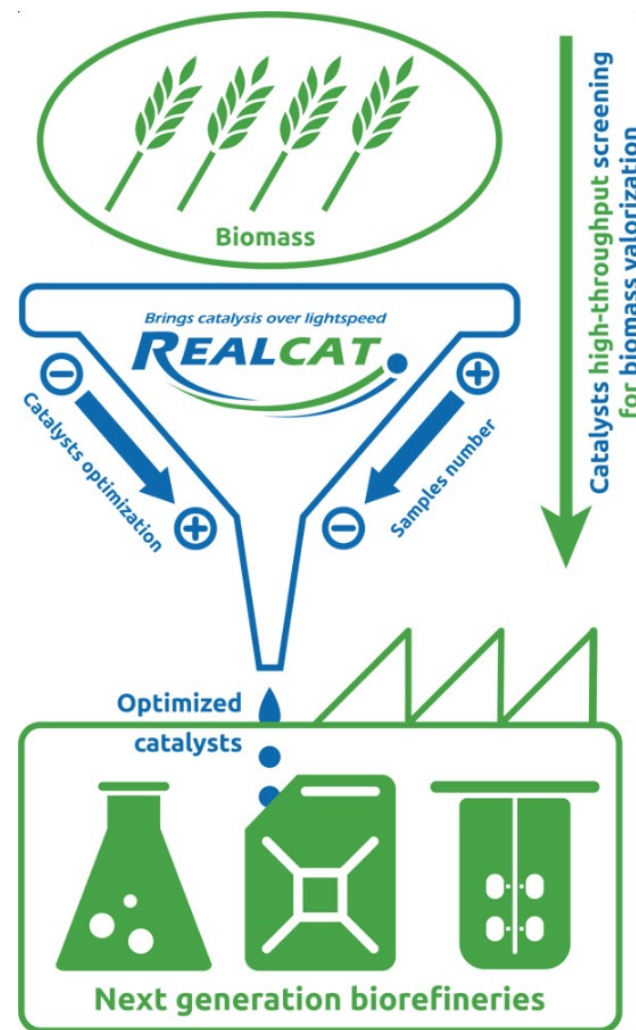
The REALCAT platform

3 main types of equipment dedicated to :

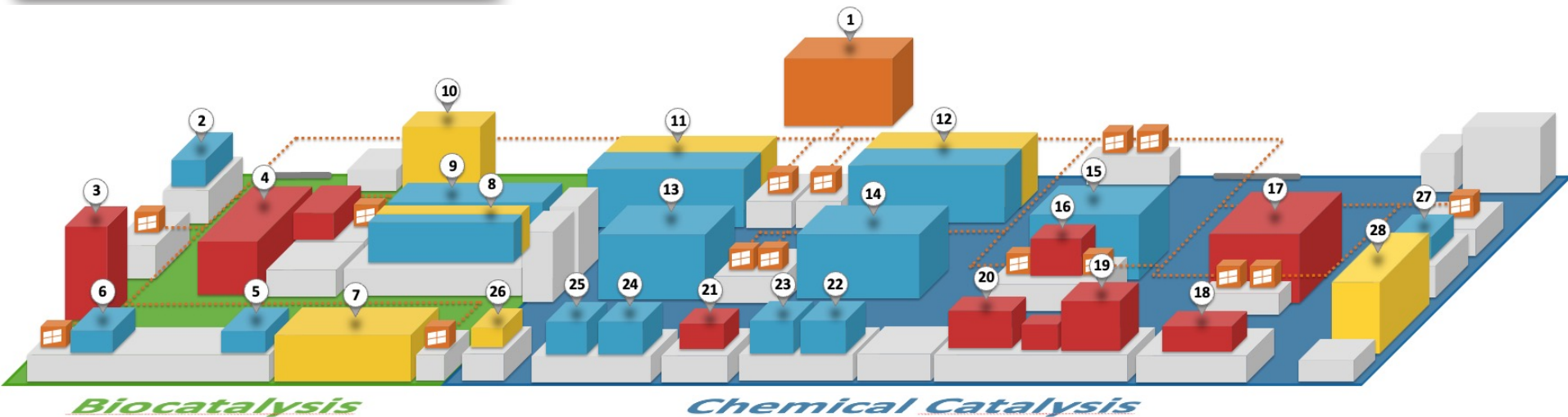
- **Synthesis** of chemical and biological materials (catalysts, strains, enzymes, metabolites, etc.)
- **Testing** of catalytic properties (catalytic, nutritional, antimicrobial, etc.)
- **Characterization** (structure, composition, physico-chemical properties, etc.)

Our final objectives :

- **Accelerate each step of the experimental phase** of a chemistry and biotechnology research project to significantly reduce the consumption of money and time
- Define new ways to **valorise biomass**



REALCAT floorplan



More than 30 automated devices

- All fully dedicated to high-throughput experiments
- Very high modularity allowing a large variety of subjects
- Fully integrated to allow synergy between machines

+ some other robots, technical rooms and offices.

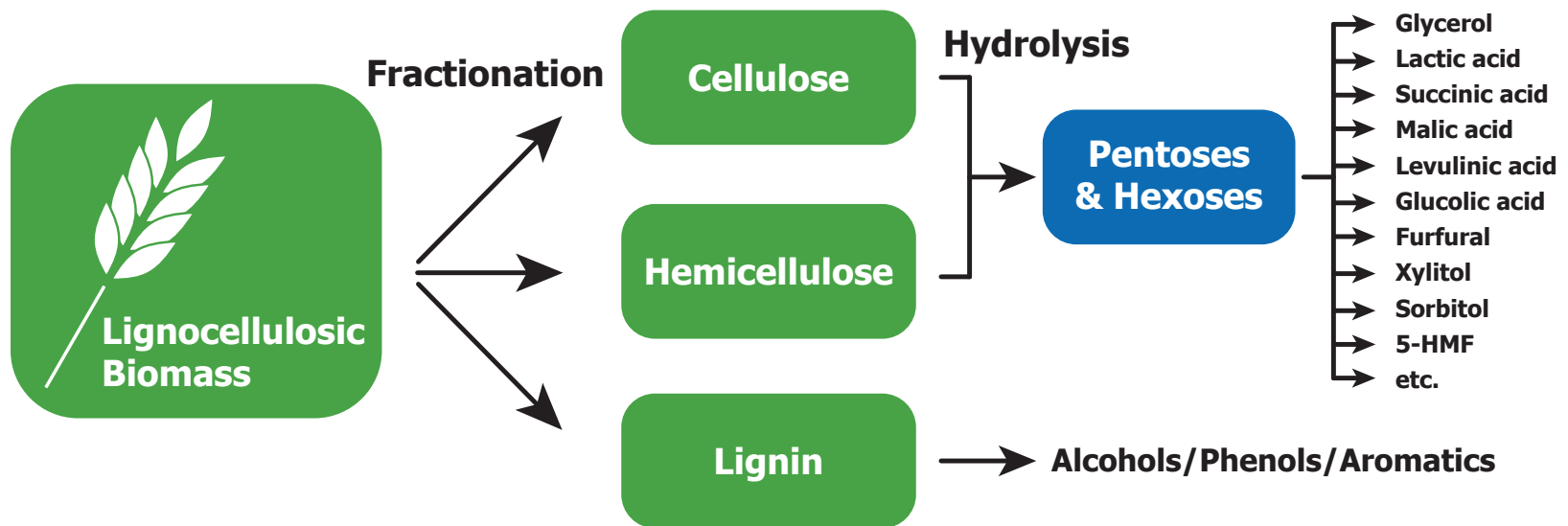
1	Central server - DELL	15	Flowrence - Avantium
2	Biomek NXp + BioLector Pro - Beckman Coulter/M2PLabs	16	M4 Tornado - Bruker
3	Autoflex Speed - Bruker	17	D8 Discover - Bruker
4	Acquity UPLC Synapt G2-Si HDMS - Waters	18	Tensor 37-HTS-XT - Bruker
5	Cary 3500 - Agilent	19	ICP-OES - Agilent
6	BioLector - M2pLabs	20	Vulcan 42S - Questron Technologies/Horiba
7	QPix 460 - Molecular Devices	21	XploRa - Horiba Jobin Yvon
8	Biomek FXp - Beckman Coulter	22	GC-FID-2010 Plus AF - Shimadzu
9	Biomek FXp - Beckman Coulter	23	GC-FID-MS-QP2010 Ultra EI - Shimadzu
10	Laminar flow hood - Aquaria	24	HPLC-UV-IR - Shimadzu
11	Catimpreg - Chemspeed	25	HPLC-DAD-MS - Shimadzu
12	Autoplant - Chemspeed	26	Calcination oven - Dislab
13	Flowrence - Avantium	27	Screening Pressure Reactor (SPR) - Freeslate
14	Flowrence - Avantium	28	Fume hood - Asem



Valorization of lignocellulosic biomass

Target: Biomass decomposition into biofuels, building blocks for fine chemistry and polymers:

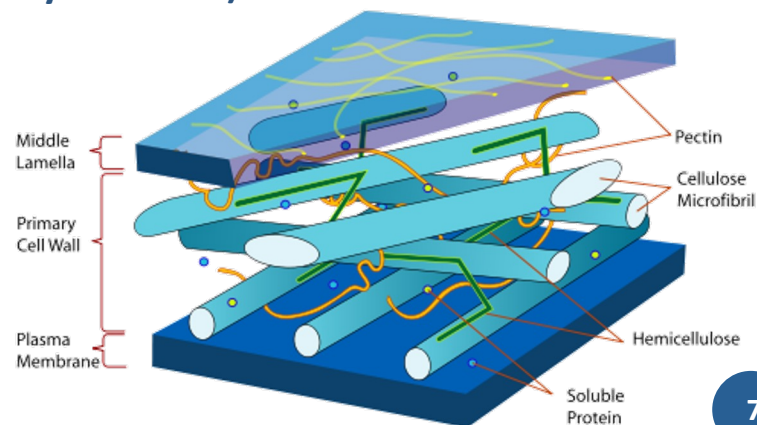
- Production of bioethanol from the fermentation of extracted sugars
- Production of 20 platform molecules from C5 and C6 sugars



- + alcohols/phenols/aromatics from lignin: methanol, benzoic acid, catechol, cinnamic acid, etc.

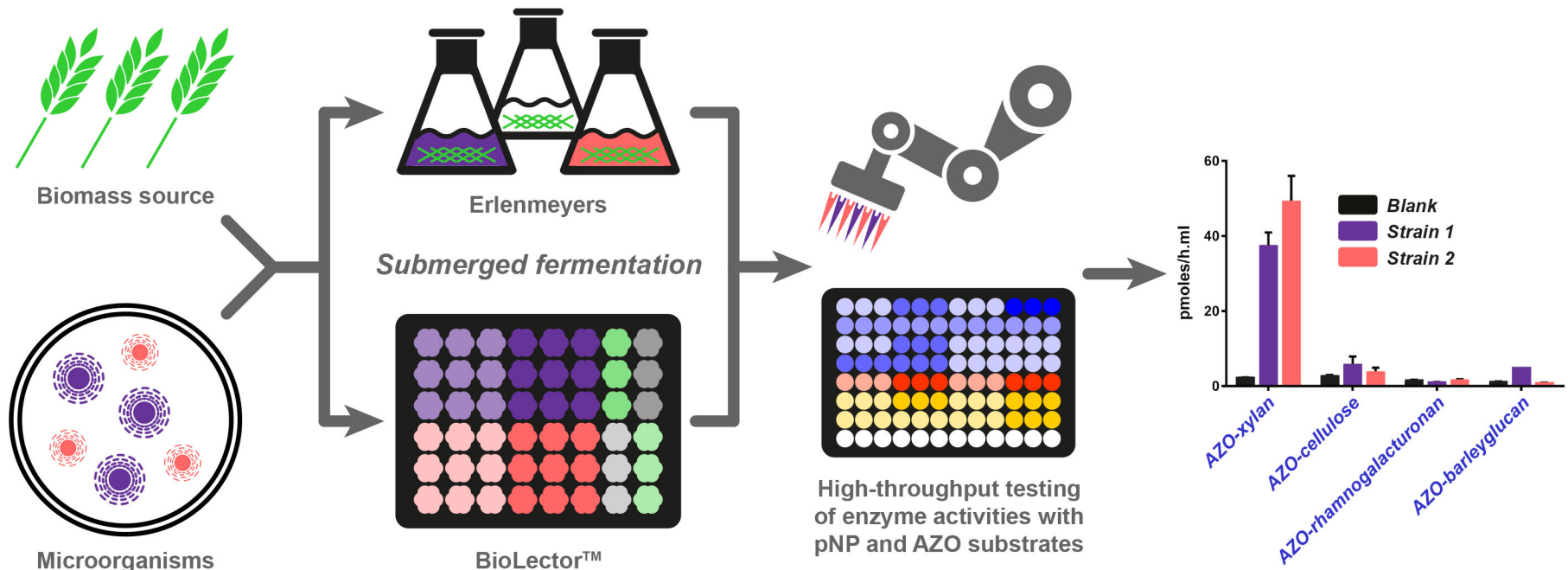
The challenge: designing the perfect enzyme cocktail for lignocellulose degradation

- Ideal case: no pre-treatment (mimic nature)
- No “universal” bacterial/fungal strain efficient on cellulose, hemicellulose AND lignin => Find the perfect enzyme combination
- Many available approaches: metagenomics, low throughput fermentation screenings, *de novo* design, commercial blends...
- Numerous drawbacks:
 - Very time and money consuming
 - High systems complexity (specially in biodiversity mixtures)
 - Model substrates are not representative
 - Lack of synergy and thermodynamic equilibriums issues



Harvesting enzyme from microorganisms living on target substrate

- Premise 1: Organisms that develop on biomass are the best equipped to degrade it
- Premise 2: No single organism can proceed to a complete degradation
- ⇒ Need for harvesting and combining numerous strains
- First semi-automated method presented in 2018





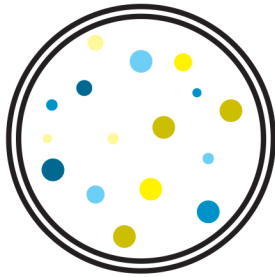
Biomass



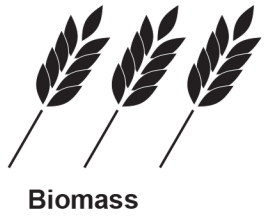
Biomass



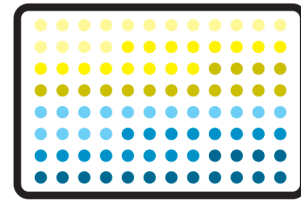
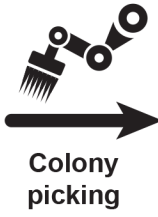
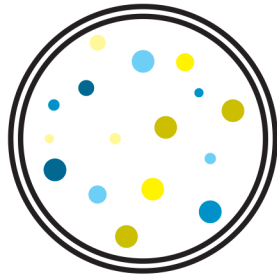
**Bacteria
isolation**

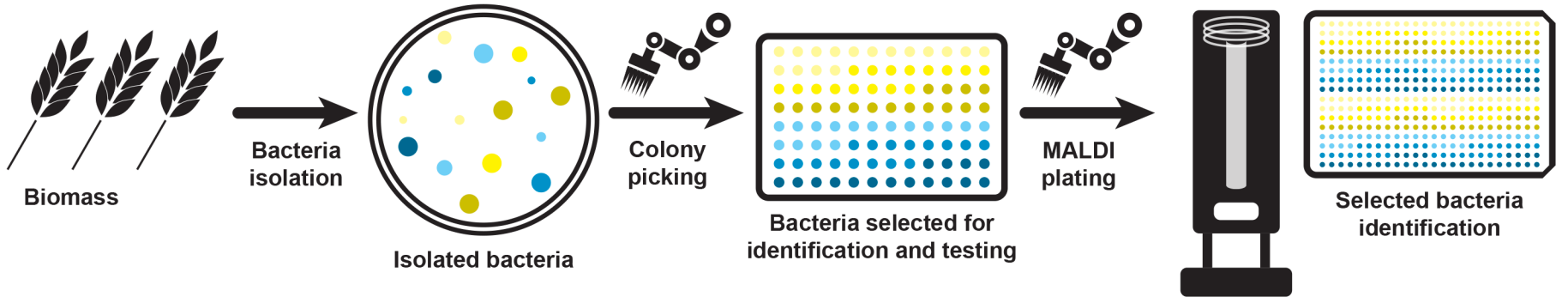


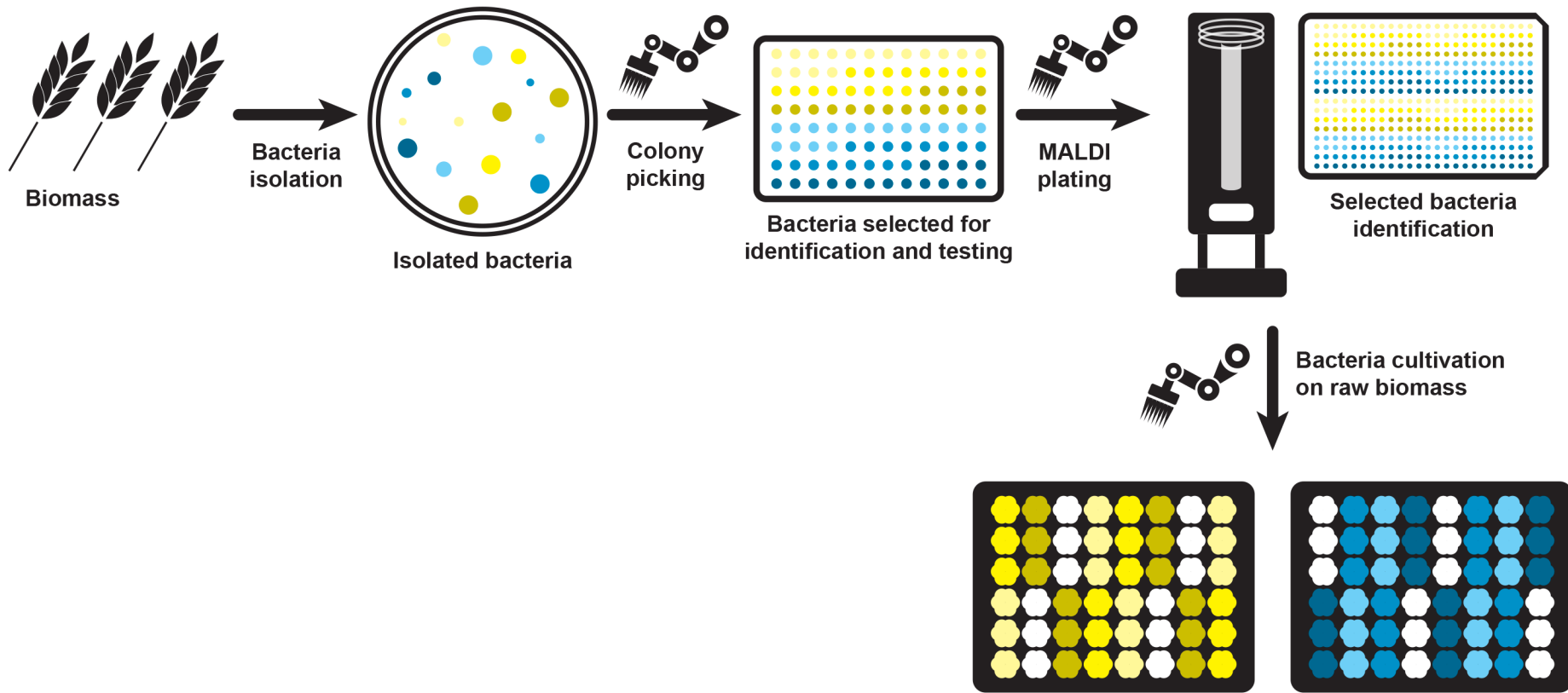
Isolated bacteria

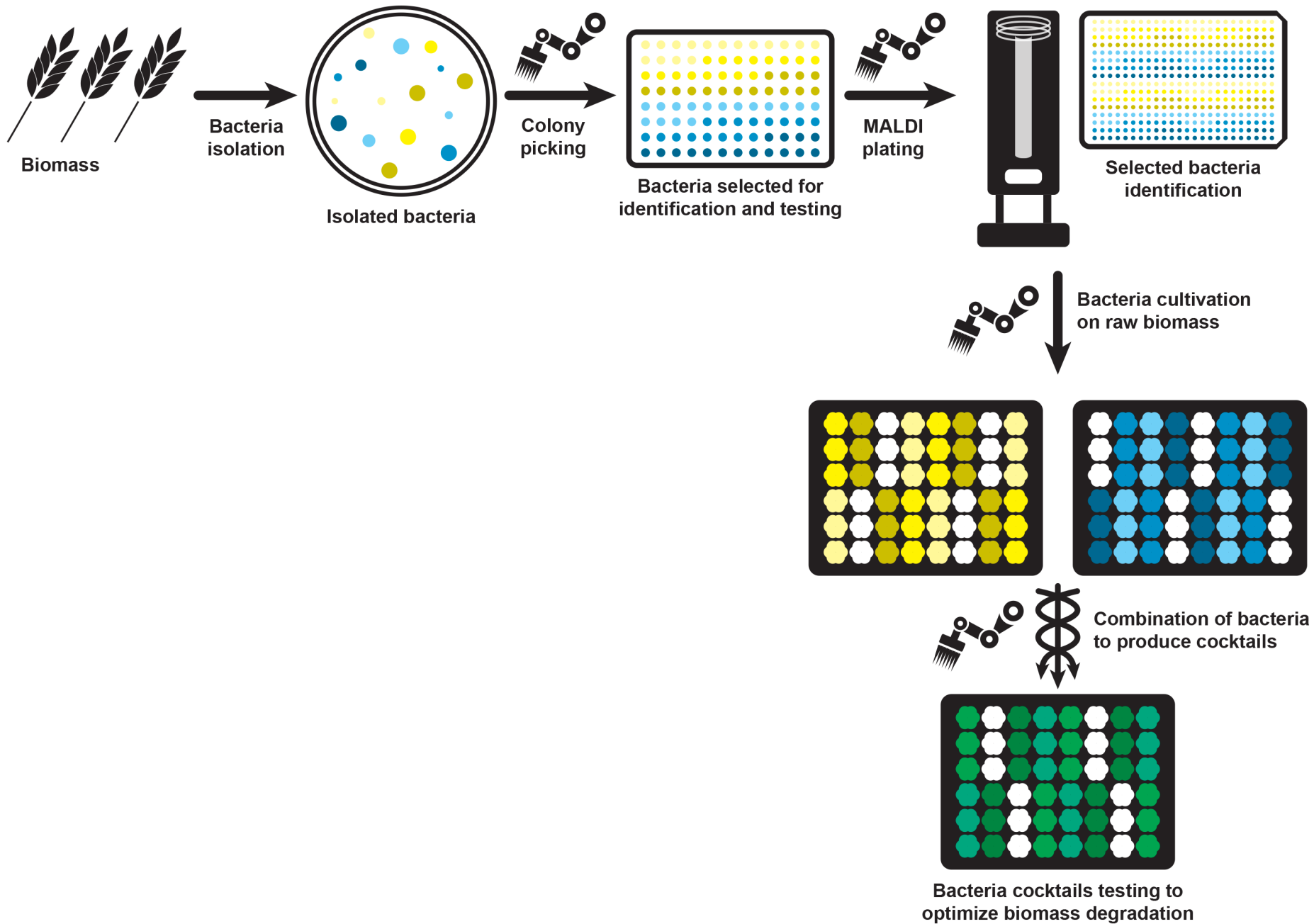


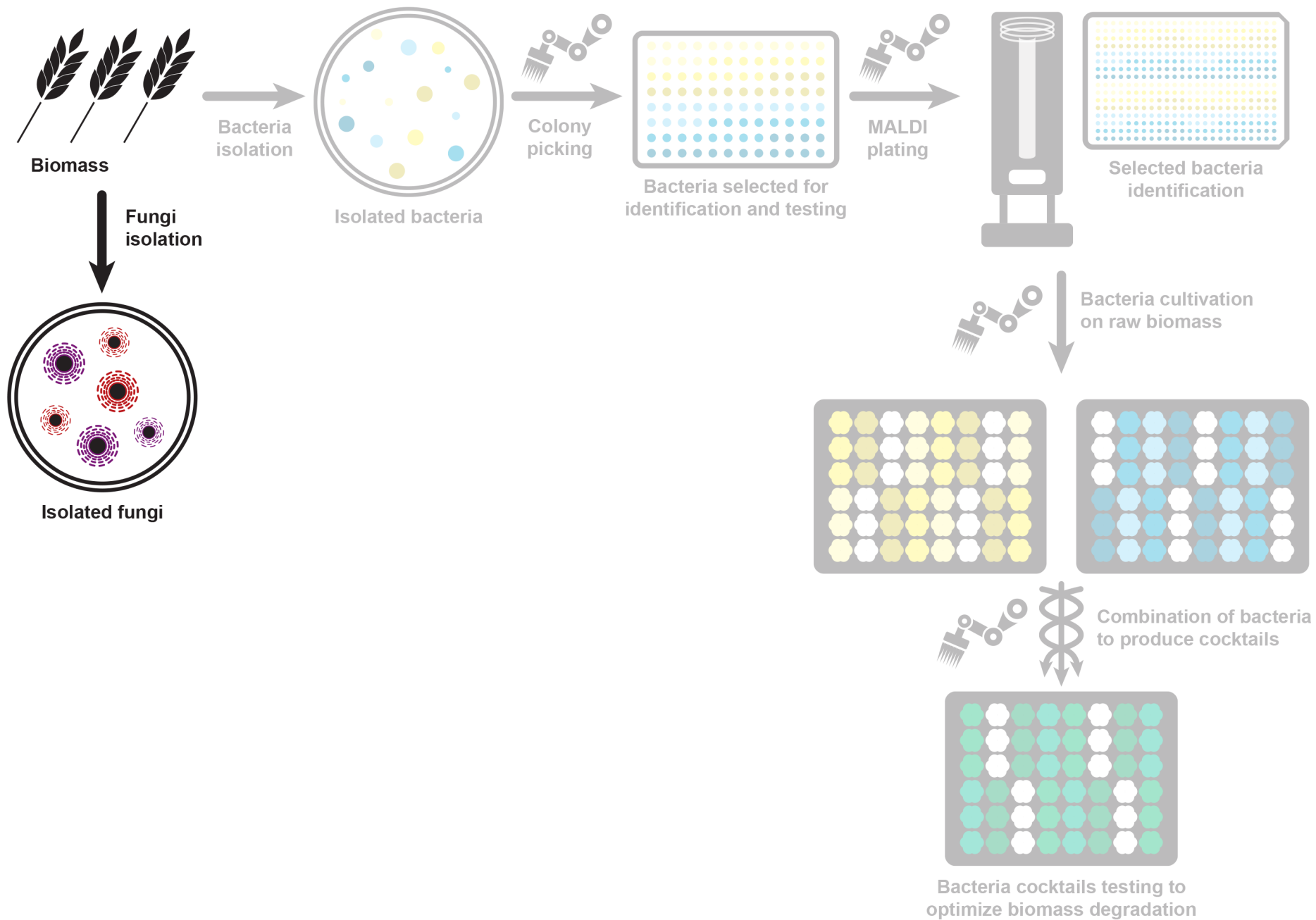
Bacteria
isolation

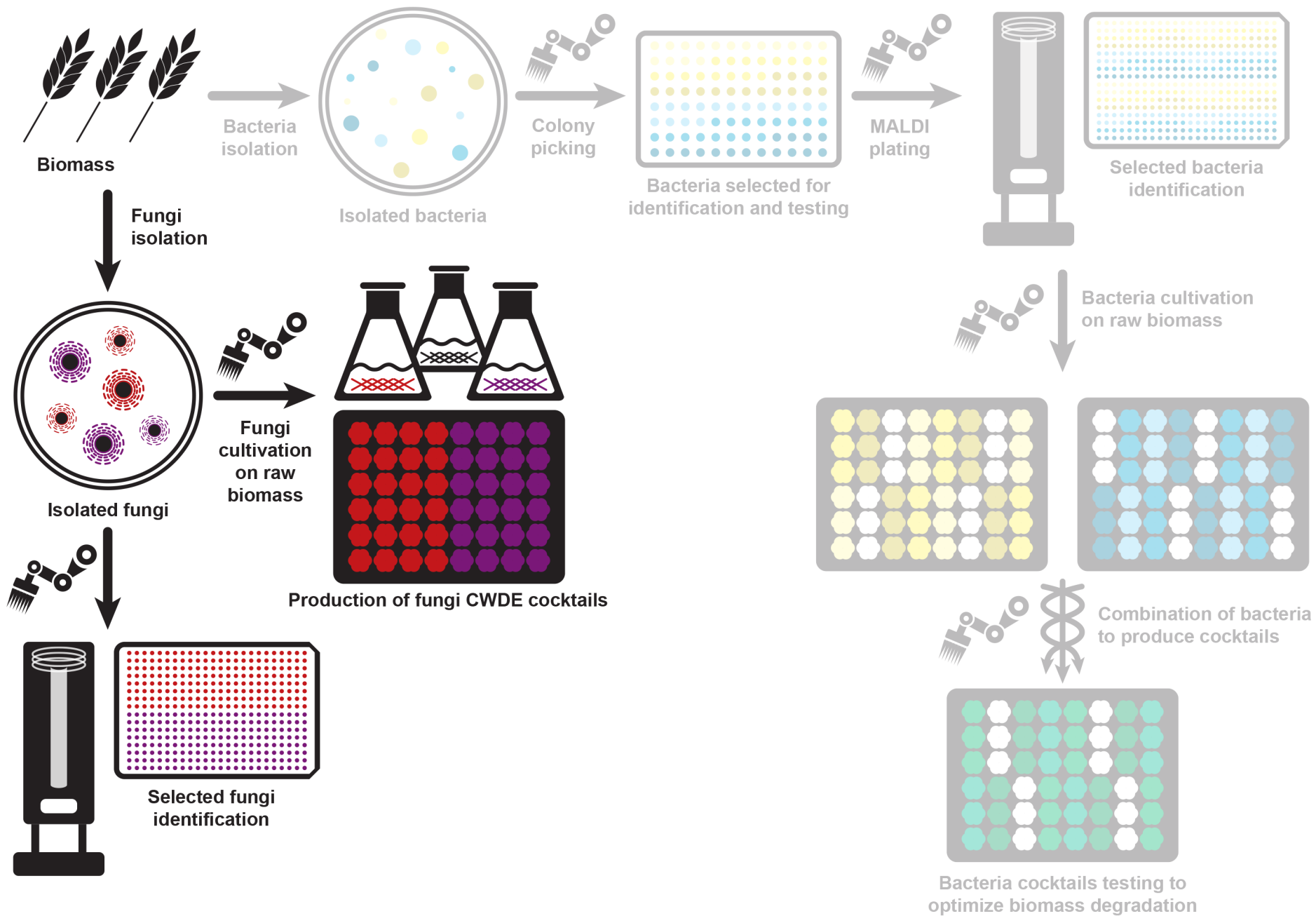


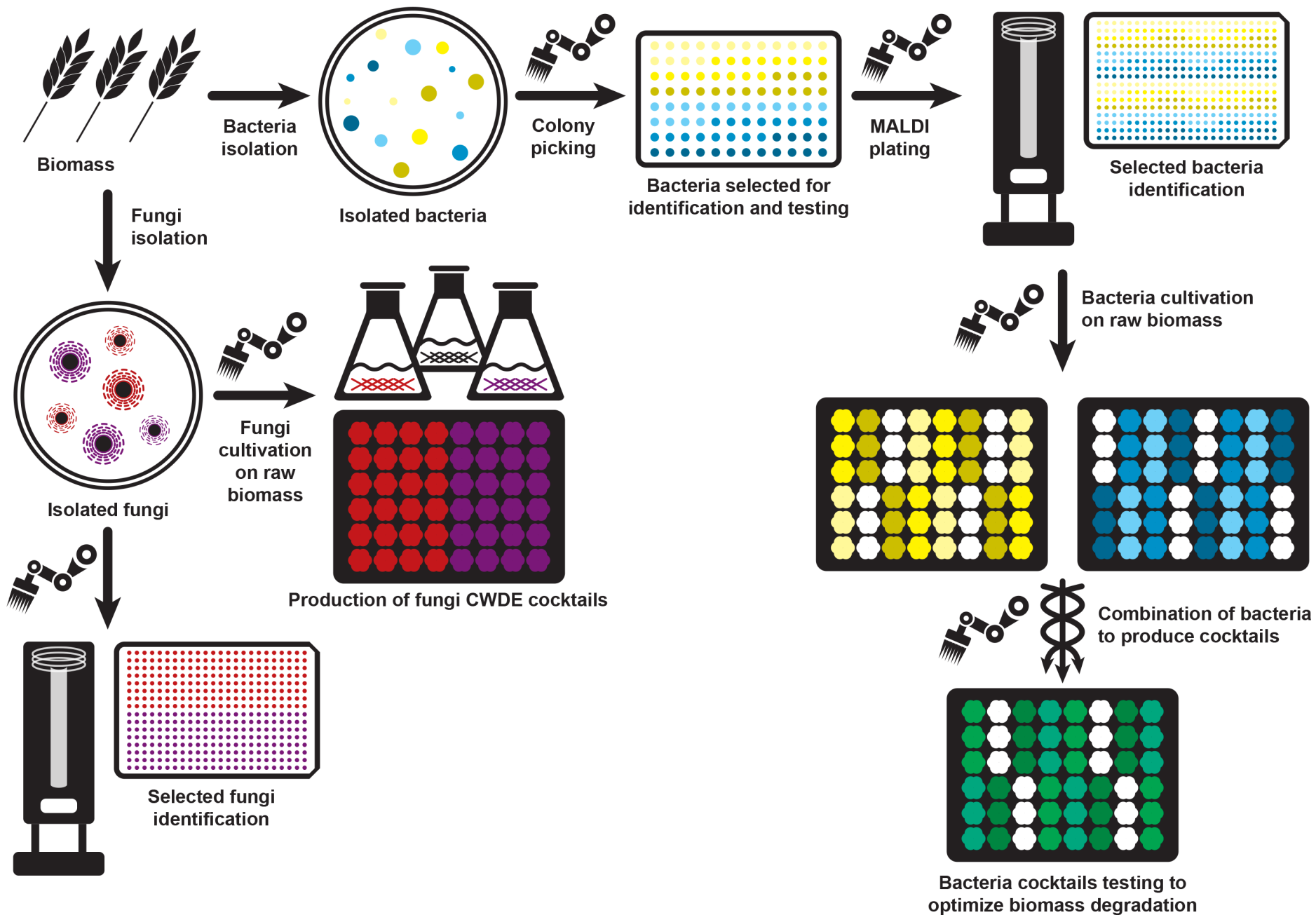


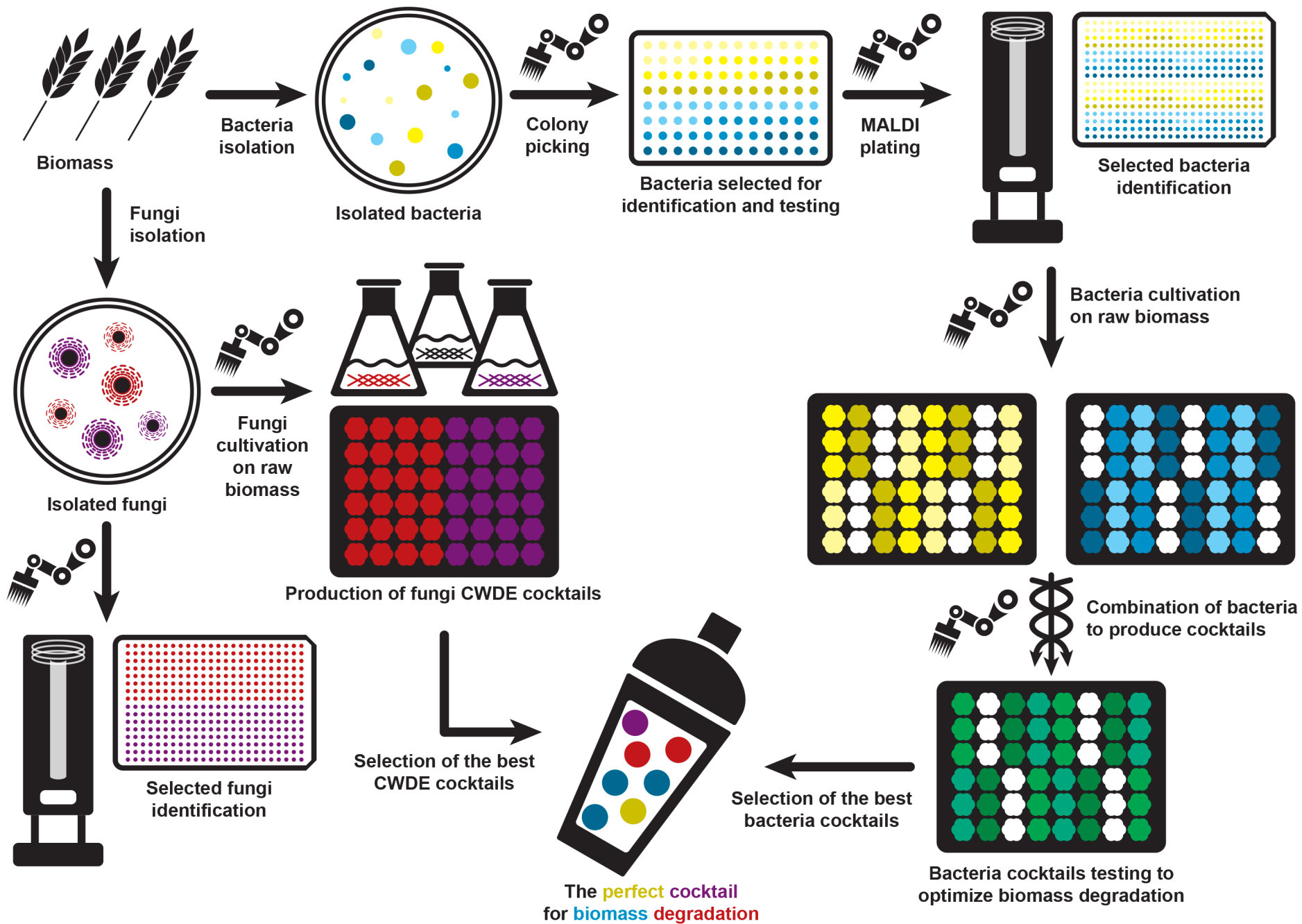


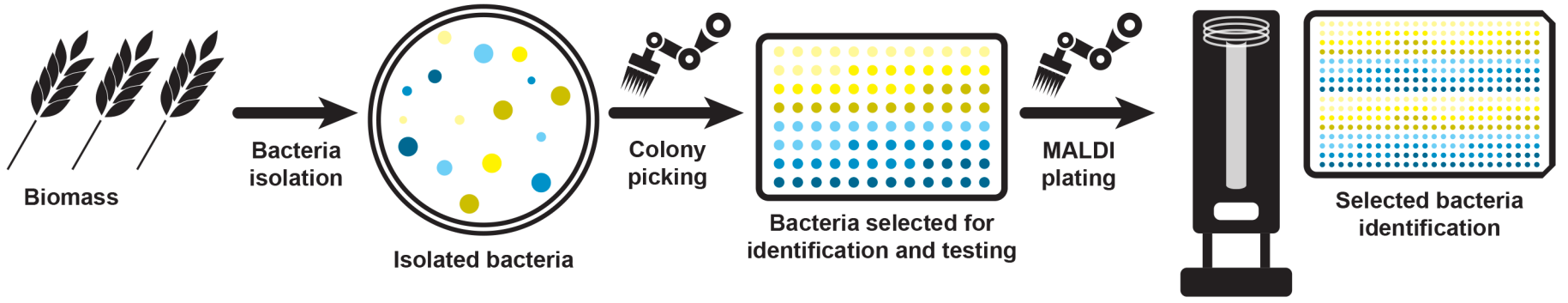








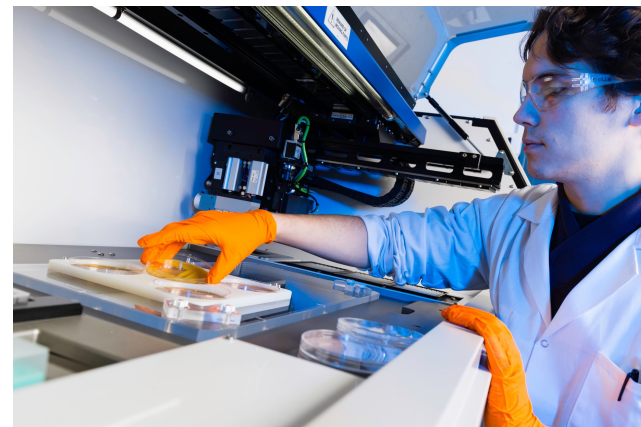
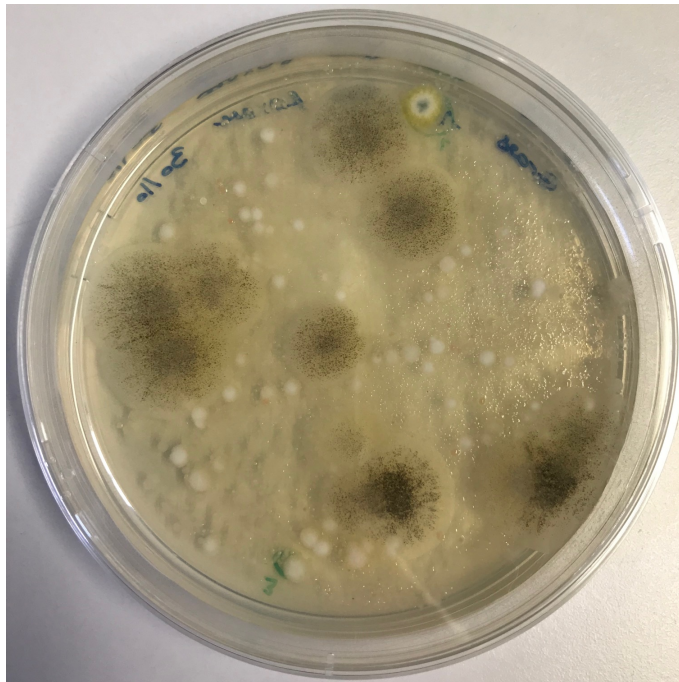




Step 1 – Colony picking

Manual and automated colony recovery: using the QPix 460 :

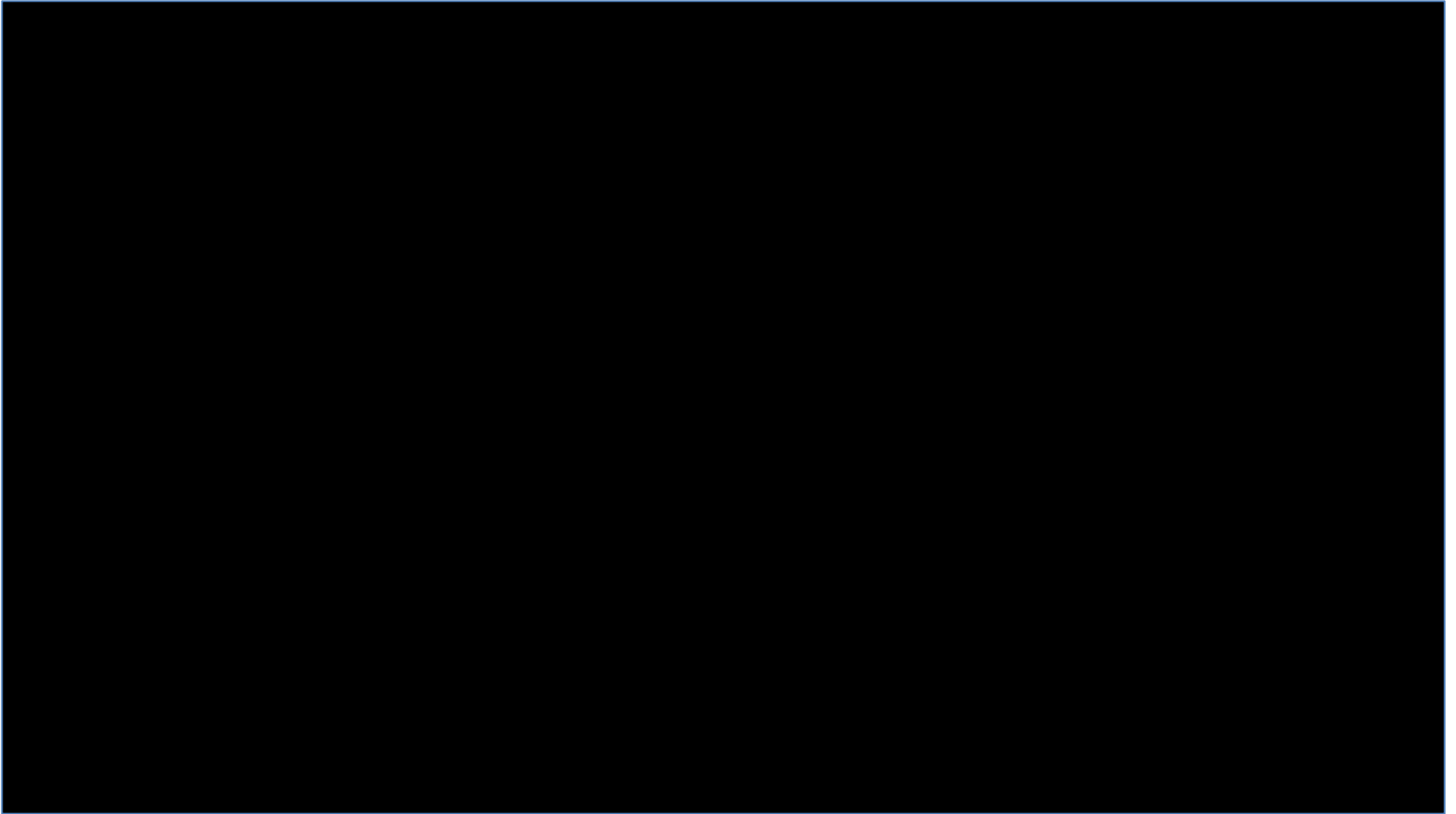
- 96-needle head (approx. 2000 colonies/hour)
- Up to 10 Petri (or 2 Q-Trays) dishes and 40 destination plates per batch
- “Intelligent” optical recognition software



Qpix 460 – Molecular devices

Step 1 – Colony picking

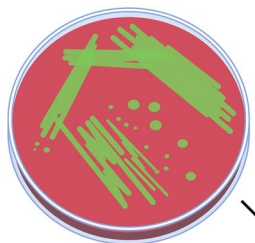
Automated colony picking



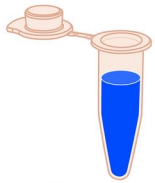
Step 2 – MALDI BioTyper

⇒ First rapid identification using a MALDI-TOF mass spectrometer

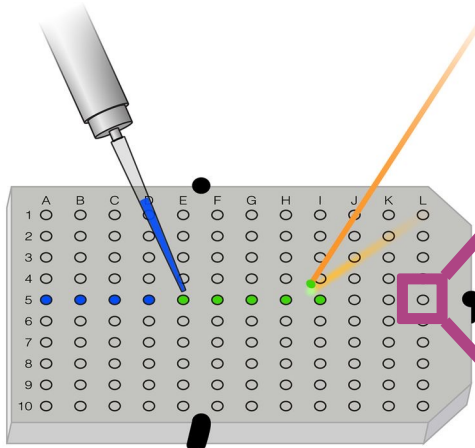
⇒ Confirmation by 16s / ITS RNA sequencing or API galleries if required



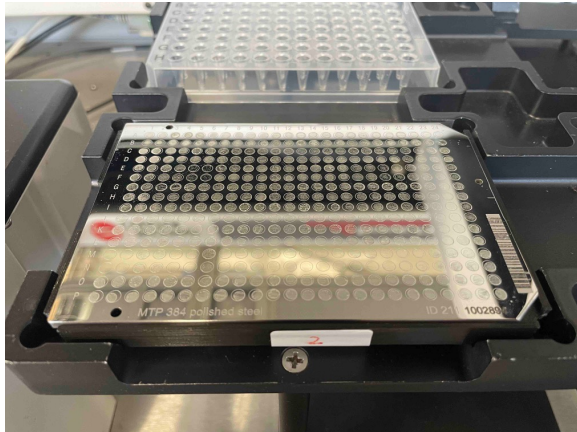
1 Sample culture



2 Matrix



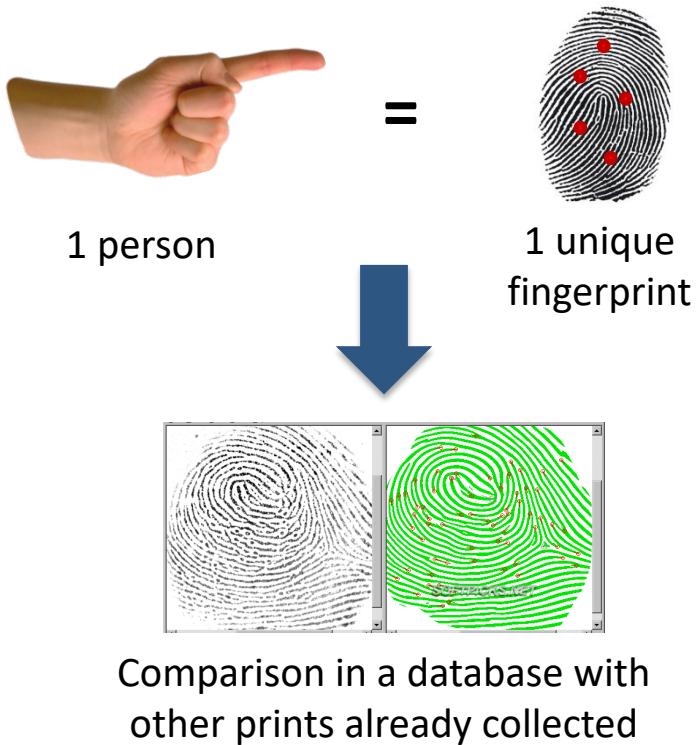
3 MALDI-TOF/MS sample plate



Autoflex Speed – Bruker

Step 2 – MALDI BioTyper

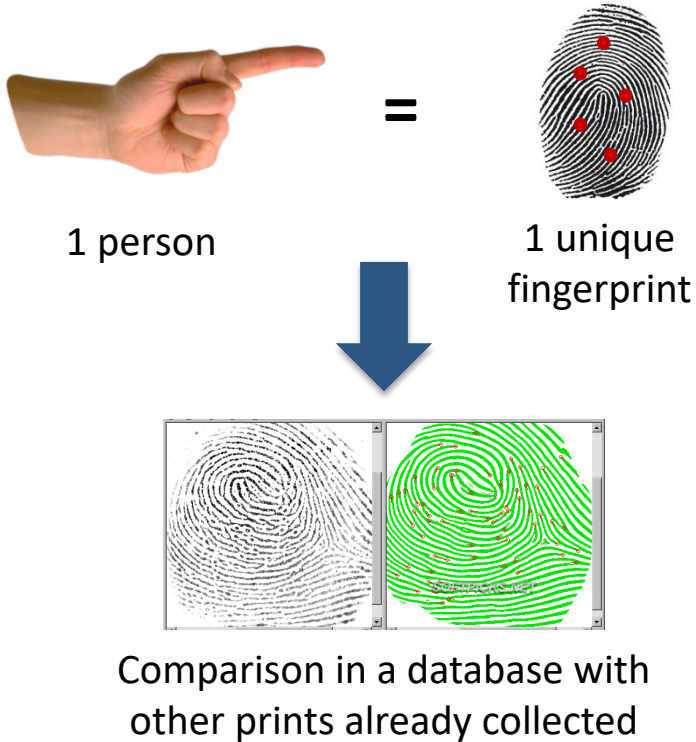
Principle of MBT identification: Digital fingerprinting technique based on the membrane proteins of microorganisms



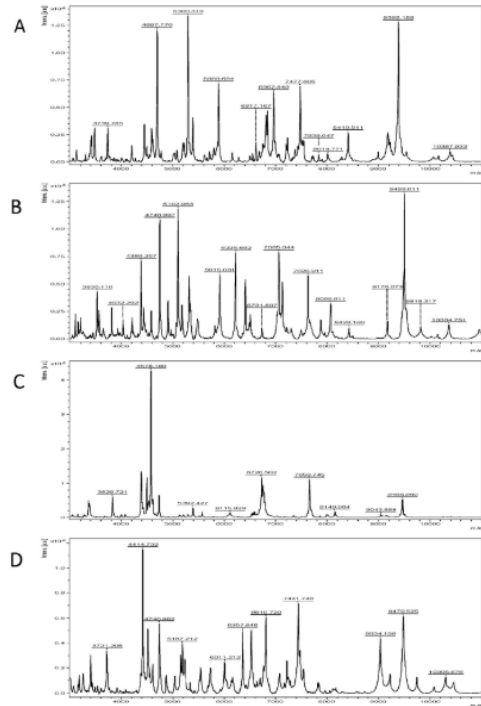
Human fingerprinting

Step 2 – MALDI BioTyper

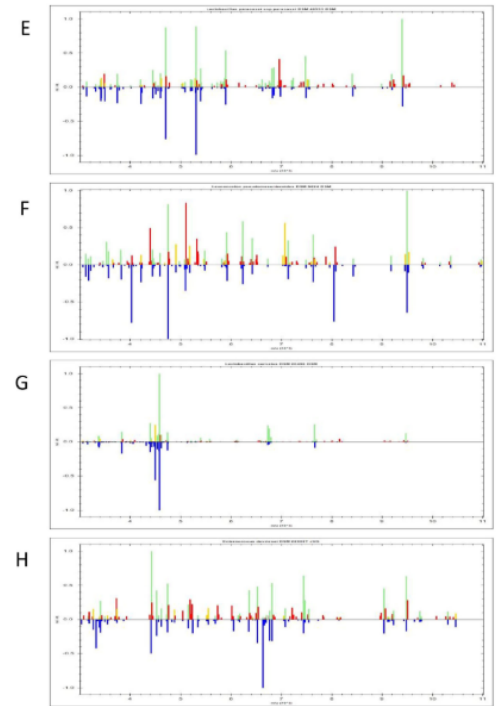
Principle of MBT identification: Digital fingerprinting technique based on the membrane proteins of microorganisms



Human fingerprinting



Mass spectra



Comparison with our in-house + Bruker databases (> 10000 strains)

Molecular fingerprinting

Step 2 – MALDI BioTyper

Principle of MBT identification: Digital fingerprinting technique based on the membrane proteins of microorganisms

Sample Name	Sample ID	Organism (best match)	Score Value	Organism (second-best match)	Score Value
N9 (+++)(C)	8 (standard)	Lactobacillus plantarum	2.40	Lactobacillus plantarum	2.39
N10 (+++)(C)	9 (standard)	Lactobacillus pentosus	2.37	Lactobacillus pentosus	2.29
N11 (+++)(C)	10 (standard)	Lactobacillus plantarum	2.24	Lactobacillus plantarum	2.24
N12 (+++)(C)	11 (standard)	Lactobacillus pentosus	2.07	Lactobacillus pentosus	2.03
N13 (+++)(C)	12 (standard)	Lactobacillus plantarum	2.47	Lactobacillus plantarum	2.45
N14 (+++)(C)	13 (standard)	Lactobacillus brevis	2.13	Lactobacillus brevis	2.13

Meaning of Score Values

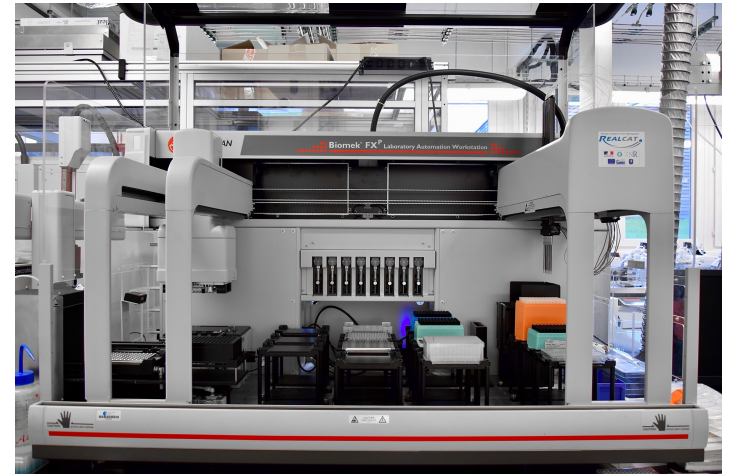
Range	Description	Symbols	Color
2.300 ... 3.000	highly probable species identification	(+++)	green
2.000 ... 2.299	secure genus identification, probable species identification	(++)	green
1.700 ... 1.999	probable genus identification	(+)	yellow
0.000 ... 1.699	not reliable identification	(-)	red

Step 2 – MALDI BioTyper

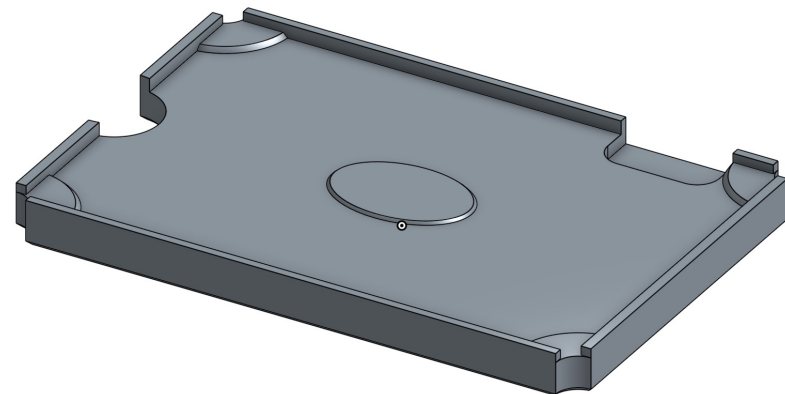
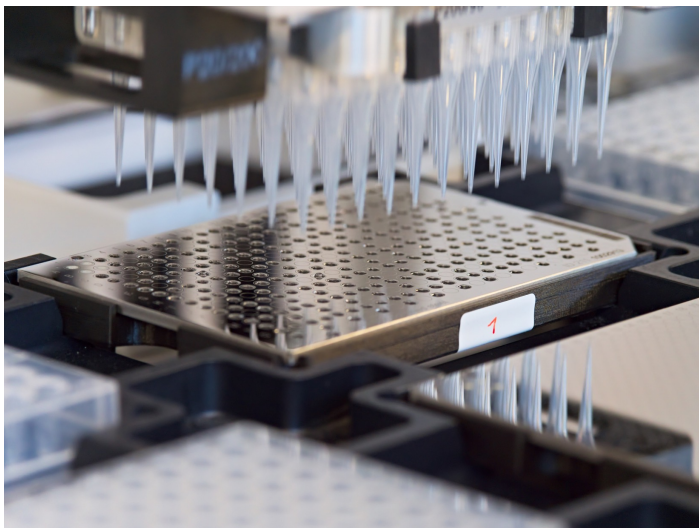
Use of a liquid handling robot : Automated protein extraction and MALDI target preparation

➤ **3 main steps:**

- ⇒ **Cryo-stocks preparation in 30% glycerol solution for bank conservation**
- ⇒ **Cells washing and protein extraction**
- ⇒ **Mixing the protein extracts with the matrix and depositing on the target**



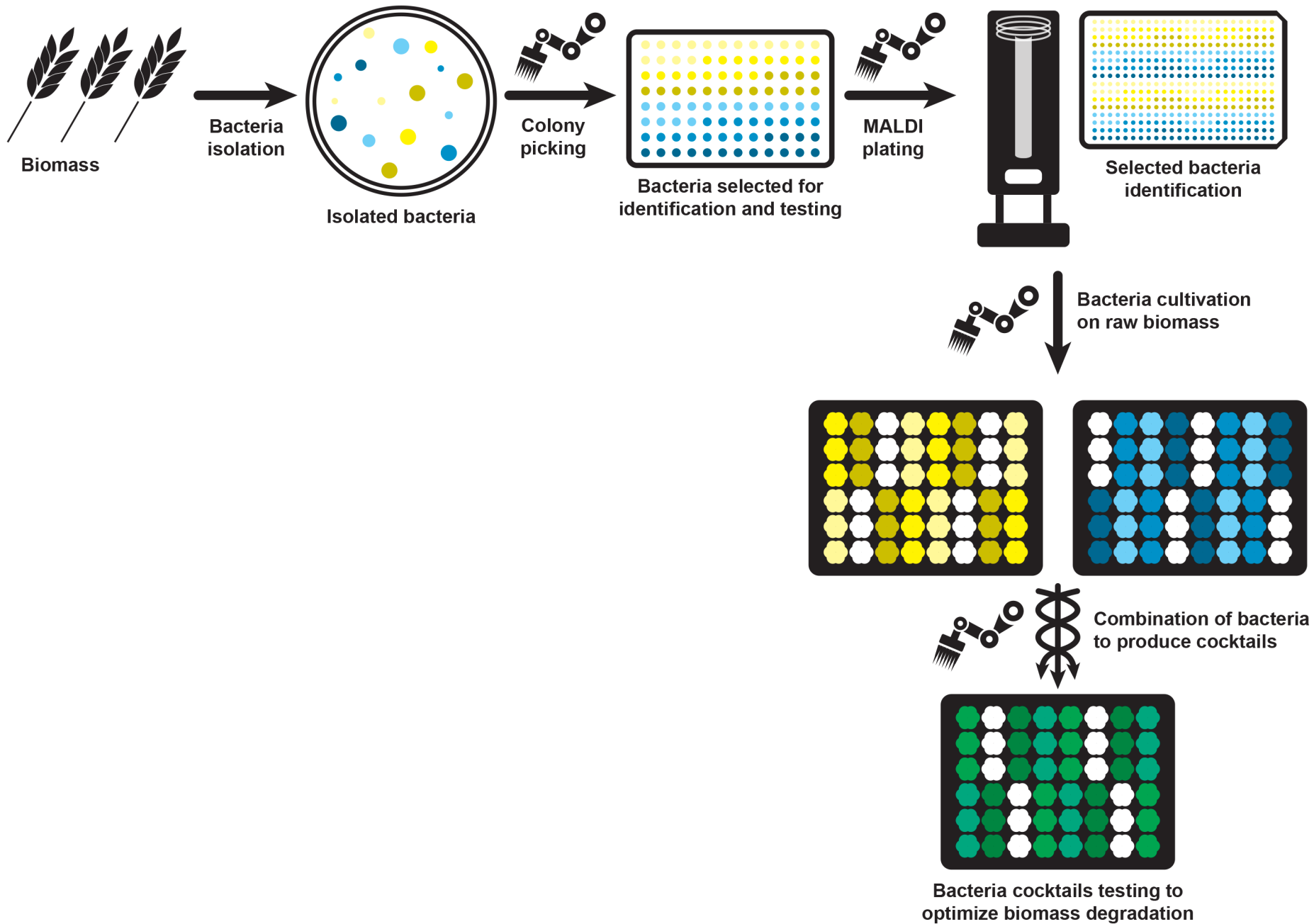
Biomek FXp – Beckman Coulter



Step 2 – MALDI BioTyper

Deposition of the sample/matrix mixture





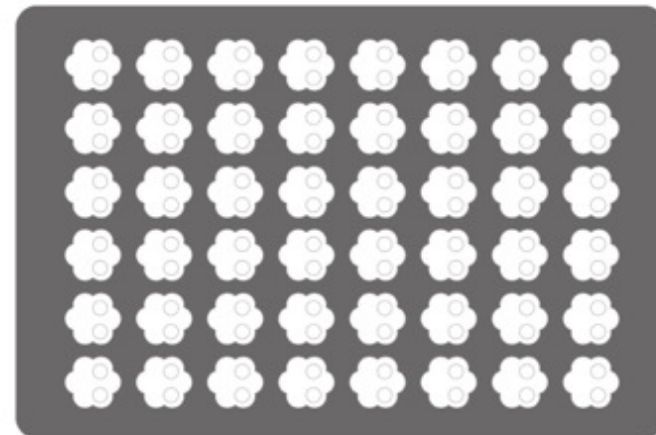
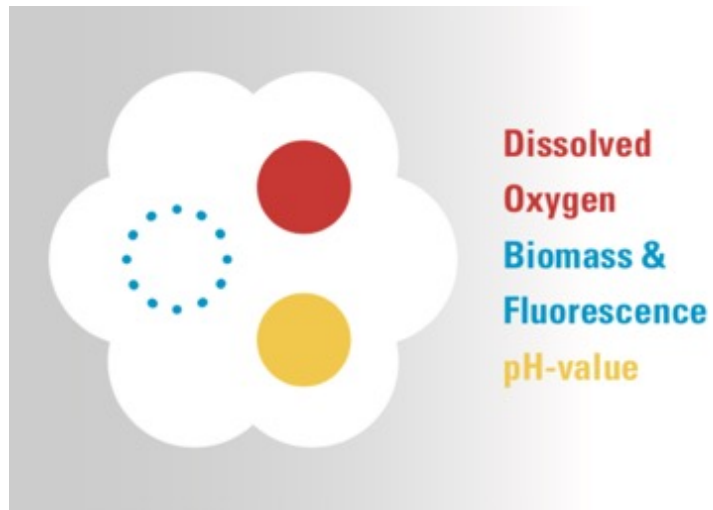
Step 1 – Cultivation and CWDE induction

Use of a high throughput culture device: *The BioLector*

- ⇒ 48 x 1mL parallel fermentations
- ⇒ On-line monitoring and control of pH, pO₂ and biomass
- ⇒ Temperature, humidity and atmosphere control (aerobic and anaerobic fermentation)



BioLector – M2PLabs



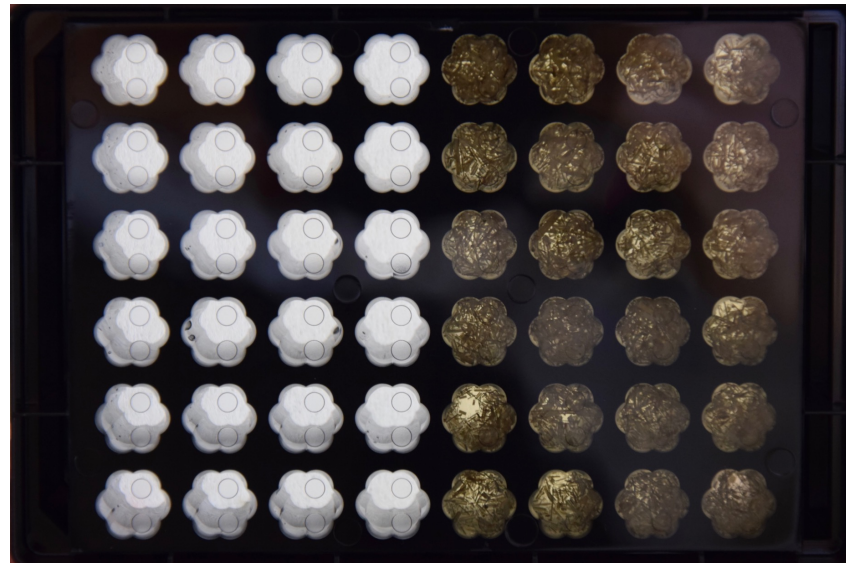
Step 1 – Cultivation and CWDE induction

*Use of a high throughput culture device: **The BioLector***

- ⇒ 48 x 1mL parallel fermentations
- ⇒ On-line monitoring and control of pH, pO₂ and biomass
- ⇒ Temperature, humidity and atmosphere control (aerobic and anaerobic fermentation)
- ⇒ Adaptation to offer the possibility to use raw materials directly (straw, stover, etc.)

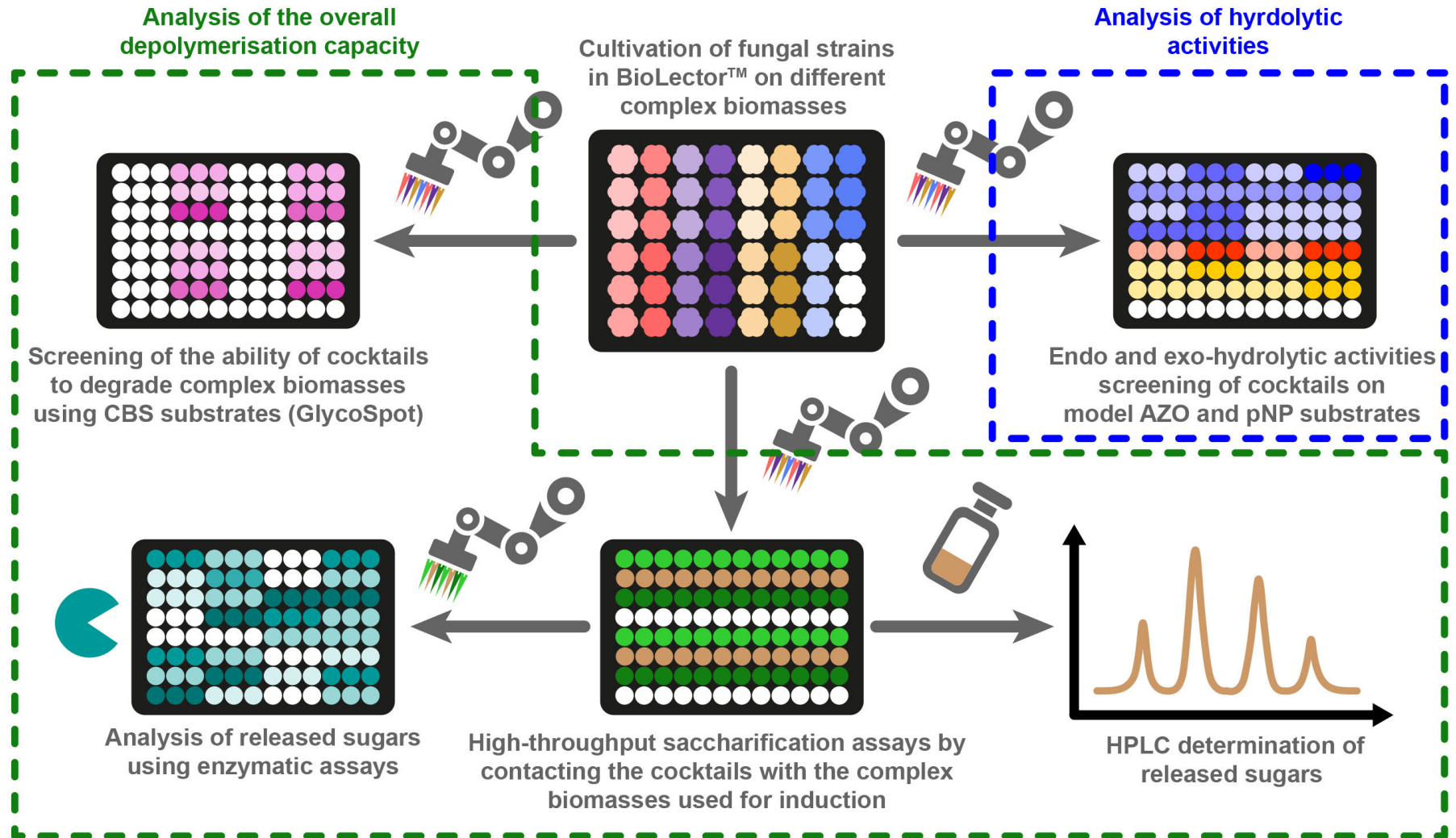


BioLector – M2PLabs



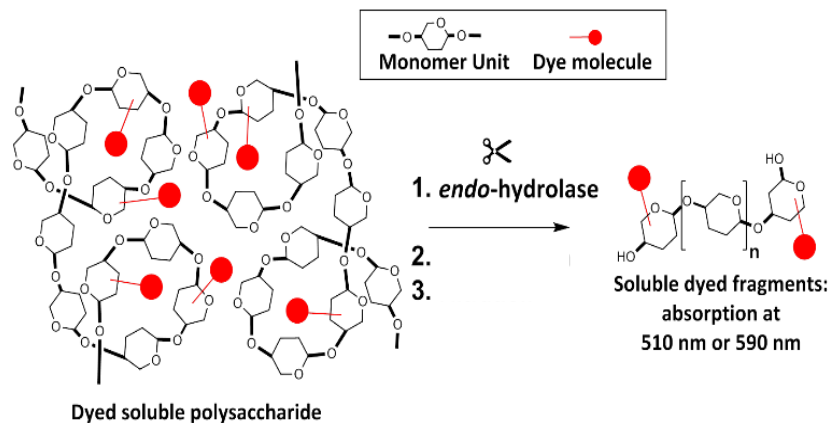
Step 2 – CWDE activity measurement

Extension of the activity measurement panel



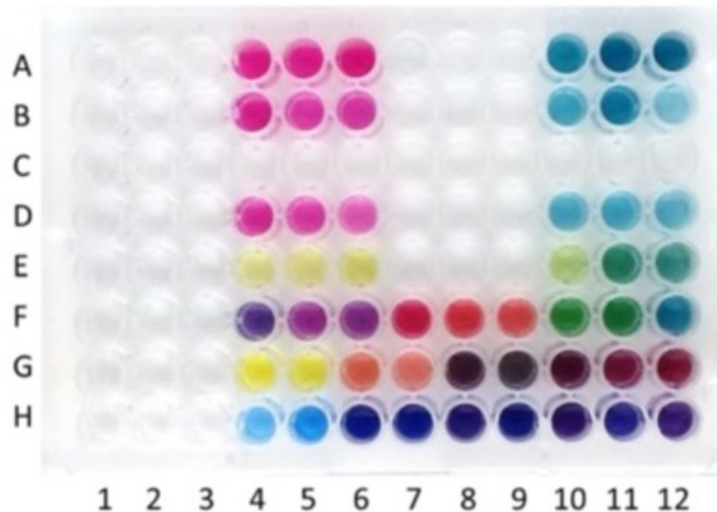
1st Screen – Model substrates for general activities measurement and classification

Substrate	Enzyme family targetted
pNP-glucopyranoside	α -glucosidases (exo)
pNP-xylopyranoside	β -xylosidase (exo)
AZO-xylan	β -xylanases (endo)
AZO-cellulose	β -glucanases (endo)
AZO-rhamnogalacturonan	rhamnogalacturonanases (endo)
AZO-barley glucan	β -glucanases (endo)



2nd Screen - Insoluble Chromogenic Biomass substrates (ICB, Glycospot)

- ⇒ *Substrates in 96-wells plates format*
- ⇒ *4 different dyes (absorption at 404 nm, 517 nm, 595 nm and 630 nm)*
- ⇒ *Complex substrates (custom-made from numerous raw biomass sources)*
- ⇒ *Perfect testing of “real” Biomass*
- ⇒ *Easy removing by filtration*

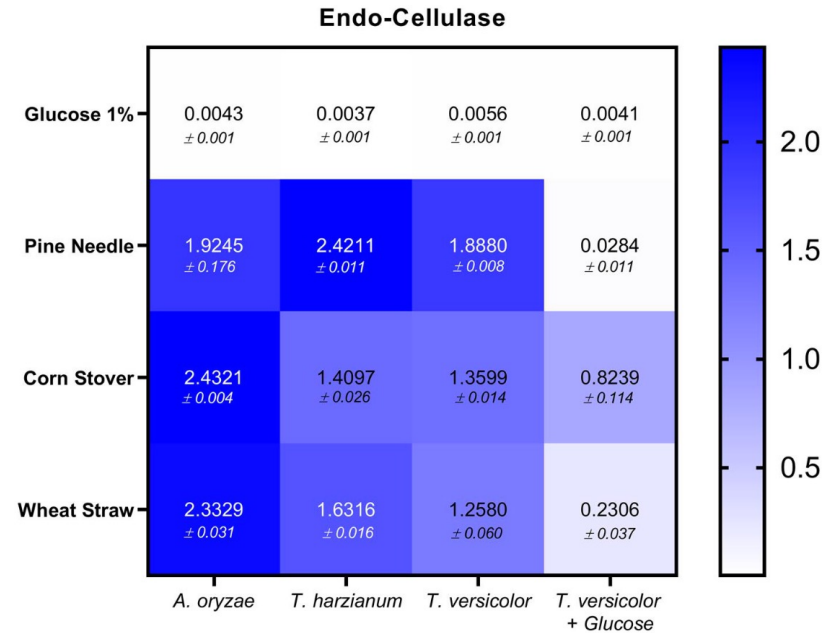
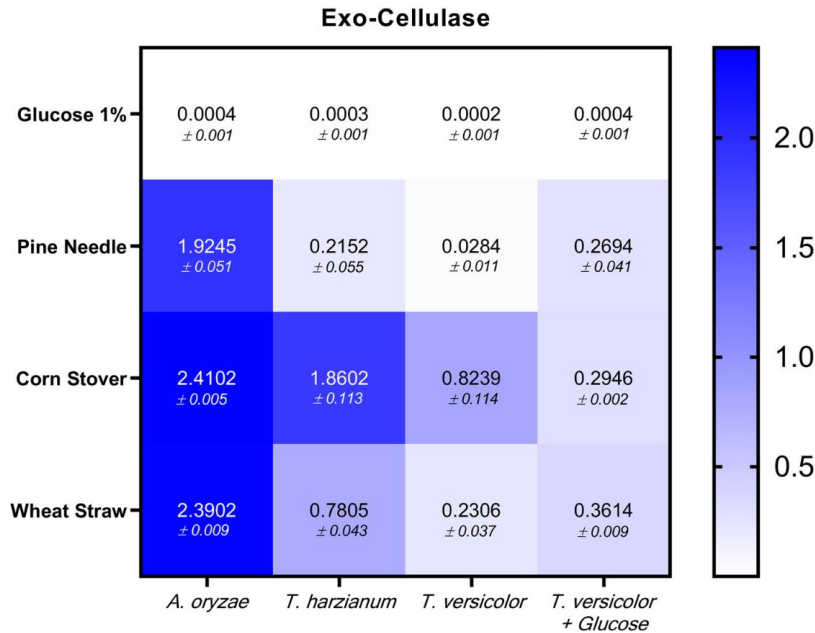


Results obtained from the first screenings

Results for CWDE activity families

⇒ Confirmation via sugar quantification by HPLC

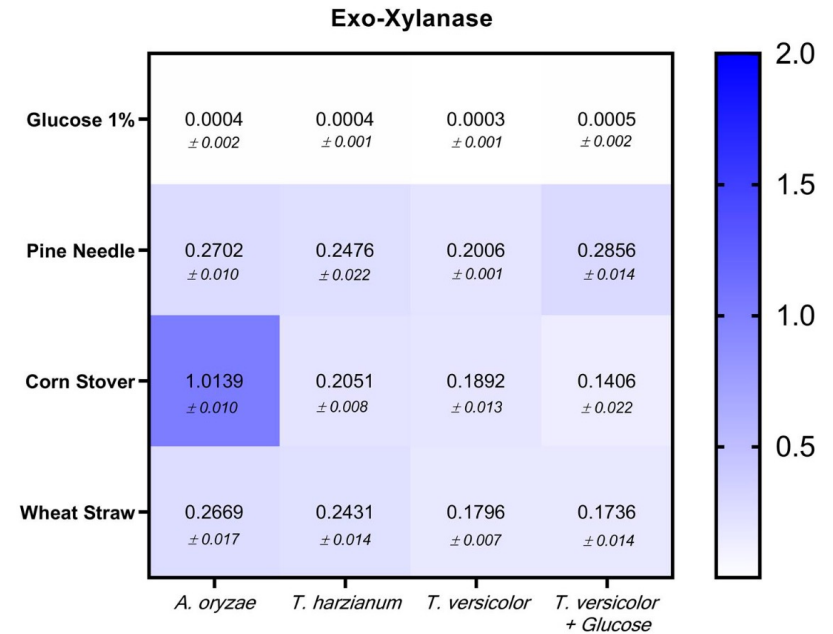
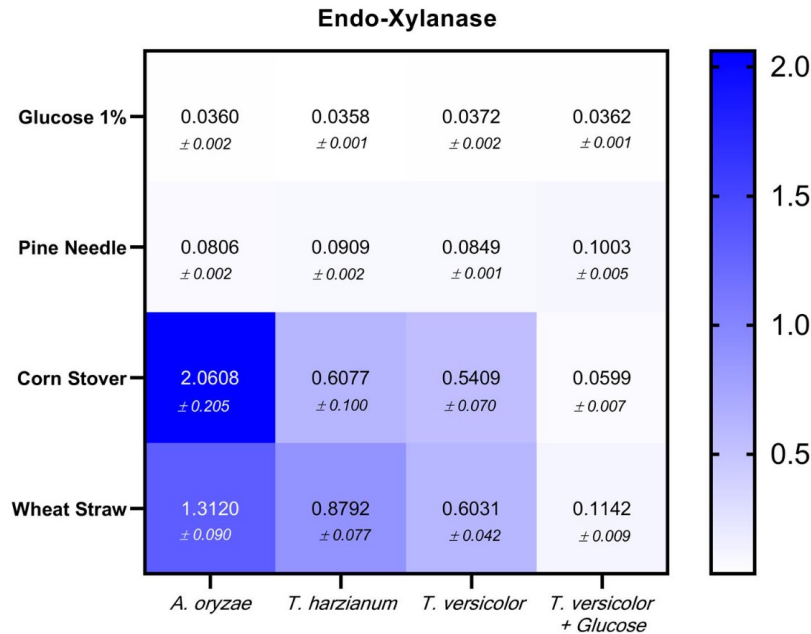
⇒ Cellulases:



Results for CWDE activity families

⇒ Confirmation via sugar quantification by HPLC

⇒ Xylanases:

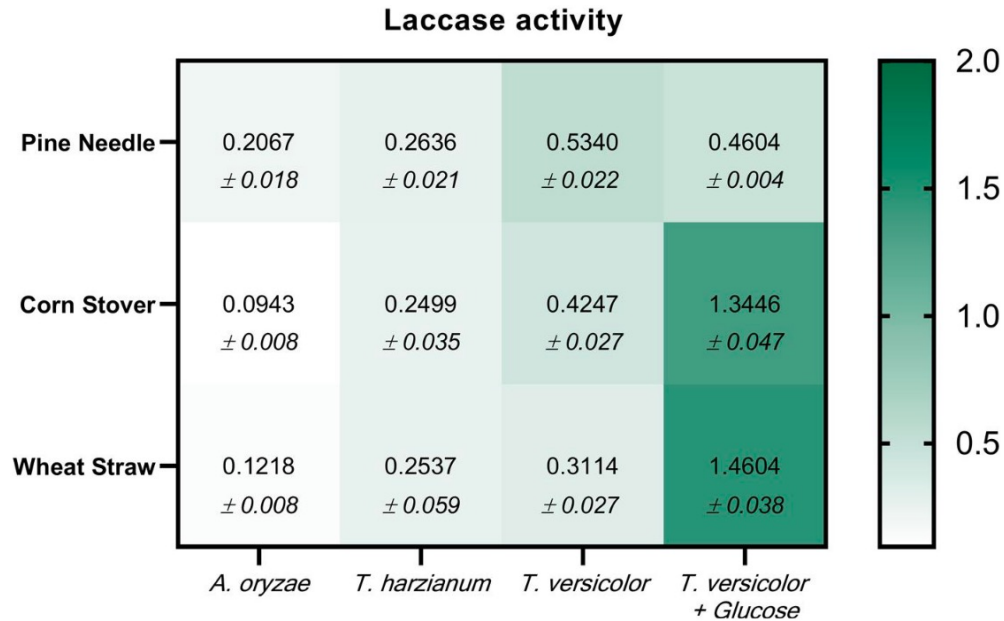


Results obtained from the first screenings

Results for CWDE activity families

⇒ Confirmation via sugar quantification by HPLC

⇒ Laccases:

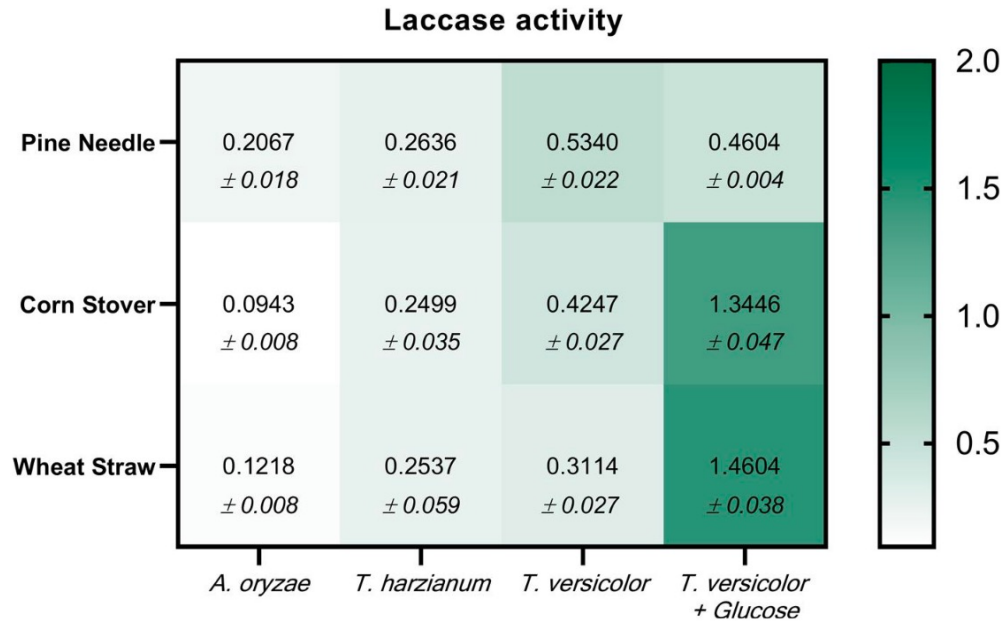


Results obtained from the first screenings

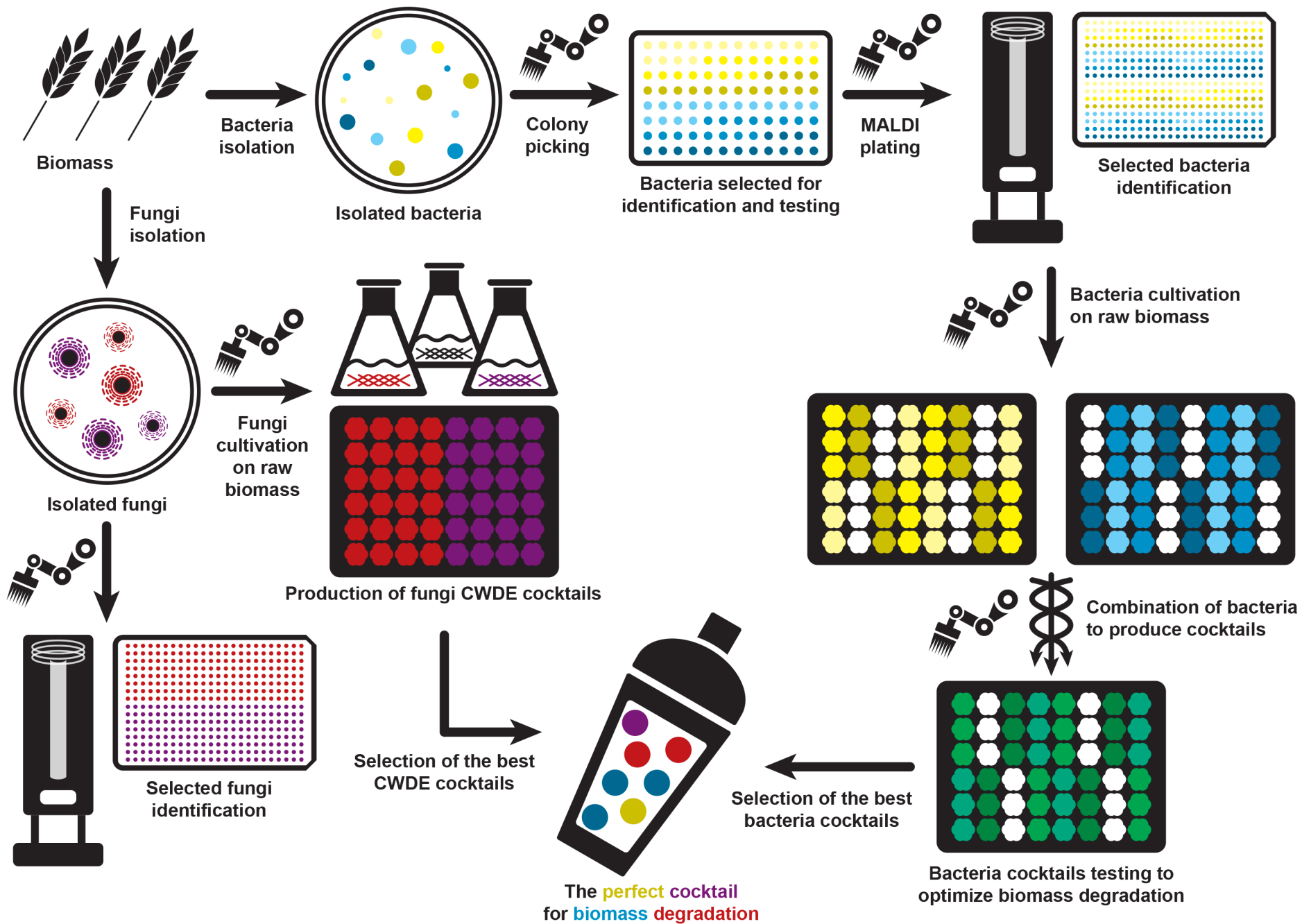
Results for CWDE activity families

⇒ Confirmation via sugar quantification by HPLC

⇒ Laccases:



⇒ Further studies on dark fermentation (short chain organic acid production) using the cocktails as biomass pre-treatment

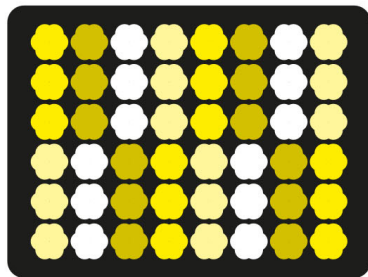
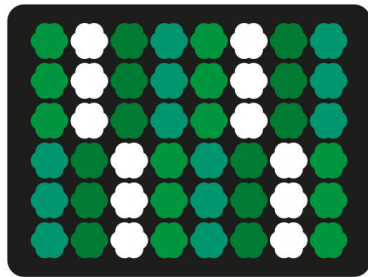


Step 3 – Secreted CWDE mixing

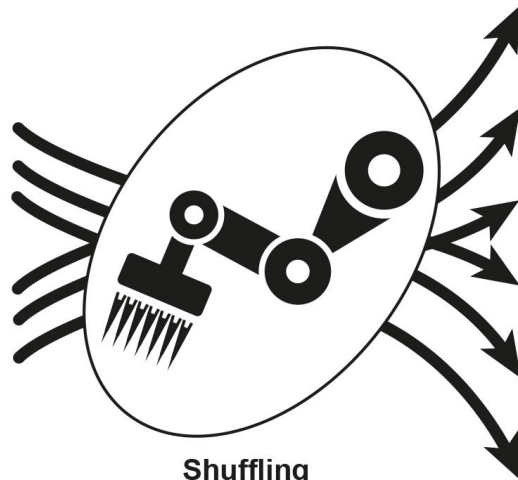
Rational mixing using fully automated workflow using Biomek FXP

⇒ **384 cocktails** combinations created from each batch of **3 strains** / **1 inducing biomass** / **2 tested substrates**

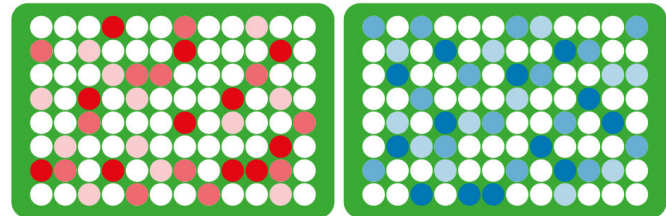
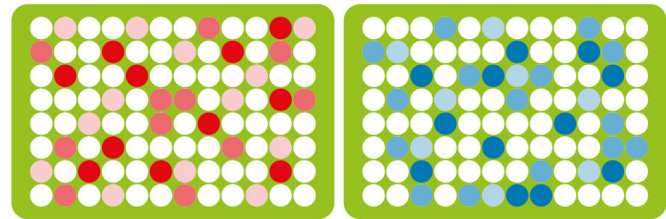
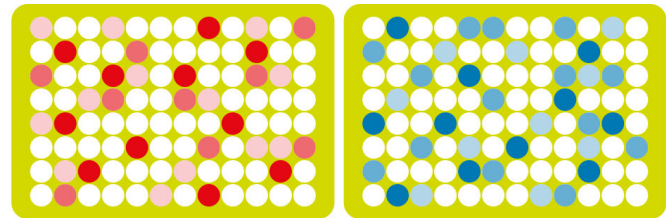
⇒ **New automation compatible DOE program developed**



Submerged fermentation with selected strains (BioLector®)



Shuffling (Biomek FXP)



High-Throughput assays on different substrates from different biomasses (Biomek FXP)

Results for strain identification and enzymes production:

- ⇒ 55 Bacteria isolated / 22 identified - 23 Fungi isolated / 14 identified from 5 biomass sources
- ⇒ Screenings of 3 Fungi + 3 inducing biomass sources for CWDE production
- ⇒ Correlation of enzyme activities detected with complementary analytical methods
- ⇒ Strong correlation between biomass composition and CWDE composition
- ⇒ Successful use of CWDE cocktails to increase the production of short chain organic acids by bacterial dark fermentation, including the implementation of a Plug Flow Reactor (PASS-BIO)



New developments in progress:

- ⇒ Analysis of the protein composition of cocktails (proteomics)
- ⇒ Attempt to correlate the enzyme composition of the cocktails with the screening parameters (biomass, conditions, strains, etc.) by machine learning.
- ⇒ Creation of artificial cocktails with additional heterologous enzymes

Thank you for your attention!

Brings catalysis over lightspeed

REALCAT

The logo for REALCAT features the word "REALCAT" in a bold, sans-serif font. The letters "REAL" are blue, and "CAT" is green. Below the text is a large, stylized graphic consisting of two curved lines, one blue and one green, that sweep upwards from left to right. A small blue sphere is positioned at the end of the green line on the right side.

www.realcat.fr

... au Nord, c'étaient les Corons!