



SFGP  
2019

Nantes

SFGP 2019 – 15-17<sup>th</sup> October 2019

## ***A dual (bio)catalysis approach for the synthesis of 5-HMF from D-glucose***

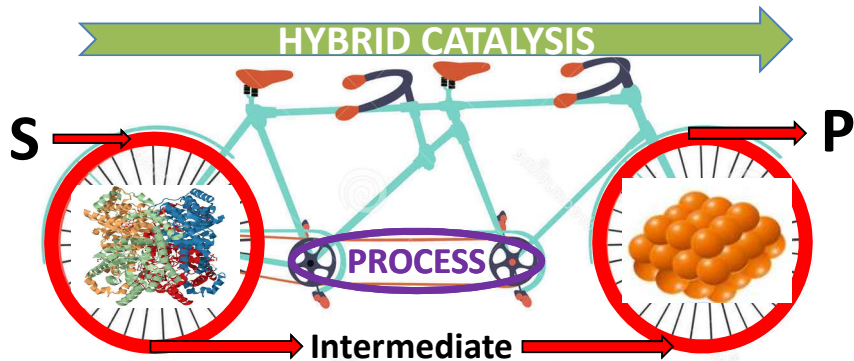
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Egon HEUSON<sup>1</sup>, Mickaël CAPRON<sup>2</sup>, Pascal DHULSTER<sup>1</sup>, Nicolas LOPES FERREIRA<sup>3</sup>,  
Damien DELCROIX<sup>3</sup> and Rénato FROIDEVAUX<sup>1</sup>**

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**Hybrid catalysis  $\Leftrightarrow$  Extension of chemoenzymatic catalysis which is often a multisequential approach**

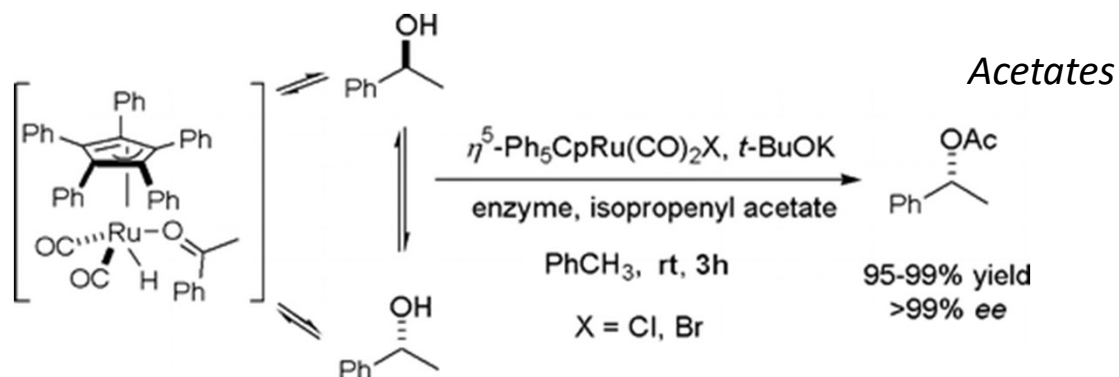
**... green synthesis approach**  
(coexistence of both (bio)catalyst)

**CHALLENGES for combining chemo- and biocatalysis :**

- $\Rightarrow$  Reciprocal poison, need of co-factor/co-enzyme
- $\Rightarrow$  Incompatibility in reaction conditions

# Hybrid catalysis for racemization of secondary alcohols

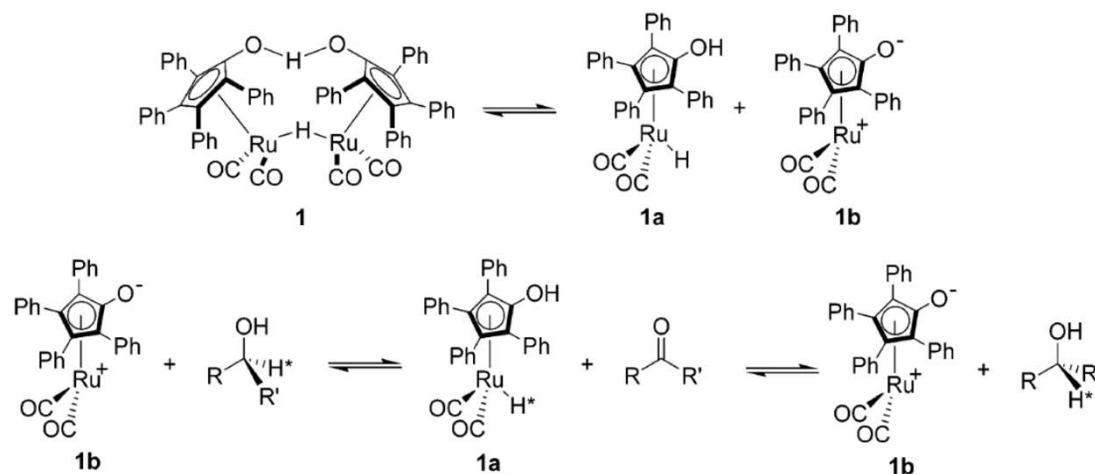
Pentaphenyl  
cyclopentadienyl  
ruthenium complexes



**Homogeneous Enzyme  
and Metalcatalyst**

**Ru complex** = hydrogenation by organometallic H transfer  
**Enzyme (lipase)** = acetylation of only one of the enantiomers

Scheme 7. Proposed Mechanism for the Racemization of Alcohols Catalyzed by **1**



Racemization mechanism for the Ru catalyst **1**

- Proton abstraction  $\Rightarrow$  ruthenium hydride intermediate **1a** + ketone
- Reduction of ketone by ruthenium intermediate **1a**  $\Rightarrow$  racemic alcohol and the ruthenium species **1b**



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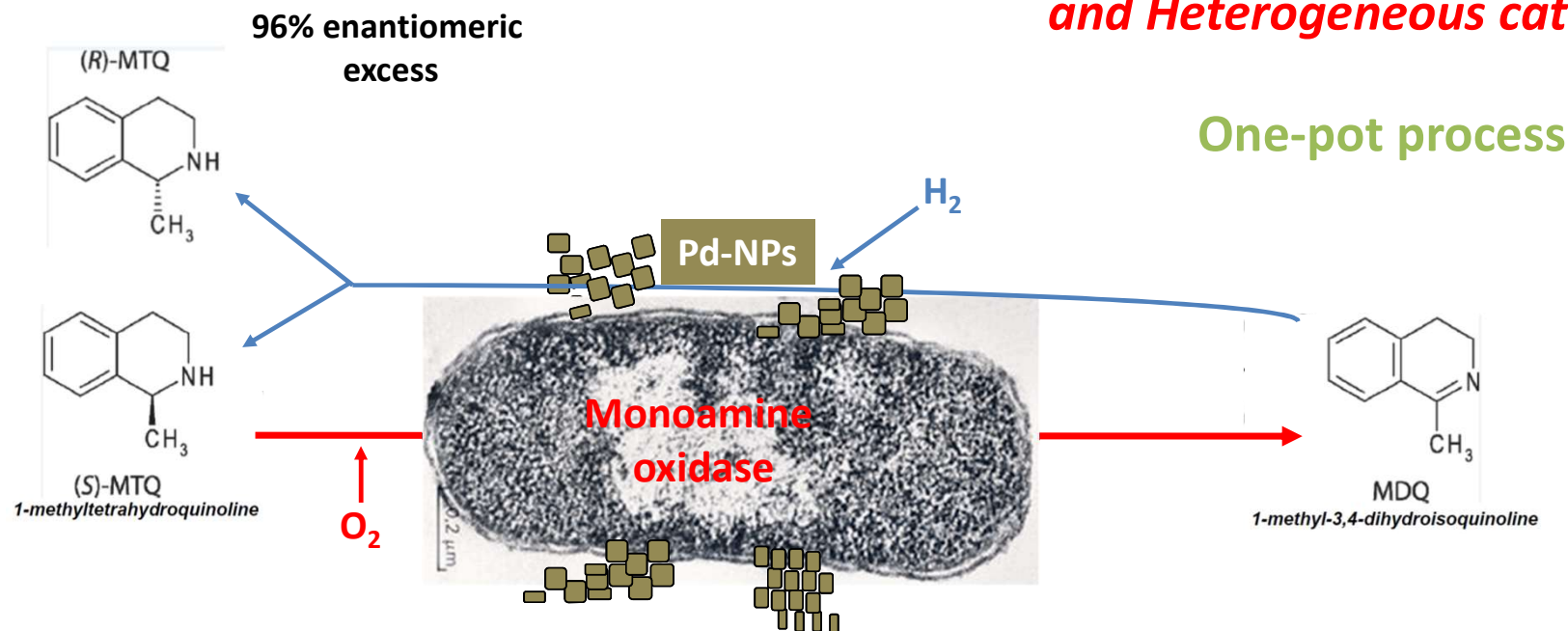
Belén Martín-Matute, Michaela Edin, Krisztián Bogár, F. Betül Kaynak, Jan-E. Bäckvall, *J. Am. Chem. Soc.* 2005 - 127248817-8825

Oscar Pàmies, Jan-E. Bäckvall, *Combination of Enzymes and Metal Catalysts. A Powerful Approach in Asymmetric Catalysis*, *Chem. Rev.* 2003 - 10383247-3262

## Development of hybrid catalysis with physical separation of (bio)catalysts

Enzyme in microorganism

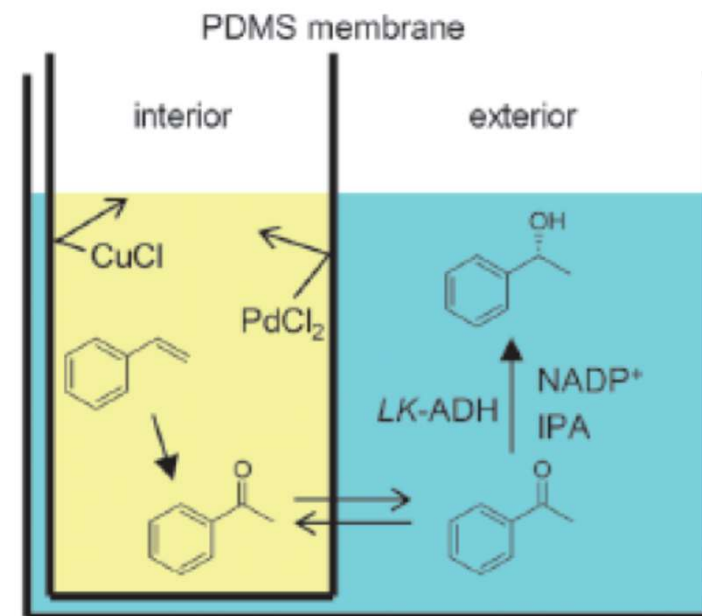
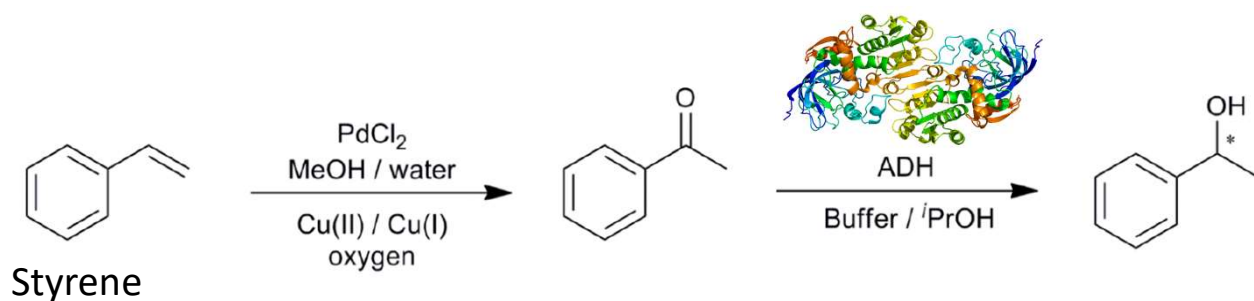
**Homogeneous Enzyme  
and Heterogeneous catalyst**



1. Non selective hydrogenation by Pd-NPs on the cell surface = production of the 2 enantiomers (S)- and (R)-MTQ
2. *E. Coli* cells coated with Pd-NPs expressing a monoamine oxidase (enzyme engineering) which catalyzes the deracemization of (S)-MTQ to the substrate MDQ
3.  $H_2/O_2$  cycles for process implementation with the two catalysts present simultaneously

## Development of hybrid catalysis with physical separation of (bio)catalysts

➔ Membrane separation



Separation of the catalysts by PDMS membrane

1. Reactions conducted in aqueous media = enzyme deactivation by Cu ions
2. Reactions conducted through compartmentalization
3. Polydimethylsiloxane porosity enables diffusion of only the organic substrate and product into the exterior where the enzymatic catalysis takes place

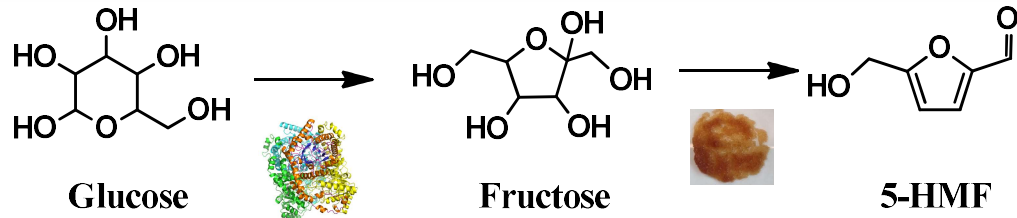
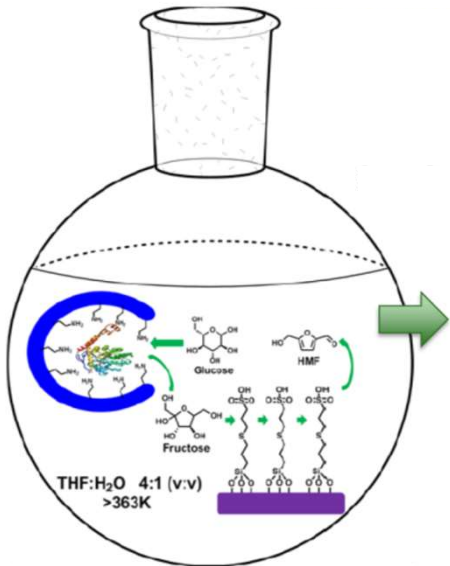
One-pot process

**Homogeneous Enzyme  
and Metalcatalyst**



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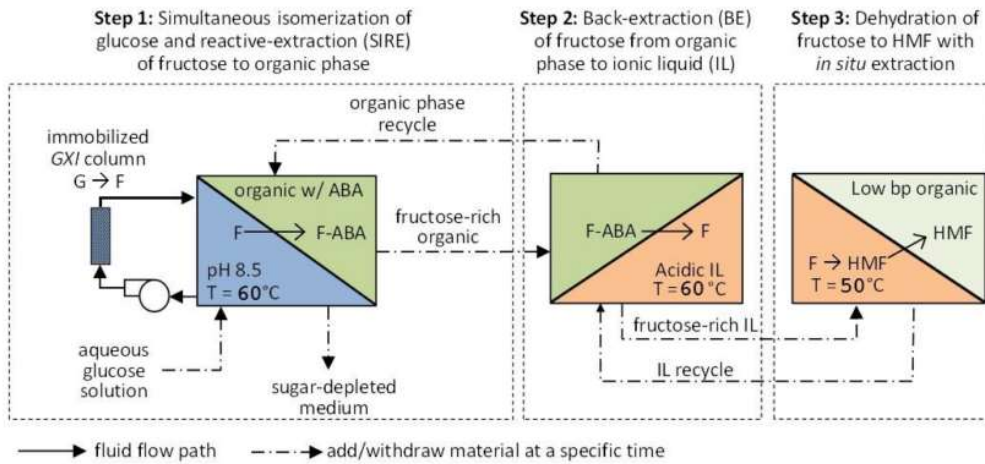
Limits : 1. Formation of humins

2. Inactivation of the isomerase by organic solvent even protected in the silica

3. Product recovery ?

⇒ Physical separation of (bio)catalyst - COMPARTMENTALIZATION

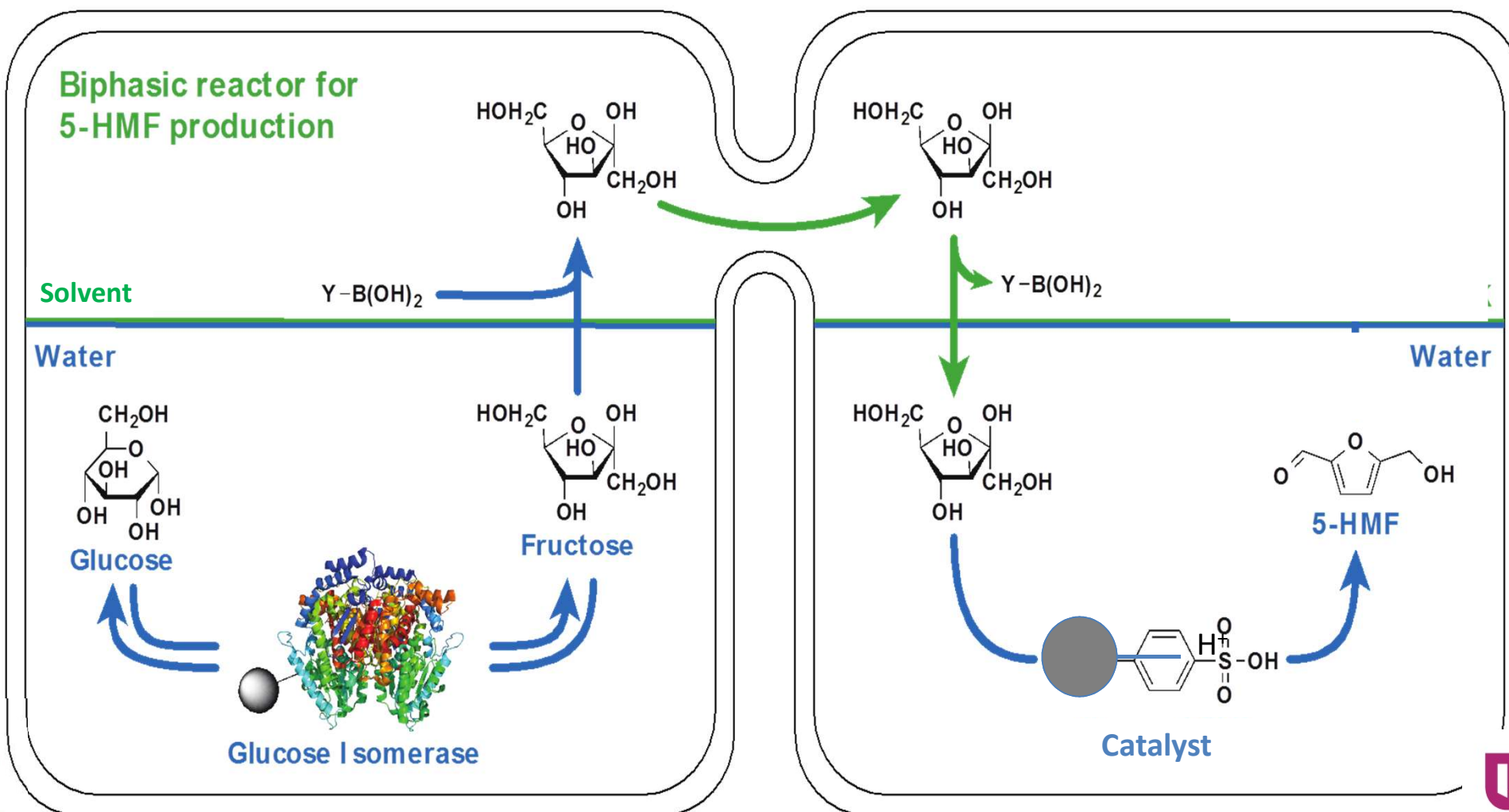
Hua Huang et al., ACS Catalysis, 4(7), 2165-2168



Sequential approach

⇒ SIMULTANEOUS approach

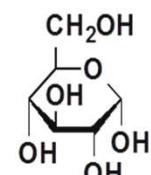
# OUR GOAL...HYBRID CATALYSIS IN DESIGNED « H » REACTOR



## Biphasic reactor for 5-HMF production

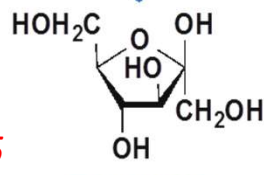
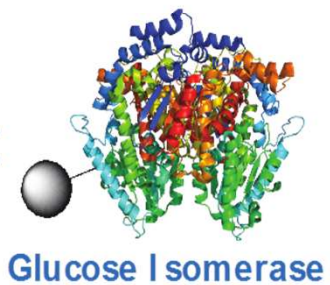
Solvent

Water



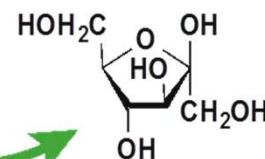
Glucose

*70°C, pH8.5*

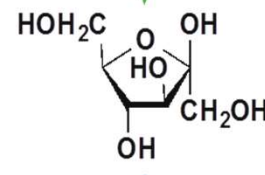


Fructose

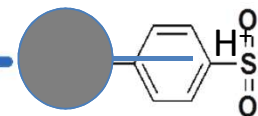
Y-B(OH)<sub>2</sub>



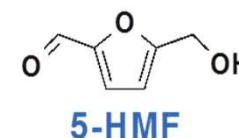
Y-B(OH)<sub>2</sub>



*80°C, pH3, H<sup>+</sup>/Fru 3/1*



*Dowex monosphere®650C*



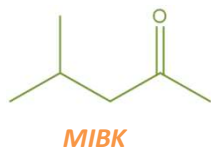
5-HMF

Water



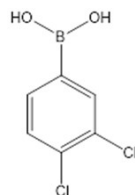
## Extraction of fructose

## Development of liquid membrane



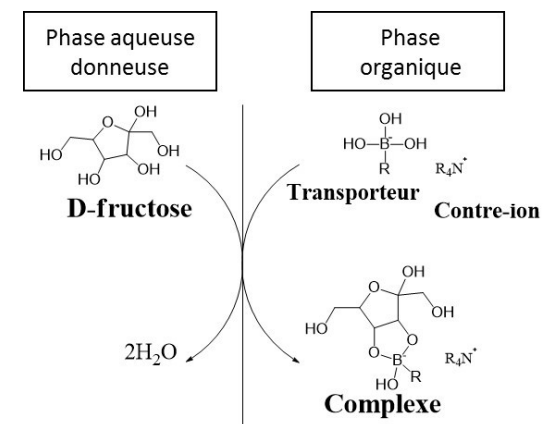
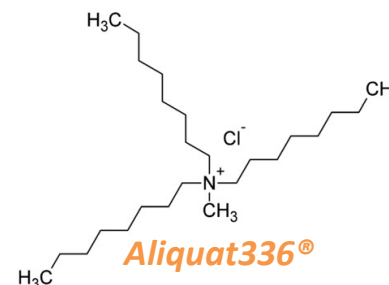
- Solvent

4-methylpentan-2-one (MIBK)



- Carrier (T)

23-DCPBA	35-DCPBA	3-TFMPBA	2-NNPBA
<b>34-DCPBA</b>	3-NPBA	4-B1nPBA	32-carboPBA
35-BTFMPBA	24-DCPBA	4-M21HPBA	
4-TFMeOPBA	2-TFMPBA	2-T5PBA	



Glucose and fructose adducts formed with boronic acids

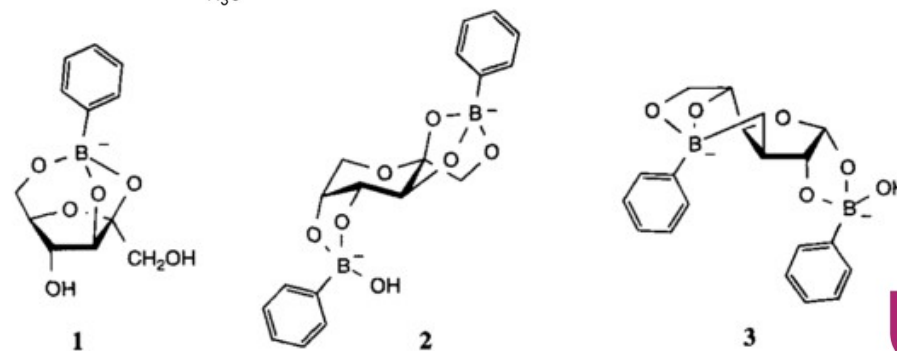
Affinity  $(34\text{-DCPBA})_{\text{Fru}} \gg (34\text{-DCPBA})_{\text{Glc}}$

This complex induces the negative ion on the bore

Ionic interaction with the counterion **Aliquat336®**

Formation of a lipophilic complex

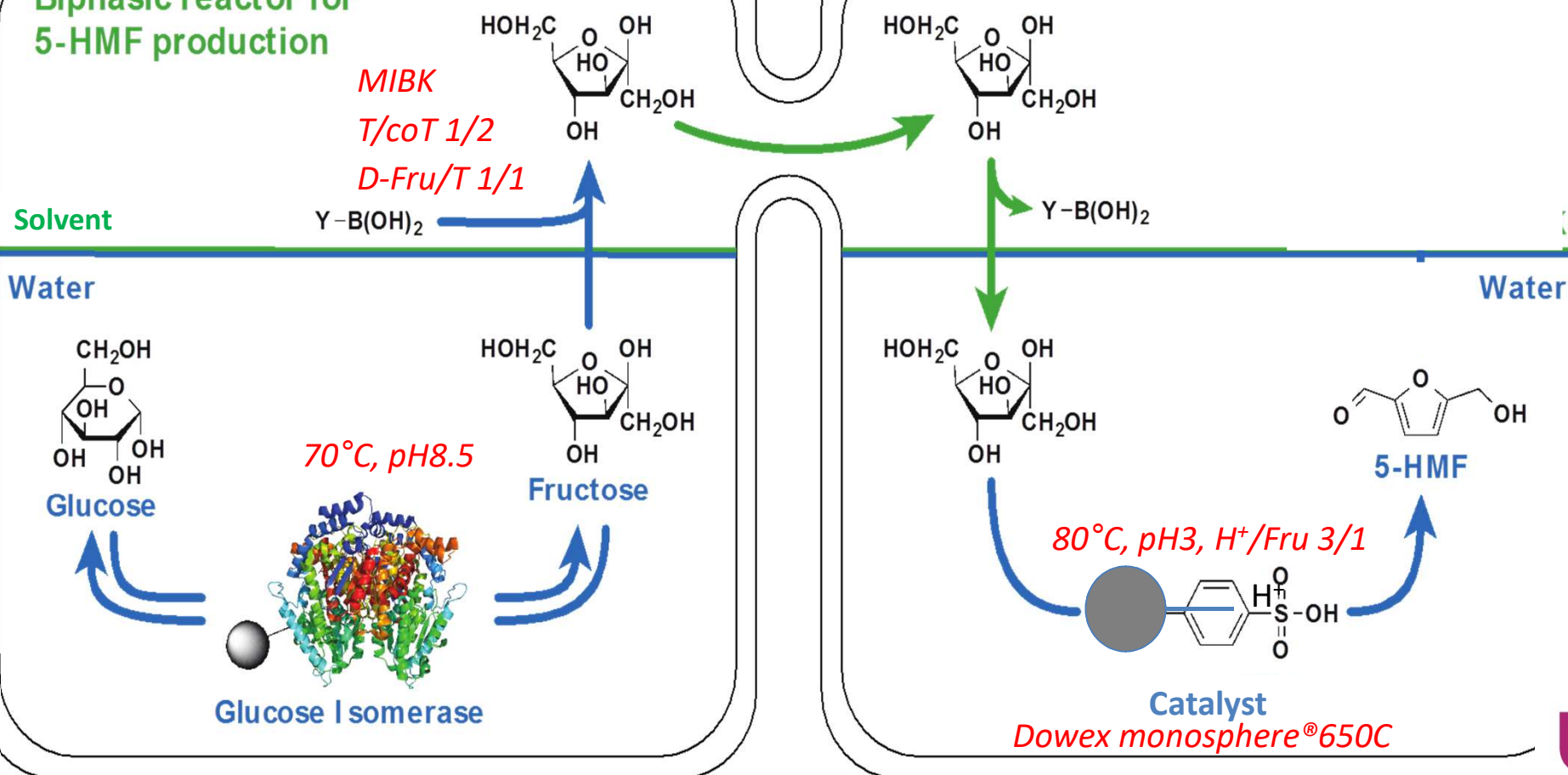
Extraction in MIBK solvent



Fructose Adducts

Glucose Adduct

## Biphasic reactor for 5-HMF production

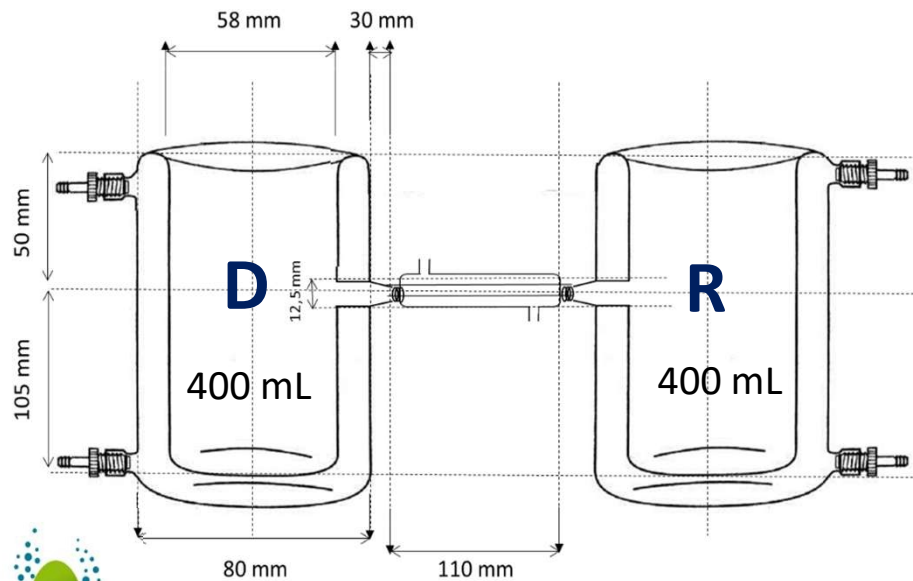


## Transport of fructose:

### Reactor configuration

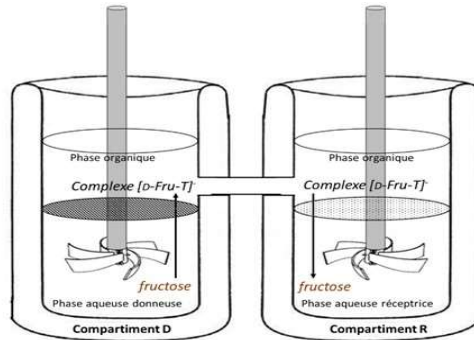
Reactor « H »:

- Bicompartimentalized
- Symmetrical
- Thermostated
- Insertion of stirring blade



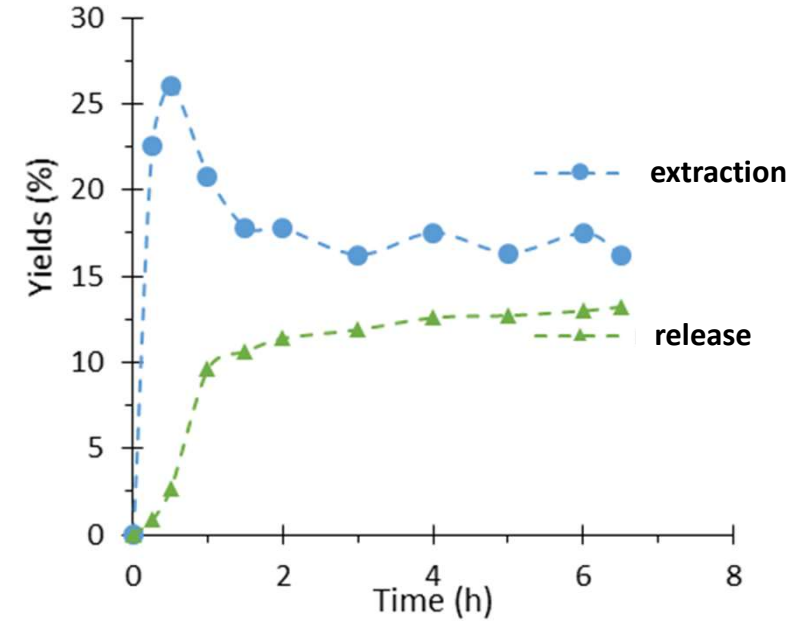
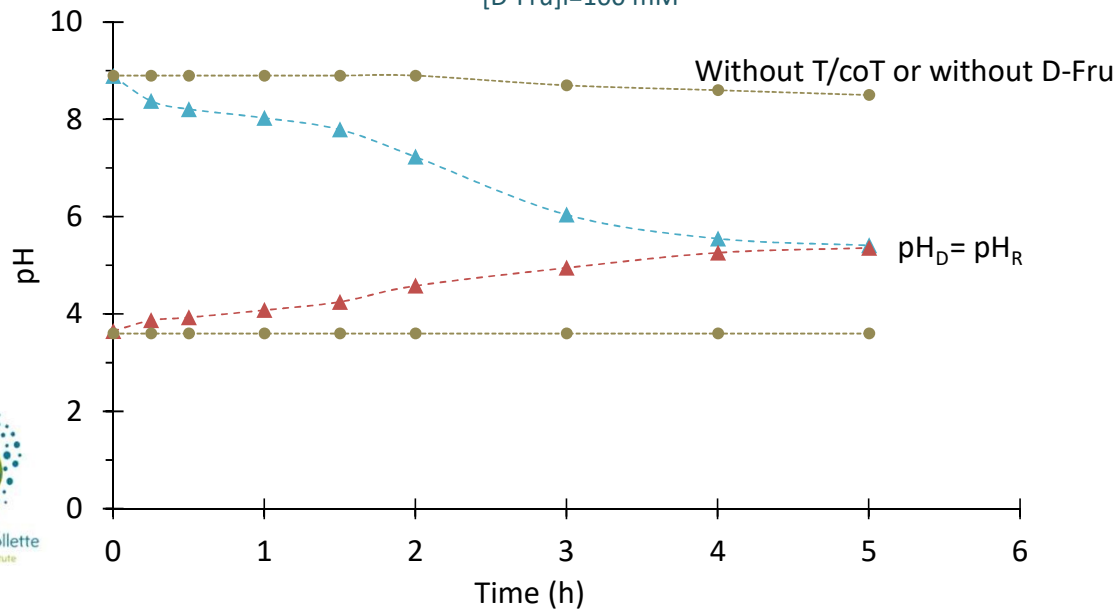
## Transport of fructose:

200 mL MIBK, 70, °C  
 [34-DCPBA]=100 mM  
 [Aliquat336®]=200 mM



Compartment D:  
 V=200 mL  
 Tris-HCl 100 mM  
 pH 8.5  
 [D-Fru]<sub>i</sub>=100 mM

Compartment R:  
 V=200 mL  
 Citrate buffer  
 100 mM pH 3

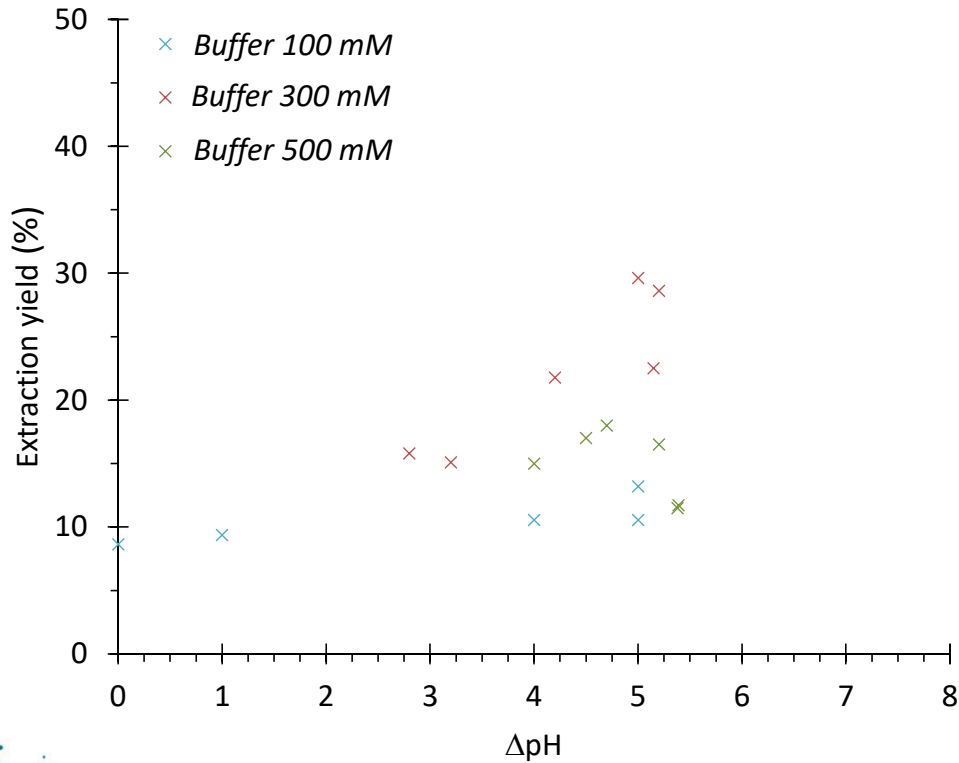


➡ pH unstable during the simultaneous process

➡ Control of pH

## Transport of fructose:

- Influence of  $\Delta\text{pH}$  and ionic strength of the buffer solutions on extraction yields:



$\Delta\text{pH} = \text{pH}_D - \text{pH}_R \approx 4.5 - 5$

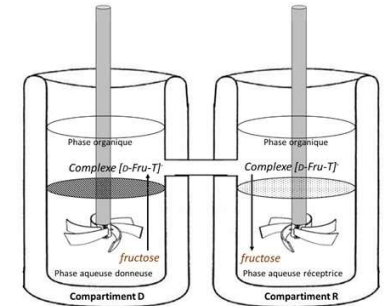


Need to control the pH in continuous during the process



$Y_{\text{extraction}} (500 \text{ mM}) < Y_{\text{extraction}} (300 \text{ mM})$

200 mL MIBK, 70, °C  
 [34-DCPBA]=100 mM  
 [Aliquat336®]=200 mM

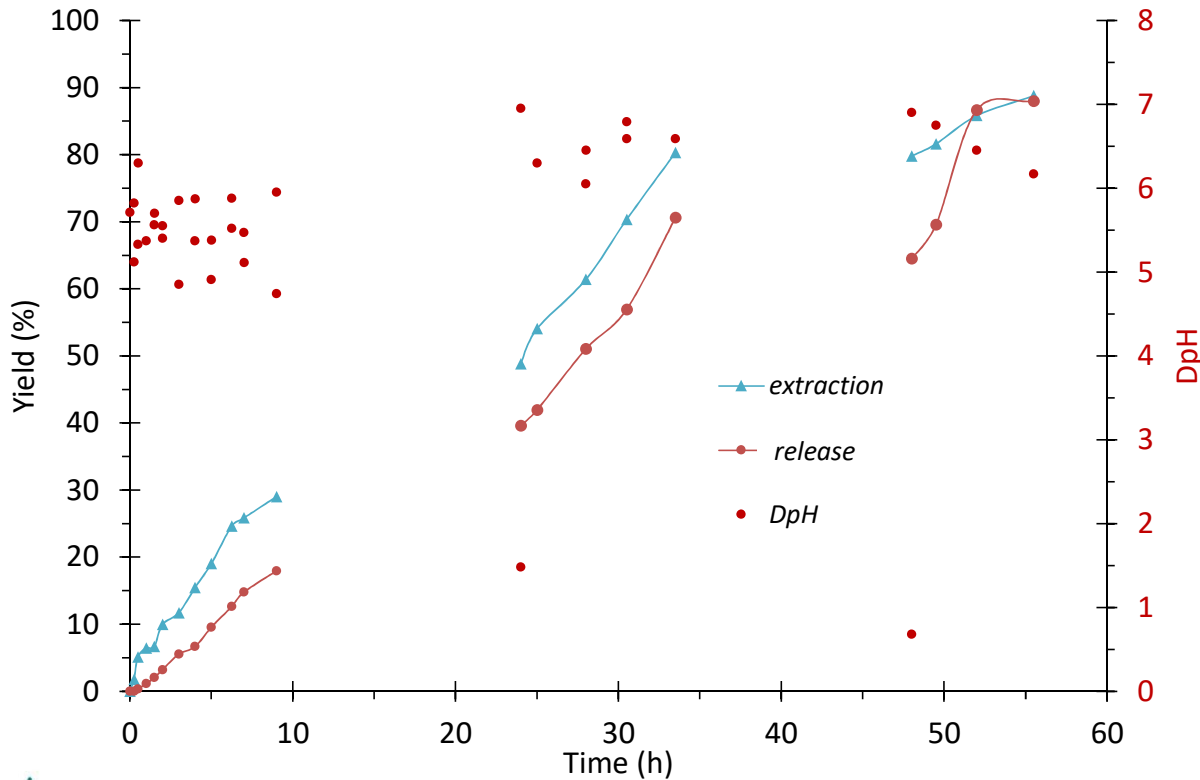


Compartment D:  
 V=200 mL  
 Tris-HCl X mM  
 pH 8.5  
 [D-Fru]<sub>i</sub>=100 mM

Compartment R:  
 V=200 mL  
 Citrate buffer  
 300 mM pH 3

## Transport of fructose:

- Cycles of 34-DCPBA / Aliquat336® :

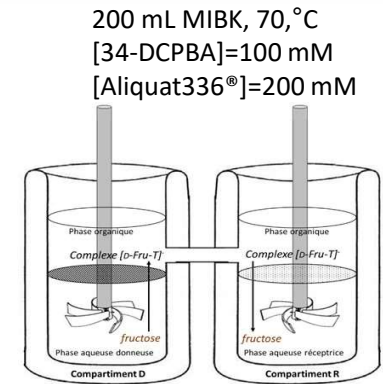


$$Y_{\text{ext}} = 89\%$$

$$Y_{\text{rel}} = 88\%$$

30 regulated hours

$$n_{\text{D-Fru}}/n_{\text{T}} = 1/0.25$$



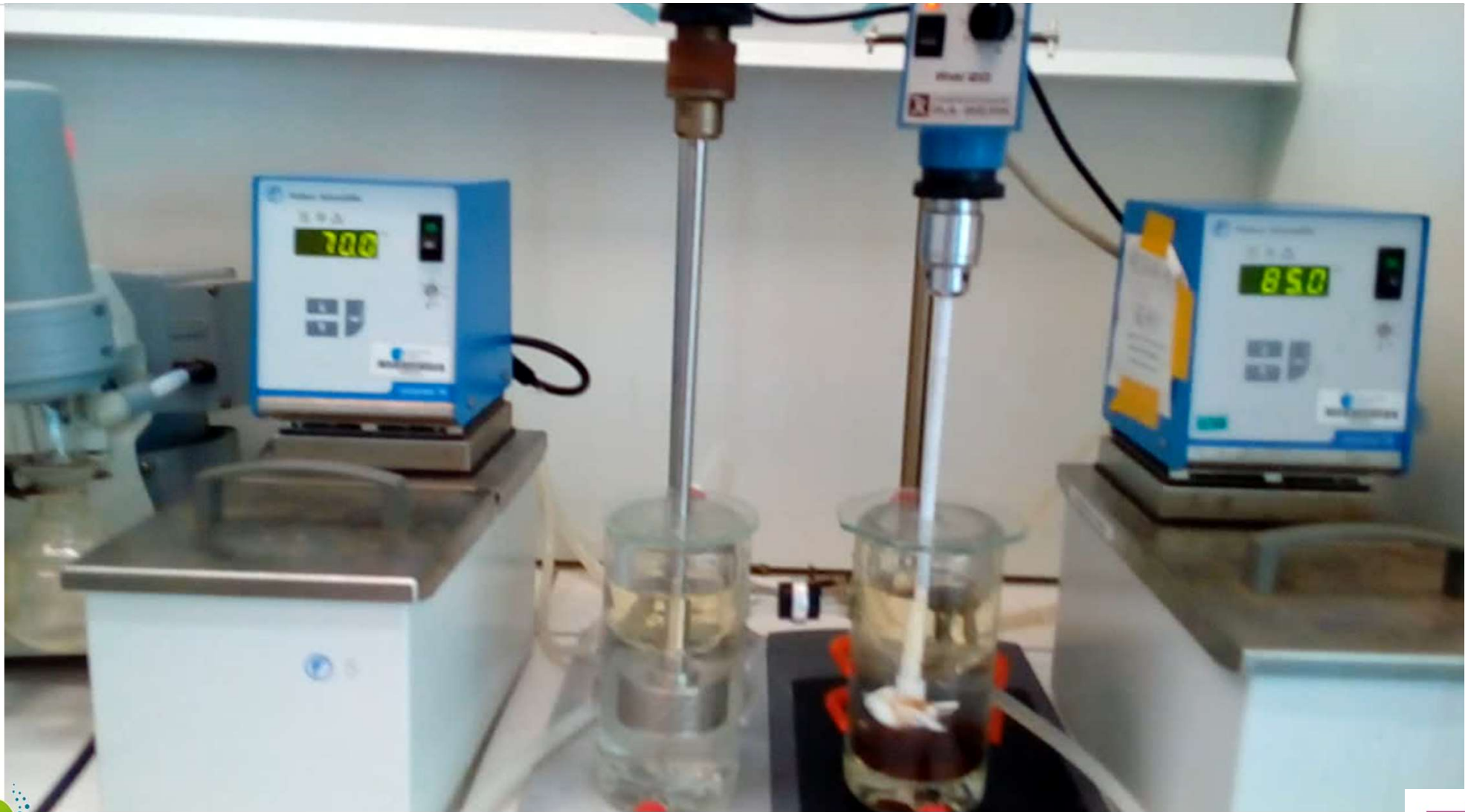
Compartment D:  
V=200 mL  
Tris-HCl 300 mM  
pH 8.5  
[D-Fru]<sub>i</sub>=100 mM

Compartment R:  
V=200 mL  
Citrate buffer  
300 mM pH 3

Amount of fructose extracted > Amount of fructose that could be extracted during 1 extraction cycle

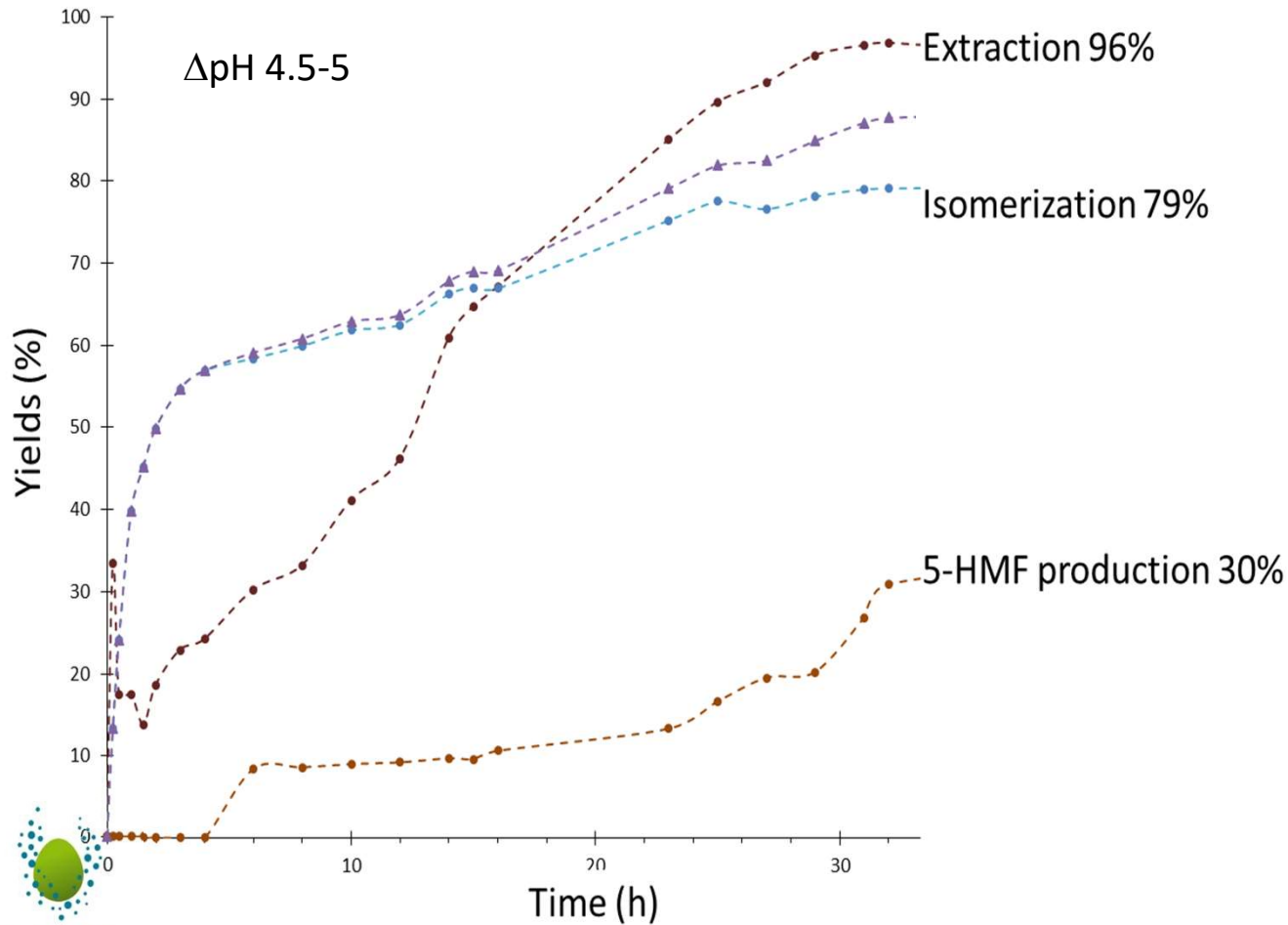
⇒ 34-DCPBA - Aliquat336® pair is recycled during the experiment

⇒ This turnover also highlights the movement of the 34-DCPBA (T) and Aliquat336® (coT) molecules in both directions

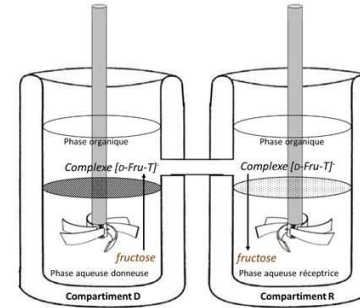


## Hybrid catalysis process implementation

- Implementation of simultaneous catalysis in the « H » reactor:

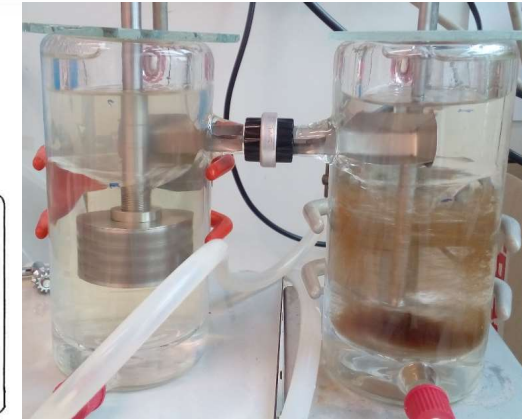


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 [34-DCPBA]=100 mM  
 [Aliquat336®]=200 mM



Compartment D:  
 V=200 mL  
 Tris-HCl 300 mM  
 pH 8.5  
 [D-Fru]<sub>i</sub>=100 mM

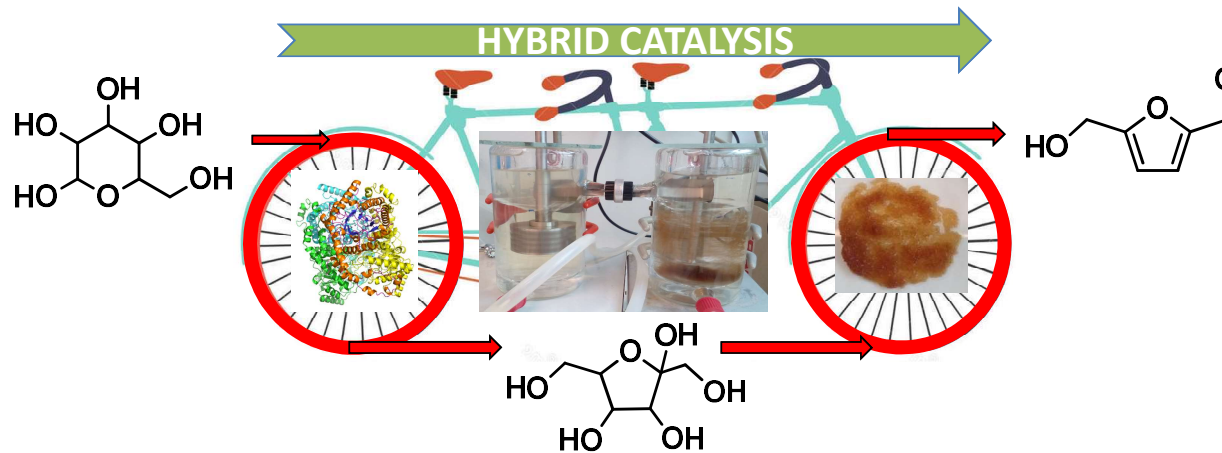
Compartment R:  
 V=200 mL  
 Citrate buffer  
 300 mM pH 3



➔  **$Y_{5\text{-HMF}}=30\%$  at 32h**  
**Conversion\*=90%**  
**Yield<sub>isomerization</sub>=79%** (55% at thermo. equil.)



# Summary



- Simultaneous (bio)catalytical reactions
- Shifting of isomerization thermodynamic equilibrium (25%)

- Study of reactions in sequential approach
- Design of the reactor « H »
- Optimization of D-Fru transport
- Implementation of (bio)catalysis in tandem in the « H » reactor

# Acknowledgements



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# Acknowledgements



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Mickael CAPRON  
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Sébastien PAUL



Sébastien Paul  
Egon HEUSON  
Svetlana HEYTE



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